

# *Francisella tularensis* subsp. *tularensis* Group A.I, United States

## Technical Appendix

### Genome sequencing and assembly

Sequencing of the 13 *F. tularensis* genomes was performed using an Illumina GA IIX instrument (Illumina Inc., San Diego, CA, USA) (100-bp pair-end reads) at the Translational Genomics Research Institute (TGen; Flagstaff, Arizona, USA) and the sequences were assembled using Abyss v1.3.3 (1). To ensure high-quality data, the genome sequences were filtered to minimize uncertain sequence positions in the phylogenetic analysis. After assembly, sequence reads were re-mapped to their corresponding genome sequence using bowtie2 v2.0.0 (2) and subsequent SNP-calling by samtools mpileup (3) and VarScan v2.3.2 (4) using default parameters except p-value=0.9. Using the SNP information, positions for nucleotides that were supported by <90% of the aligned reads were replaced by the nucleotide symbol “N.” After filtering, a multiple genome alignment was calculated by the progressive Mauve algorithm (5) using the 13 filtered genome sequences and two public *F. tularensis* genome sequences. These were strain SCHU S4 (acc. AJ749949.2), representing the A.I clade, and strain WY96-3418 (acc. CP000608.1), representing clade A.II and also serving as an outgroup for the phylogenetic analyses. A second filter was then applied to remove all positions within 30-bp of gaps (“-“) or uncertain positions (“n”) to minimize potential misalignment errors. One SNP found to be incorrectly called due to inaccurate mapping in a repetitive region was manually excluded from the alignment.

### Whole genome phylogeny

Based on the filtered and aligned genome data, a Neighbor-Joining tree was inferred using MEGA5 software 2 (Figure 1 in article main text) with gaps/missing data treated as complete deletions.

The naming of the branch leading up to major groups separated by deeply rooted splits was based on SNP nomenclature: A.I.12, A.I.8, and A.I.3 (Figure 1 in article main text). All short read archives were submitted to SRA (NCBI BioProject Accessions: PRJNA187553, PRJNA187555, PRJNA187556, PRJNA187557, PRJNA187558, PRJNA187559, PRJNA187562, PRJNA187563, PRJNA187564, PRJNA187565, PRJNA187567, PRJNA187568, PRJNA187569).

### **Single Nucleotide Polymorphism (SNP) identification for the development of new canonical SNP assays**

SNPs were identified by mapping paired-end reads to a high quality reference genome (*F. tularensis* SCHU S4, acc. AJ749949.2) (6) using BWA short read alignment software (7) followed by SNP-calling using samtool pileup (3) and VarScan v2.2 (min-var-freq 0.9, min-reads 5 and min-coverage 20) (4). Finally, SNPs defining the three clades A.I.12, A.I.8, and A.I.3 were confirmed using an in-house Perl script based on their presence in a multiple alignment of de novo assembled genome sequences. From this information 16 canonical SNP (canSNP) assays were created as previously described (8).

### **References**

1. Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJ, Birol I. ABySS: a parallel assembler for short read sequence data. *Genome Res.* 2009;19:1117–23. [PubMed](#)  
<http://dx.doi.org/10.1101/gr.089532.108>
2. Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2. *Nat Methods.* 2012;9:357–9. [PubMed](#) <http://dx.doi.org/10.1038/nmeth.1923>
3. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, et al. The Sequence Alignment/Map format and SAMtools. *Bioinformatics.* 2009;25:2078–9. [PubMed](#)  
<http://dx.doi.org/10.1093/bioinformatics/btp352>
4. Koboldt DC, Chen K, Wylie T, Larson DE, McLellan MD, Mardis ER, et al. VarScan: variant detection in massively parallel sequencing of individual and pooled samples. *Bioinformatics.* 2009;25:2283–5. [PubMed](#) <http://dx.doi.org/10.1093/bioinformatics/btp373>

5. Darling AC, Mau B, Blattner FR, Perna NT. Mauve: multiple alignment of conserved genomic sequence with rearrangements. *Genome Res.* 2004;14:1394–403. [PubMed](#)  
<http://dx.doi.org/10.1101/gr.2289704>
6. Larsson P, Oyston PC, Chain P, Chu MC, Duffield M, Fuxelius HH, et al. The complete genome sequence of *Francisella tularensis*, the causative agent of tularemia. *Nat Genet.* 2005;37:153–9. [PubMed](#) <http://dx.doi.org/10.1038/ng1499>
7. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics.* 2009;25:1754–60. [PubMed](#) <http://dx.doi.org/10.1093/bioinformatics/btp324>
8. Birdsell DN, Pearson T, Price EP, Hornstra HM, Nera RD, Stone N, et al. Melt analysis of mismatch amplification mutation assays (Melt-MAMA): a functional study of a cost-effective SNP genotyping assay in bacterial models. *PLoS ONE.* 2012;7:e32866. [PubMed](#)  
<http://dx.doi.org/10.1371/journal.pone.0032866>