cific. The high plasma creatinine level in the newborn sometimes reflects the mother's plasma creatinine level (9). However, kidney function of the mother of the newborn was within normal limits at the time of Caesarean section; plasma creatinine level of 0.7 mg/dL. An elevated plasma creatinine level is observed frequently in premature infants due to immaturity of the kidney tissue and will usually decrease within a few weeks. Oseltamivir was administered with dose adjustment based on the infant's estimated glomerular filtration rate. The recommended dose of oseltamivir for glomerular filtration rate <30 mL/ min/1.73 m² is 2-3 mg/kg/day, based on preliminary data obtained by a National Institutes of Health-funded Collaborative Antiviral Study Group (10). The success of our management strategy for this case suggests early treatment with oseltamivir can prevent severe illness in newborns with perinatal influenza A pandemic (H1N1) 2009 infection.

Acknowledgments

We thank the staff of Ratchaburi Hospital for taking care of the neonate and Petra Hirsch for reviewing the manuscript.

This research was supported by the Center of Excellence in Clinical Virology Fund, Faculty of Medicine, Chulalongkorn University and Hospital; Thai Red Cross Society; Commission on Higher Education, Ministry of Education, Chulalongkorn University; and Chulalongkorn University Centernary Academic Development Project.

Wut Dulyachai, Jarika Makkoch, Pornpimol Rianthavorn, Mutita Changpinyo, Slinporn Prayangprecha, Sunchai Payungporn, Rachod Tantilertcharoen, Pravina Kitikoon, and Yong Poovorawan Author affiliations: Ratchaburi Hospital, Ratchaburi, Thailand (W. Dulyachai, M. Changpinyo); Chulalongkorn University, Bangkok, Thailand (J. Makkoch, P. Rianthavorn, S. Prayangprecha, S. Payungporn, R. Tantilertcharoen, P. Kitikoon, Y. Poovorawan); and King Chulalongkorn Memorial Hospital, Bangkok (P. Rianthavorn)

DOI: 10.3201/eid1602.091733

References

- Centers for Disease Control and Prevention. Swine influenza A (H1N1) infection in two children—southern California, March–April 2009. MMWR Morb Mortal Wkly Rep. 2009;58:400–42.
- Centers for Disease Control and Prevention. Use of influenza A (H1N1) 2009 monovalent vaccine: recommendations of the Advisory Committee on Immunization Practices (ACIP), 2009. MMWR Recomm Rep. 2009;58(RR-10):1–8.
- Schwartz GJ, Brion LP, Spitzer A. The use of plasma creatinine concentration for estimating glomerular filtration rates in infants, children, and adolescents. Pediatr Clin North Am. 1987;34:571–90.
- Rowe T, Abernathy RA, Hu-Primmer J, Thompson WW, Lu X, Lim W, et al. Detection of antibody to avian influenza A (H5N1) virus in human serum by using a combination of serologic assays. J Clin Microbiol. 1999;37:937–43.
- Irving WL, James DK, Stephenson T, Laing P, Jameson C, Oxford JS, et al. Influenza virus infection in the second and third trimesters of pregnancy: a clinical and seroepidemiological study. BJOG. 2000;107:1282–9. DOI: 10.1111/j.1471-0528.2000.tb11621.x
- McGregor JA, Burns JC, Levin MJ, Burlington B, Meiklejohn G. Transplacental passage of influenza A/Bangkok (H3N2) mimicking amniotic fluid infection syndrome. Am J Obstet Gynecol. 1984;149:856–9.
- Yawn DH, Pyeatte JC, Joseph JM, Eichler SL, Garcia-Bunuel R. Transplacental transfer of influenza virus. JAMA. 1971;216:1022–3. DOI: 10.1001/ jama.216.6.1022
- Purtilo DT, Hallgren HM, Yunis EJ. Depressed maternal lymphocyte response to phytohaemagglutinin in human pregnancy. Lancet. 1972;1:769. DOI: 10.1016/ S0140-6736(72)90522-3
- Bueva A, Guignard JP. Renal function in preterm infants. Pediatr Res. 1994;36:572–7.

 Allen U, Blumberg EA, Fischer SA, Green M, Humar A, Ison MG, et al. American Society of Transplantation Infectious Diseases Community of Practice Transplant Infectious Disease Section of the Transplantation Society Guidance on Novel Influenza A/H1N1 [cited 2009 Nov 19]. http://www.transplantation-soc.org/downloads

Address for correspondence: Yong Poovorawan, Center of Excellence in Clinical Virology, Department of Pediatrics, Faculty of Medicine, Chulalongkorn University, Bangkok, 10330 Thailand; email: yong.p@chula.ac.th

Bronchial Casts and Pandemic (H1N1) 2009 Virus Infection

To the Editor: In the late 1990s, triple-reassortant influenza A viruses containing genes from avian, human, and swine influenza viruses emerged and became enzootic in swine herds in North America (1). The first 11 human cases of novel influenza A virus infection were reported to the Centers for Disease Control and Prevention (CDC; Atlanta, GA, USA) from December 2005 through February 2009 (1). In response to those reports, surveillance for human infection with nonsubtypeable influenza A viruses was implemented.

In the spring of 2009, outbreaks of febrile respiratory infections caused by a novel influenza A virus (H1N1) were reported among persons in Mexico, the United States, and Canada (2). Patient specimens were sent to CDC for real-time reverse transcription– PCR (RT-PCR) testing, and from April 15 through May 5, 2009, a total of 642 infections with the virus, now called pandemic (H1N1) 2009 virus, were confirmed. Of those 642 patients, 60% were \leq 18 years of age, indicating that children may be particularly susceptible to pandemic (H1N1) 2009 (2).

Children and adults with preexisting underlying respiratory conditions, such as asthma, are at increased risk for complications from infection with pandemic (H1N1) 2009 virus. One possible complication is plastic bronchitis, a rare respiratory illness characterized by formation of large gelatinous or rigid branching airway casts (3). Plastic bronchitis is a potentially fatal condition induced by bronchial obstruction from mucus accumulation resulting from infection, inflammation, or vascular stasis (4). We report a case of bronchial casts that caused atelectasis of the right lung of a child infected with influenza A pandemic (H1N1) 2009 virus.

A 6-year-old boy with asthma and a 1-day history of fever and cough was referred to a hospital pediatrics department because of dyspnea. Clinical examination at hospital admission found respiratory distress, as shown by tachypnea (respiratory rate 66 breaths/ min) and inspiratory retraction, deficient vesicular sounds over the right lung field, elevated blood levels of immunoglobulin E (1,770 IU/mL) and a reduced number of lymphocytes (483 cells/µL), and radiographic evidence of atelectasis of the right lung and hyperinflation of the left lung without air leakage (Figure, panel A). Pandemic (H1N1) 2009 virus infection was confirmed by real-time RT-PCR, as described (5), of an endotracheal as-

pirate. Real-time PCR ruled out Streptococcus pneumoniae, Haemophilus influenzae, Mycoplasma pneumoniae, Legionella pneumophila, Chlamydophila pneumoniae, S. pyogenes, respiratory syncytial viruses A and B, seasonal influenza viruses A and B, parainfluenza viruses 1-3, rhinovirus, enterovirus, human metapneumovirus, human bocavirus, and adenovirus (6). While the patient was breathing room air, his percutaneously monitored oxygen saturation was 86%; respiratory support by mechanical ventilation was then initiated. Mucus casts were extracted by intratracheal suction (Figure, panel B). The patient was treated with an inhaled bronchodilator, intravenous methylprednisolone (20-60 mg/day for 7 days), and antiviral (oseltamivir) and antimicrobial (ampicillin/sulbactam) drugs.

On hospital day 2, chest radiographs showed that atelectasis of the right lower lobe had partially resolved (Figure, panel C). A histologic examination of casts (May-Giemsa stain; Figure, panel D) indicated a mucoid substance containing a predominantly eosinophilic infiltrate (>90% of cells). The patient's respiratory condition during 11 days of oxygen supplementation gradually improved, and he was discharged on hospital day 18.

Plastic bronchitis is related mainly to respiratory, cyanotic cardiac (post-Fontan), and hematologic (sickle cell anemia) diseases. A diagnosis of plastic bronchitis is determined on the basis of clinical findings (pointing to allergic and asthmatic, cardiac, or idiopathic etiologies) and pathologic findings (inflammatory vs. noninflammatory) on examination of casts (*3*). Inflammatory casts contain fibrin, eosinophils, and Charcot-Leyden crystals; noninflammatory casts contain mucin and exhibit vascular hydrostatic changes. The case presented here was the allergic-inflammatory type of plastic bronchitis.

Various treatments for plastic bronchitis have been described and vary from cast removal by expectoration or by bronchoscopy (7,8). Other interventions involve cast disruption by tissue plasminogen activator or urokinase and prevention of cast formation by use of mucolytic agents, steroids, or anticoagulants. However, evidence remains anecdotal because too few plastic bronchitis patients are available for clinical trials. Details of steroid dosage will need to be clarified for pandemic (H1N1) 2009 virus-infected children with respiratory distress from bronchitis and pneumonia.

In Iran during 1998–2001, avian influenza (H9N2) infection among broiler chickens resulted in 20%–60% mortality rates on affected farms (9). Macroscopic examination of specimens from infected chickens showed extensive hyperemia of the respiratory tract, followed by exudate and casts extending from the tracheal bifurcation to the secondary bronchi. Light microscopy indicated severe necrotizing tracheitis. Pandemic (H1N1) 2009

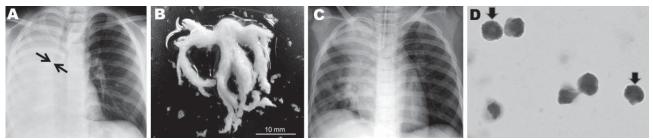


Figure. A) Chest radiograph obtained at hospital admission from a child infected with influenza subtype H1N1 virus. The image shows atelectasis of the right lung and hyperinflation of the left lung; arrows indicate obstruction of the right main bronchus. B) Macroscopic bronchial casts extracted by intratracheal suction. C) Chest radiograph obtained on hospital day 2, indicating partial resolution of atelectasis of the right lower lobe. D) Light micrograph of casts, characterized by predominant eosinophil infiltration (>90% of cells) (May-Giemsa stain, original magnification ×1,000). Arrows indicate typical eosinophil granules. A color version of this figure is available online (www.cdc. gov/EID/content/16/2/344-F.htm).

LETTERS

can produce similar airway cast formation in humans; severe respiratory distress reflects extensive obstruction of the respiratory system.

Healthcare providers should be aware of the possibility of bronchial casts when examining children with influenza (H1N1) infection accompanied by atelectasis. Steroids can be administered early in infection to avoid cast formation, and antiviral drug therapy and respiratory support can be used for influenza (H1N1)–infected children in whom airway casts have developed.

Acknowledgments

We thank Naoko Chiba and Akiko Ono for assistance with manuscript preparation.

This work was supported in part by a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology of Japan (no. 21390306) to T.T.

Maki Hasegawa, Yasuji Inamo, Tatsuo Fuchigami, Koji Hashimoto, Miyuki Morozumi, Kimiko Ubukata, Haruo Watanabe, and Takashi Takahashi

Author affiliations: Nihon University Nerima-Hikarigaoka Hospital, Tokyo, Japan (M. Hasegawa, Y. Inamo, T. Fuchigami, K. Hashimoto); Graduate School of Infection Control Sciences, Kitasato University, Tokyo (M. Morozumi, K. Ubukata, T. Takahashi); and National Institute of Infectious Diseases, Tokyo (H. Watanabe)

DOI: 10.3201/eid1602.091607

References

 Shinde V, Bridges CB, Uyeki TM, Shu B, Balish A, Xu X, et al. Triple-reassortant swine influenza A (H1) in humans in the United States, 2005–2009. N Engl J Med. 2009;360:2616–25. DOI: 10.1056/NEJM oa0903812

- Novel Swine-Origin Influenza A (H1N1) Virus Investigation Team, Dawood FS, Jain S, Finelli L, Shaw MW, Lindstrom S, et al. Emergence of a novel swine-origin influenza A (H1N1) virus in humans. N Engl J Med. 2009;360:2605–15. DOI: 10.1056/NEJMoa0903810
- Madsen P, Shah SA, Rubin BK. Plastic bronchitis: new insights and a classification scheme. Paediatr Respir Rev. 2005;6:292– 300. DOI: 10.1016/j.prrv.2005.09.001
- Kruger J, Shpringer C, Picard E, Kerem E. Thoracic air leakage in the presentation of cast bronchitis. Chest. 2009;136:615–7. DOI: 10.1378/chest.08-0383
- Hasegawa M, Hashimoto K, Morozumi M, Ubukata K, Takahashi T, Inamo Y. Spontaneous pneumomediastinum complicating pneumonia in children infected with 2009 pandemic influenza A(H1N1) v virus. Clin Microbiol Infect. 2009 Oct; [Epub ahead of print]. DOI: 10.1111/ j.1469-0691.2009.03086.x
- Hamano-Hasegawa K, Morozumi M, Nakayama E, Chiba N, Murayama SY, Takayanagi R, et al. Comprehensive detection of causative pathogens using real-time PCR to diagnose pediatric communityacquired pneumonia. J Infect Chemother. 2008;14:424–32. DOI: 10.1007/s10156-008-0648-6
- Noizet O, Leclerc F, Leteurtre S, Brichet A, Pouessel G, Dorkenoo A, et al. Plastic bronchitis mimicking foreign body aspiration that needs a specific diagnostic procedure. Intensive Care Med. 2003;29:329– 31. DOI: 10.1007/s00134-002-1610-1
- Nayar S, Parmar R, Kulkarni S, Cherian KM. Treatment of plastic bronchitis. Ann Thorac Surg. 2007;83:1884–6. DOI: 10.1016/j.athoracsur.2006.12.027
- Nili H, Asasi K. Natural cases and an experimental study of H9N2 avian influenza in commercial broiler chickens of Iran. Avian Pathol. 2002;31:247–52. DOI: 10.1080/03079450220136567

Addresses for correspondence: Yasuji Inamo, Department of General Pediatrics, Nihon University Nerima-Hikarigaoka Hospital, Nihon University School of Medicine, 2-11-1, Hikarigaoka, Nerima-ku, Tokyo, 179-0072, Japan; email: y-inamo@pb3.so-net.ne.jp; or Takashi Takahashi, Laboratory of Infectious Diseases, Graduate School of Infection Control Sciences, Kitasato University, 5-9-1 Shirokane, Minato-ku, Tokyo 108-8641, Japan; email: taka2si@lisci.kitasato-u.ac.jp

The opinions expressed by authors contributing to this journal do not necessarily reflect the opinions of the Centers for Disease Control and Prevention or the institutions with which the authors are affiliated.

Methicillin-Resistant *Staphylococcus aureus* ST398, Italy

To the Editor: It has recently become apparent that livestock can constitute a new methicillin-resistant Staphylococcus aureus (MRSA) reservoir and be a source of a novel and rapidly emerging type of MRSA. These livestock-associated MRSA clones are nontypeable by use of pulsed-field gel electrophoresis with SmaI and belong to sequence type (ST) 398 (1). MRSA ST398 clones account for 20% of all MRSA in the Netherlands (2), but the emergence of such clones has been described worldwide (3). Although ST398 transmission has been reported primarily between animals, persons with occupational exposure to livestock are at higher risk for MRSA carriage than the general population. Even though MRSA ST398 usually causes colonization, several cases of infections of variable clinical relevance, varying from skin and soft tissue infections (4) to endocarditis (5) and pneumonia (6), have been described over the past few years. Most instances of ST398 human carriers have been identified among persons who work at pig farms (7). Data regarding MRSA colonization of dairy farmers are less exhaustive and, to our knowledge, only 1 instance of direct transmission between cattle and humans has been proven. MRSA isolates from cows with subclinical mastitis in 2007 in Hungary were indistinguishable from MRSA isolates from the tonsil swab of a farmer who worked with these animals (8). We report a case of MRSA ST398 invasive disease in a cattle farmer, as well as a case of MRSA ST398 necrotizing fasciitis.

In early April 2008, a 52-year-old man was admitted to an intensive care unit in Manerbio, Italy, because of severe sepsis and a large ulcerative and