unrecognized stages. Many affected travelers may not seek medical attention or may have received a wrong diagnosis. Therefore, surveillance based only on reported cases is likely to underestimate the true incidence of travel-associated *R. africae* infection.

## Paul M. Oostvogel,\* Gerard J. van Doornum,† Russouw Ferreira,‡ Jacqueline Vink,\* Florence Fenollar,§ and Didier Raoult§

\*Medical Center Haaglanden, The Hague, the Netherlands; †Erasmus University Hospital, Rotterdam, the Netherlands; ‡Skukuza, Republic of South Africa; and §Université de la Mediterranée, Marseille, France

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Address for correspondence: Paul M. Oostvogel, Department of Clinical Microbiology, Medical Center Haaglanden, PO Box 432, 2501 CK The Hague, the Netherlands; email: p.oostvogel@mchaaglanden.nl

# Catheter-related Bacteremia and Multidrug-resistant Acinetobacter Iwoffii

To the Editor: Acinetobacter species are ubiquitous in the environment. In recent years, some species, particularly A. baumannii, have emerged as important nosocomial pathogens because of their persistence in the hospital environment and broad antimicrobial drug resistance patterns (1,2). They are often associated with clinical illness including bacteremia, pneumonia, meningitis, peritonitis, endocarditis, and infections of the urinary tract and skin (3). These conditions are more frequently found in immunocompromised patients, in those admitted to intensive care units, or in those who have intravenous catheters, and those who are receiving mechanical ventilation (4,5).

The role of *A. baumannii* in nosocomial infections has been documented (2), but the clinical effect of other *Acinetobacter* species has not been investigated. *A. lwoffii* (formerly *A. calcoaceticus* var. *lwoffii*) is a commensal organism of human skin, oropharynx, and perineum that shows tropism for urinary tract mucosa (6). Few cases of *A. lwoffii* bacteremia have been reported (3,5–7). We report a 4-year (2002–2005) retrospective study of 10 patients with *A. lwoffii* bacteremia admitted to a 600-bed teaching hospital in central Italy.

All 10 patients were immunocompromised; 8 had used an intravascular catheter (peripheral or central) and 2 had used a urinary catheter. Blood cultures of the patients were analyzed with the BacT/ALERT 3D system (bioMérieux, Marcy l'Etoile, France). Isolates were identified as *A. lwoffii* by using the Vitek 2 system and the API 20NE system (both from bioMérieux).

Susceptibilities of 10 A. lwoffii isolates to 18 antimicrobial drugs were determined by the broth microdilution method, according to Clinical and Laboratory Standards Institute (CLSI, formerly NCCLS) guidelines (8). The drugs tested were amikacin, ampicillin-sulbactam, aztreonam, cefepime, cefotaxime, ceftazidime, ceftriaxone, ciprofloxacin, gentamicin, imipenem, levofloxacin, meropenem, ofloxacin, piperacillin, piperacillintazobactam, tetracycline, tobramycin, and trimethoprim-sulfamethoxazole. MIC was defined as the lowest drug concentration that prevented visible bacterial growth. Interpretative criteria for each drug tested were as in CLSI guidelines (8). A. lwoffii resistant to  $\geq 4$  classes of drugs were defined as multidrug-resistant (MDR) isolates.

A. *lwoffii* isolates were genotyped by pulsed-field gel electrophoresis (PFGE) to determine their epidemiologic relatedness. Chromosomal DNA was digested with *SmaI* (9) and analyzed with a CHEF DR II apparatus (Bio-Rad Laboratories, Hercules, CA, USA). PFGE patterns were classified as identical, similar (differed by 1–3 bands), or distinct (differed by  $\geq$ 4 bands) (10).

Among the 10 A. lwoffii isolates, 6 were susceptible to all drugs except cephalosporins (cefepime, cefotaxime, ceftazidime, and ceftriaxone) and aztreonam. The other 4 isolates were MDR: 3 were susceptible only to imipenem (MICs 1-4 µg/mL), meropenem (MICs 1-2 µg/mL), and amikacin (MICs 2-4 µg/mL). The fourth MDR strain was susceptible to imipenem (MIC 2 µg/mL), meropenem (MIC 2 µg/mL), amikacin (MIC 4 µg/mL), and ciprofloxacin (MIC 1 µg/mL). Seven antimicrobial drug resistance profiles were detected (Table).

Macrorestriction analysis of the *A*. *lwoffii* isolates identified 8 distinct PFGE types. Two MDR strains (strains 2 and 3 in the Table), which

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Table. Antimicrobial drug susceptibility and pulsed-field gel electrophoresis (PFGE) patterns of 10 Acinetobacter lwoffii strains, Italy, 2002–2005

|       | Drug†        |         |        |       |       |      |          |         |       |        |       |    |    |    |    |    |    |    |    |             |      |
|-------|--------------|---------|--------|-------|-------|------|----------|---------|-------|--------|-------|----|----|----|----|----|----|----|----|-------------|------|
| No.   | Source*      | AS      | ΡI     | ΡT    | CE    | СТ   | CA       | CF      | AT    | CI     | LE    | OF | GM | ΤМ | AM | тс | IP | MP | ΤS | Antibiotype | PFGE |
| 1     | ICU          | R       | R      | R     | R     | R    | R        | R       | R     | R      | R     | R  | R  | R  | S  | R  | S  | S  | R  | а           | Α    |
| 2     | ICU          | R       | R      | R     | R     | R    | R        | R       | R     | R      | R     | R  | R  | R  | S  | R  | S  | S  | R  | а           | В    |
| 3     | OW           | R       | R      | R     | R     | R    | R        | R       | R     | R      | R     | R  | R  | R  | S  | R  | S  | S  | R  | а           | В    |
| 4     | ICU          | R       | R      | R     | R     | R    | R        | R       | R     | S      | R     | R  | R  | R  | S  | R  | S  | S  | R  | b           | С    |
| 5     | ICU          | S       | S      | R     | S     | S    | R        | R       | R     | R      | R     | R  | S  | S  | S  | S  | S  | S  | S  | с           | D    |
| 6     | ICU          | R       | S      | S     | S     | R    | R        | R       | S     | S      | S     | S  | R  | R  | S  | S  | S  | S  | S  | d           | E    |
| 7     | UW           | S       | S      | S     | R     | R    | R        | R       | R     | S      | S     | S  | S  | S  | S  | R  | S  | S  | S  | е           | F    |
| 8     | ICU          | S       | R      | S     | R     | R    | S        | R       | R     | S      | S     | S  | S  | S  | S  | S  | S  | S  | R  | f           | G    |
| 9     | ICU          | S       | R      | S     | R     | R    | S        | R       | R     | S      | S     | S  | S  | S  | S  | S  | S  | S  | R  | f           | G    |
| 10    | MW           | S       | S      | S     | S     | S    | R        | S       | R     | S      | R     | R  | R  | S  | R  | S  | S  | S  | S  | g           | Н    |
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\*ICU, intensive care unit; OW, orthopedic ward; UW, urologic ward; MW, medical ward.

†AS, ampicillin-sulbactam; PI, piperacillin; PT, piperacillin-tazobactam; CE, cefepime; CT, cefotaxime; CA, ceftazidime; CF, ceftriaxone; AT, aztreonam; CI, ciprofloxacin; LE, levofloxacin; OF, ofloxacin; GM, gentamicin; TM, tobramycin; AM, amikacin; TC, tetracycline; IP, imipenem; MP, meropenem; TS, trimethoprim-sulfamethoxazole; R, resistant; S, susceptible.

were isolated from patients in different wards, and 2 non-MDR strains (strains 8 and 9), which were isolated from patients in the same ward, had similar PFGE patterns and identical resistance phenotypes. These findings suggest nosocomial transmission. Nine of the 10 patients survived after catheter removal or treatment with appropriate antimicrobial drugs. These results confirm that catheterrelated *A. lwoffii* bacteremia in immunocompromised hosts is associated with a low risk for death (4,6).

This study identified *A. lwoffii* MDR strains that cause bacteremia in immunocompromised catheterized patients. Our data are consistent with those of previous reports on the role of catheters as the principal source of *A. lwoffii* infections.

### Luciano Tega,\* Katia Raieta,† Donatella Ottaviani,‡ Gian Luigi Russo,† Giovanni Blanco,\* and Antonio Carraturo\*

\*Ospedale Santa Maria Goretti, Latina, Italy; †Istituto di Scienze dell'Alimentazione, Avellino, Italy; and ‡Istituto Zooprofilattico Sperimentale dell'Umbria e delle Marche, Ancona, Italy

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Address for correspondence: Antonio Carraturo, Laboratorio di Microbiologia, Ospedale Santa Maria Goretti, Azienda Unità Sanitaria Locale di Latina, Via Canova, 04100 Latina, Italy; email: acarraturo@yahoo.com

