# Characteristics of Staphylococcus aureus Infections, Chicago Pediatric Hospital

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Invasive and skin community-associated (CA)-methicillin-resistant *Staphylococcus aureus* isolates from children were matched with invasive CA-methicillin-sensitive *S. aureus* strains during 2000–2004. Isolates were analyzed for presence of Panton-Valentine leukocidin. A USA400 lineage clone (n = 6) and the predominant USA300 lineage clone emerged.

Community-associated methicillin-resistant *Staphylococcus aureus* (CA-MRSA) infections have been increasing in children since the 1990s. Panton-Valentine leukocidin (PVL) has been associated with CA-MRSA strains (1–4).

As CA-MRSA infections have been increasing in previously healthy pediatric patients, we sought to do the following: 1) describe the clonal relatedness of these CA-MRSA isolates by using pulsed-field gel electrophoresis (PFGE), 2) detect the presence of PVL genes among CA-MRSA pediatric isolates causing invasive disease and among isolates causing skin and soft tissue infections (SSTI), 3) determine clinical and epidemiologic differences among patients with invasive disease caused by community-associated methicillin-sensitive *S. aureus* (CA-MSSA) versus those with disease caused by CA-MRSA strains, 4) assess the geographic pattern of infection, and 5) measure the antimicrobial agent susceptibility for CA-MRSA strains.

The institutional review board at Children's Memorial Hospital, a 253-bed, freestanding children's hospital in Chicago, Illinois, approved this study. A CA-MRSA strain was defined as a clinical MRSA isolate recovered from a pediatric patient (<1–18 years of age) who had no established risk factors for MRSA infection (no residence in long-term care facility, no hospitalization except for routine birth, and no permanent indwelling medical devices). For most patients, strains were recovered within 72 hours of admission. Exceptions included patients who had clinical evidence of community-associated disease and whose isolates were obtained after 72 hours of hospitalization. Isolates recovered from normally sterile sites were defined as invasive. We identified patients with *S. aureus* infections retrospectively by reviewing microbiology log books from March 1, 2000, through November 30, 2004.

Demographic and clinical data retrieved included age, sex, race, ZIP code of residence, length of hospitalization, and clinical outcomes. When possible, patients with invasive cases were matched to patients with CA-MRSA SSTI and to those with invasive CA-MSSA infections by age (within 12 months for those <18 months or within 3 years for those >18 months), geographic location of patient residence, or year of infection.

*S. aureus* isolates were identified by standard microbiologic methods. For all *S. aureus* isolates that appeared erythromycin resistant and clindamycin susceptible, antibiotic double disk diffusion assay was performed (5).

Isolates of *S. aureus*, including control strain NCTC 8325 (Bio-Rad, Hercules, CA, USA), were analyzed by PFGE after DNA digestion with *Sma*I. Resulting fragments were separated by using the *Staphylococcus* program 5 (GenePath System, Bio-Rad), and DNA banding patterns were compared (*6*,*7*). PCR was used to detect the PVL genes (8).

Categorical variables were analyzed by  $\chi^2$  analysis. Variables with non-normal distribution were analyzed by Mann-Whitney U test; 2-tailed p value  $\leq 0.05$  was statistically significant (SPSS Inc., version 11.0, Chicago, IL, USA).

From 166 MRSA patient strains noted in the microbiology records, 21 patients with invasive CA-MRSA infection were identified. Three patient isolates were unavailable and were excluded from analysis. Patients with invasive CA-MRSA strains were case-matched with patients with CA-MRSA SSTI (16/18 were matched by age). During the study period,  $\approx$ 31 cases of invasive CA-MRSA disease were identified and 10 cases were able to be retrieved and matched with cases of invasive CA-MRSA (9/10 case-patients were matched by age).

Groups with invasive CA-MRSA and groups with invasive CA-MSSA did not differ significantly regarding sex, initial leukocyte count, duration of fever, or length of hospital stay. Pediatric patients with invasive CA-MRSA infection were more likely to be African American (p =0.01) and were febrile significantly longer than patients with invasive CA-MSSA (p = 0.03). One of the patients with invasive CA-MRSA died (Table 1).

Among the 18 patients with invasive CA-MRSA, 17 patients required surgical drainage, and 1 patient was given extracorporeal membranous oxygenation. Among the 10 patients with invasive CA-MSSA disease, all required surgical intervention, but none died. Of 18

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Table 1. Comparison of clinical characteristics between pediatric patients with invasive CA-MRSA and CA-MSSA*				
Characteristic	Invasive CA-MRSA, n (%)	Invasive CA-MSSA, n (%)	p value	
Race				
Black	16/18 (88)	3/10 (30)	0.01	
Caucasian	0	2/10 (20)	0.01	
Hispanic	1/18 (6)	5/10 (50)	0.01	
Other	0	0	0.01	
Unknown	1/18 (6)	1/10 (10)	0.01	
Days of discordant therapy (CA-MRSA)				
Mean ± SD (range)	2.22 ± 1.76 (0-6)	N/A		
Days febrile, mean ± SD (range)	7.0 ± 4 (0–17)	5.20 ± 10.0 (0-32)	0.03	
Days in hospital, mean ± SD (range)	14.2 ± 7.6 (4–28)	13.8 ± 16.8 (4–60)	NS	
Diagnosis†				
Osteomyelitis, acute	3/18 (38)	1/10 (10)	NS	
Ostyeomyelitis, chronic	2/18 (11)	4/10 (40)	NS	
Bacteremia	5/18 (28)	2/10 (20)	NS	
Endocarditis	0/18 (0)	2/10 (20)	NS	
Pyomyositis	2/18 (11)	0/10 (0)	NS	
Liver abscess	1/18 (6)	0/10 (0)	NS	
Pneumonia + empyema	6/18 (33)	1/10 (10)	NS	
Septic joint	1/18 (6)	2/10 (20)	NS	
Fasciitis	1/18 (6)	0/10 (0)	NS	
Toxic-shock syndrome	1/18 (6)	0/10 (0)	NS	
Other‡	3/18 (38)	0/10 (0)	NS	
Patients with >1 disease manifestation (%)	7/18 (39)	1/10 (10)	NS	
Days of illness before hospitalization, mean ± SD (range)	6.9 ± 11.8 (1–45)	10.9 d ± 15 (2–45)	NS	
Days of positive cultures, mean ± SD (range)	6.94 ± 9.9 (1–45)	4.4 ± 2.98 (1–11)	NS	
Initial leukocyte count§ (thousand/ $\mu$ L), mean ± SD (range)	14.8 ± 13.07 (2.4–57.0)	9.95 ± 4.45 (3.3–17.0)	NS	

\*CA-MRSA, community-associated methicillin-resistant Staphylococcus aureus; MSSA, methicillin-sensitive S. aureus; NS, not significant; SD, standard deviation. Among patients with invasive CA-MRSA infection, 1 died.

+Bonferroni correction for multiple comparisons was applied for comparisons of diagnosis.

‡Other diseases included extensive perineal abscess with fistula, subgaleal hematoma, and a subscapular abscess and prostatitis in the same patient. SOne initial leukocyte count was unavailable in each patient group.

patients with SSTI CA-MRSA disease, 12 were African American, and 2 were Hispanic. The average hospital stay for this group was 2.3 days. Seven children required no hospitalization or hospitalization <24 hours.

A representative PFGE result is depicted in the Figure. Two predominant clones emerged among the local and invasive CA-MRSA isolates. The clone A (n = 6) was identified to be of the USA400 lineage, and the B clone (n =30) was identified to be of the USA300 lineage. No predominant clones emerged from the invasive MSSA isolates; 6 unique clones were identified from 10 isolates (data not shown). No isolates were of the USA300 or 400 lineage. The clinical manifestations of invasive disease in patients with CA-MRSA disease from clone A were pneumonia with empyema, osteomyelitis, bacteremia, and septic arthritis. The clinical manifestations of invasive disease in patients with clone B infection included sepsis, toxicshock syndrome, osteomyelitis, fasciitis, bacteremia, pyomyositis, pneumonia with empyema, deep visceral abscesses, and perirectal abscess with prostatitis; 1 patient in this group died. Clone A was found in a wide geographic distribution around metropolitan Chicago. In contrast,

clone B was located with more frequency within the city limits of Chicago.

Antimicrobial susceptibility patterns among CA-MRSA strains are detailed in Table 2. Two CA-MRSA isolates that were positive by 3-D test belonged to the A0 clone. None of the MRSA isolates that caused SSTI was Dtest positive. The PVL gene was found in all of the CA-MRSA isolates that caused invasive and SSTI disease but in only 1 of 10 of the invasive CA-MSSA isolates (p<0.001).

## Conclusions

PVL genes can be transmitted by means of bacteriophages, which allows them to be transmitted from 1 organism to another (9). When injected intradermally in rabbits, PVL induces necrotic skin lesions (10), and PVL has been described in S. aureus isolates from patients with necrotizing pneumonia, skin infections, and musculoskeletal infecbeen tions. These outbreaks have widespread (1,2,8,11-13). We found a wide range of severity of infection caused by clonally related CA-MRSA, PVL-positive isolates within our community, from superficial skin abscesses to fatal disease.





Figure. Pulsed-field gel electrophoresis (PFGE) results for community-associated methicillin-resistant *Staphylococcus aureus* (MRSA) isolates causing disease. A) Local skin and soft tissue isolates. B) Invasive isolates. \*, PFGE control; \*\*, clinical isolate, hospital-associated MRSA.

Results of PFGE correlate well with results of other molecular typing methods, such as multilocus sequence typing (MLST), which characterizes *S. aureus* species by using sequences of 7 conserved housekeeping genes (7). In accordance with other published studies in the United

States, the USA300 strain was the most frequently isolated among CA-MRSA disease (12) in contrast to the invasive CA-MSSA disease, which had no predominant clonality. We also detected isolates of the USA400 lineage that harbored PVL genes. This lineage was previously described as a cause of severe and fatal CA-MRSA disease in children in the Midwest (14). Previous reports from University of Chicago have described a cluster of 4 cases in which USA400 isolates caused empyema and sepsis syndrome with some features of toxic-shock syndrome (4); 3 children died with necrotizing pneumonia and Waterhouse-Friderichsen syndrome due to a PVL-positive, MLSTidentified type 1 strain (15). In contrast, our patient with fatal toxic-shock syndrome did not have any primary pulmonary pathology and had disease caused by a USA300 lineage strain.

Limitations to our study include its retrospective nature and the limited numbers of patients. The geographic distribution of CA-MRSA isolates within the city likely reflects the geographic distribution of our patient population. Future prospective studies may further elucidate possible epidemiologic risk factors associated with acquiring CA-MRSA invasive infection.

Dr Jaggi is a pediatric infectious disease fellow at Children's Memorial Hospital (Northwestern). Her major research interests have included group A streptococcal infections and pediatric community-associated MRSA infections.

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Table 2. Susceptibility data for CA-MRSA isolates from children*				
Susceptibility to antimicrobial agent†	CA-MRSA isolates causing invasive disease, n (%)	CA-MRSA isolates causing local skin/ soft tissue infections, n (%)	p value	
Resistant to erythromycin	16/18 (88.9)	11/18 (61)	NS	
Apparently susceptible to clindamycin	18/18 (100)	18/18 (100)	NS	
Inducible clindamycin resistance	3/16 (19)	0/10 (0)	NS	
Resistant to ciprofloxacin	2/18 (11)	1/18 (6)	NS	
Resistant to levofloxacin	1/18 (6)	1/18 (6)	NS	
Resistant to tetracycline	1/18 (5)	2/18 (11)	NS	

\*CA-MRSA, community-associated methicillin-resistant Staphylococcus aureus; NS, not significant.

†In addition to the antimicrobial agents listed, all isolates were susceptible to vancomycin, linezolid, and rifampin.

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