LETTERS

during the clinical stage of the disease, and in 1 case, at the preclinical or asymptomatic stage. Our findings suggest that PrP^{sc} is likely to be detected in the saliva of BSE-affected cattle during the clinical stage of disease, after accumulation of PrP^{sc} in the brain. PrP^{sc} was found in the salivary glands of BSE-affected cattle at the terminal stage of infection (*1*). Therefore, once the infectious agent reaches the central nervous system, it may spread centrifugally from the brain to the salivary glands through the autonomic nervous system.

Infectivity of saliva and the presence of PrPsc in saliva have been reported in other ruminants affected with transmissible spongiform encephalopathy. Infectivity of saliva was demonstrated in deer with chronic wasting disease (3) and in scrapieaffected sheep (4); the immunolabeled PrP^{Sc} accumulated in the salivary glands of scrapie-affected sheep (5). A low level of PrP^{sc} was detected in concentrated buccal swab samples of preclinical scrapie-infected sheep by using sPMCA (6,7). These results suggest that small amounts of PrPsc may accumulate in the salivary glands and are then secreted into saliva.

The presence of infectious prions in saliva may explain the facile horizontal transmission of scrapie in sheep (4-6) and chronic wasting disease in deer (4,8). There has been no epidemiologic evidence, however, that saliva, milk, blood, and cerebrospinal fluid from BSE-infected cattle are infectious (9). Nonetheless, the potential risk for BSE transmission by body fluids or excretions from BSE-infected cattle is cannot be ruled out by the current data.

This work was supported by a grantin-aid from the BSE and Other Prion Disease Project of the Ministry of Agriculture, Forestry and Fisheries, Japan.

Hiroyuki Okada, Yuichi Murayama, Noriko Shimozaki, Miyako Yoshioka, Kentaro Masujin, Morikazu Imamura, Yoshifumi Iwamaru, Yuichi Matsuura, Kohtaro Miyazawa, Shigeo Fukuda, Takashi Yokoyama, and Shirou Mohri

Author affiliations: National Agriculture and Food Research Organization, Tsukuba, Japan (H. Okada, Y. Murayama, N. Shimozaki, M. Yoshioka, K. Masujin, M. Imamura, Y. Iwamaru, Y. Matsuura, K. Miyazawa, T. Yokoyama, S. Mohri); and Hokkaido Research Organization, Shintoku, Japan (S. Fukuda)

DOI: http://dx.doi.org/10.3201/eid1812.120528

References

- Murayama Y, Yoshioka M, Masujin K, Okada H, Iwamaru Y, Imamura M, et al. Sulfated dextrans enhance in vitro amplification of bovine spongiform encephalopathy PrP^{Sc} and enable ultrasensitive detection of bovine PrP^{Sc}. PLoS ONE. 2010;5:e13152. http://dx.doi.org/10.1371/ journal.pone.0013152
- Okada H, Iwamaru Y, Imamura M, Masujin K, Matsuura Y, Murayama Y, et al. Detection of disease-associated prion protein in the posterior portion of the small intestine involving the continuous Peyer's patch in cattle orally infected with bovine spongiform encephalopathy agent. Transbound Emerg Dis. 2011;58:333–43. http://dx.doi.org/10.1111/j.1865-1682. 2011.01208.x
- Haley NJ, Seelig DM, Zabel MD, Telling GC, Hoover EA. Detection of CWD prions in urine and saliva of deer by transgenic mouse bioassay. PLoS ONE. 2009;4:e4848. http://dx.doi.org/10.1371/ journal.pone.0004848
- Tamgüney G, Richt JA, Hamir AN, Greenlee JJ, Miller MW, Wolfe LL, et al. Salivary prions in sheep and deer. Prion. 2012;6:52–61. http://dx.doi.org/10.4161/ pri.6.1.16984
- Vascellari M, Nonno R, Mutinelli F, Bigolaro M, Di Bari MA, Melchiotti E, et al. PrPSc in salivary glands of scrapie-affected sheep. J Virol. 2007;81:4872–6. http:// dx.doi.org/10.1128/JVI.02148-06
- Maddison BC, Rees HC, Baker CA, Taema M, Bellworthy SJ, Thorne L, et al. Prions are secreted into the oral cav-

ity in sheep with preclinical scrapie. J Infect Dis. 2010;201:1672–6. http://dx.doi. org/10.1086/652457

- Gough KC, Baker CA, Rees HC, Terry LA, Spiropoulos J, Thorne L, et al. The oral secretion of infectious scrapie prions occurs in preclinical sheep with a range of PRNP genotypes. J Virol. 2012;86:566– 71. http://dx.doi.org/10.1128/JVI.05579-11
- Mathiason CK, Powers JG, Dahmes SJ, Osborn DA, Miller KV, Warren RJ, et al. Infectious prions in the saliva and blood of deer with chronic wasting disease. Science. 2006;314:133–6. http://dx.doi. org/10.1126/science.1132661
- Brown P, Andréoletti O, Bradley R, Budka H, Deslys JP, Groschup M, et al. WHO tables on tissue infectivity distribution in transmissible spongiform encephalopathies. Geneva: World Health Organization; 2010 [cited 2011 Nov 2]. http://www.who. int/bloodproducts/tablestissueinfectivity. pdf

Address for correspondence: Yuichi Murayama, Prion Disease Research Center, National Institute of Animal Health, National Agriculture and Food Research Organization, 3-1-5 Kannondai, Tsukuba, Ibaraki 305-0856, Japan; email: ymura@affrc.go.jp

Reptile- and Amphibianassociated Salmonellosis in Childcare Centers, United States

To the Editor: Salmonella spp. infection represents a major public health problem in the United States; nearly 1.4 million human cases and 600 associated deaths are reported each year (1). Reptile and amphibian exposures might cause >70,000 of these cases annually (2). Furthermore, children are at increased risk of acquiring *Salmonella* spp. and experiencing severe manifestations of disease (3,4). Given the increasing popularity of reptiles and amphibians as pets, reptile- and amphibian-associated salmonellosis is a substantial public health concern (5).

The public has a generally low level of awareness that Salmonella spp. can be acquired from reptiles and amphibians (6); a poll conducted by the US Centers for Disease Control and Prevention (CDC) during 2003 showed that as few as 4 of 49 states require pet stores to provide information about salmonellosis to persons purchasing reptiles (4). A Food and Drug Administration ban, activated in 1975, on the sale of small turtles subsequently prevented an estimated 100,000 cases of salmonellosis in children each year (7). To further reduce the risk of reptile- and amphibian-associated salmonellosis, the CDC has issued recommendations advising that children <5 years of age avoid contact with reptiles and amphibians and that these animals not be kept in childcare centers. The CDC also recommends that all persons wash their hands after handling reptiles and amphibians (8).

We reviewed the regulations as of December 2011 for childcare centers in all US states aimed at preventing reptile- and amphibian-associated salmonellosis (Table). To gather these data, we searched the websites for each state's public health department or the state's equivalent of an early childhood learning agency. When searches on the Internet did not yield the desired information, the appropriate state agencies were contacted by phone or email. In some instances, we corresponded with the designated State Public Health Veterinarian.

Overall, only 50% of states had regulations that required staff and/ or children to wash their hands after touching any animals in childcare centers. Twelve states banned reptiles from childcare centers; 3 of these 12 states also banned amphibians, and these were the only states we found to have banned amphibians from childcare centers. While some states did not allow potentially dangerous or harmful animals in childcare centers, a minority of these states went further to expressly ban reptiles as well (of the 23 states that banned potentially dangerous or harmful animals, 8 states also banned reptiles). One state (Colorado) explicitly banned reptiles, amphibians, and potentially dangerous or harmful animals from childcare centers and also required staff and children in the center to wash their hands after touching animals.

This survey has several limitations. Given the ambiguity in the language used in some regulations and that the language was not standardized between states, we might have misinterpreted some of the documents we reviewed. Furthermore, we might have unintentionally overlooked regulations that were already in place during our investigation, and hence our findings might underestimate the true number of states that have such policies. In some cases, cities and counties have regulations that provide increased protection beyond those implemented at the state level.

In summary, we found great variation between state regulations for childcare centers aimed at reducing transmission of *Salmonella* spp. from reptiles and amphibians to humans. The discrepancy in the regulations of states that banned potentially dangerous or harmful animals from childcare centers but that did not also specifically ban reptiles and amphibians was paradoxical, considering the well-recognized risk that these animals pose for transmitting Salmonella spp. We do not know how many childcare centers across the United States currently house reptiles or amphibians. However, our data suggest that there is room for revision of the regulations in many states which could in turn augment efforts to prevent Salmonella spp. transmission from reptiles and amphibians. We believe that the recommendations issued by the CDC for the prevention of salmonellosis from reptiles and amphibians (4) could serve as a practical guide as state regulations are updated. Our own experience has indicated that greater collaboration between public health organizations and the agencies responsible for setting regulations for childcare centers can be informative and productive. Similarly, state agencies can work with the pet industry and childcare centers to develop approaches that are mutually beneficial.

Although pets provide many benefits to humans, particularly during the early years of life (9), any exposure that children have to animals must pose minimal risk to the children's health. Ultimately, keeping reptiles and amphibians out of childcare centers and requiring that staff and children wash their hands after touching animals offers a simple way to better safeguard the health of children while having a minimal effect on practices of childcare centers.

Acknowledgments

We thank Casey Barton Behravesh, Carina Blackmore, Bryan Cherry, John Dunn, Karl Musgrave, Joni Scheftel, Sally

Table. State regulations for contact between children and animals in childcare centers, United States, 20	11
Description of state regulation	No.(%) states
Bans all animals that show evidence of disease from childcare centers	22 (44)
Bans all potentially dangerous or harmful animals from childcare centers	23 (46)
Bans all reptiles from childcare centers	12 (24)
Bans all amphibians from childcare centers	3 (6)
Requires staff and/or children to wash hands after handling animals	25 (50)

LETTERS

Slavinski, Faye Sorhage, and Carl Williams for their clarification on state and national regulations aimed at reducing the risks of salmonellosis and their advice on conducting this survey. We also thank members and staff of the National Association of State Public Health Veterinarians and the National Resource Center for Health and Safety in Child Care and Early Education for their assistance.

This survey was generously funded by the Mars Foundation and New York Community Trust.

Neil M. Vora, Kristine M. Smith, Catherine C. Machalaba, and William B. Karesh

Author affiliations: Columbia University, New York, New York, USA (N.M. Vora); and EcoHealth Alliance, New York (N.M. Vora, K.M. Smith, C.C. Machalaba, W.B. Karesh)

DOI: http://dx.doi.org/10.3201/eid1812.120784

References

- Mead PS, Slutsker L, Dietz V, McCaig LF, Bresee JS, Shapiro C, et al. Food-related illness and death in the United States. Emerg Infect Dis. 1999;5:607–25. http:// dx.doi.org/10.3201/eid0505.990502
- Mermin J, Hutwagner L, Vugia D, Shallow S, Daily P, Bender J, et al. Reptiles, amphibians, and human *Salmonella* infection: a population-based, case-control study. Clin Infect Dis. 2004;38(Suppl 3):S253– 61. http://dx.doi.org/10.1086/381594
- Mermin J, Hoar B, Angulo FJ. Iguanas and Salmonella Marina infection in children: a reflection of the increasing incidence of reptile-associated salmonellosis in the United States. Pediatrics. 1997;99:399–402. http://dx.doi. org/10.1542/peds.99.3.399
- Centers for Disease Control and Prevention. Reptile-associated salmonellosis selected states, 1998–2002. MMWR Morb Mortal Wkly Rep. 2003;52:1206–9.
- Pickering LK, Marano N, Bocchini JA, Angulo FJ. Exposure to nontraditional pets at home and to animals in public settings: risks to children. Pediatrics. 2008;122:876–86. http://dx.doi. org/10.1542/peds.2008-1942
- Centers for Disease Control and Prevention. Multistate outbreak of human Salmonella Typhimurium infections associated with aquatic frogs—United States, 2009. MMWR Morb Mortal Wkly Rep. 2010;58:1433–6.

- Cohen ML, Potter M, Pollard R, Feldman RA. Turtle-associated salmonellosis in the United States. Effect of Public Health Action, 1970 to 1976. JAMA. 1980;243:1247–9. http://dx.doi. org/10.1001/jama.1980.03300380027016
- Centers for Disease Control and Prevention. Turtle-associated salmonellosis in humans—United States, 2006–2007. MMWR Morb Mortal Wkly Rep. 2007;56:649–52.
- National Association of State Public Health Veterinarians, Inc, Compendium of measures to prevent disease associated with animals in public settings, 2011: National Association of State Public Health Veterinarians, Inc. MMWR Recomm Rep. 2011;60(RR-04):1–24.

Address for correspondence: Neil M. Vora, 460 W 34th St, 17th Floor, New York, NY 10001-2320, USA; email: neilvora@gmail.com

Wild Boars as Hosts of Human-Pathogenic Anaplasma phagocytophilum Variants

To the Editor: Michalik et al. (1) reported a 12% prevalence of Anaplasma phagocytophilum, the causative agent of human granulocytic anaplasmosis and tick-borne fever of ruminants, in wild boars in Poland. A. phagocytophilum has been reported with low prevalence among wild boar in the Czech Republic, Slovenia (2), and Japan (3). In Spain and Mississippi, United States, A. phagocytophilum in wild boars or feral pigs, respectively, has not been reported (4,5). Furthermore, in Slovenia and Poland, the A. phagocytophilum gene sequences found in samples from wild boars were identical to those found in samples from humans and the tick vector

Ixodes ricinus (1). These results suggested, as pointed out by Michalik et al. (1), that wild boar might play a role in the epizootiology of *A. phagocytophilum* by serving as a natural reservoir host, at least in some regions.

To test this hypothesis, we conducted transcriptomics studies to characterize host response to A. phagocytophilum infection in naturally and experimentally infected boars (6,7). The results suggested that boars are susceptible to A. phagocytophilum, but are able to control infection, mainly through activation of innate immune responses and cytoskeleton rearrangement to promote phagocytosis and autophagy. Control of A. phagocytophilum infection in boars might result in infection levels below PCR detection or infection clearance, contributing to the low percentage of infection prevalence detected for this species in most regions.

The low detection levels suggest that boars have a low or no impact as a reservoir host for *A. phagocytophilum*. Even if boars remain persistently infected with *A. phagocytophilum* at low levels by downregulating some adaptive immune genes and delaying the apoptotic death of neutrophils through activation of the Jak-STAT pathway, among other mechanisms (*6*), their role as a source of infection for ticks remains to be demonstrated.

José de la Fuente and Christian Gortazar

Author affiliations: Instituto de Investigación en Recursos Cinegéticos, Ciudad Real, Spain (J. de la Fuente, C. Gortazar); and Oklahoma State University, Stillwater, Oklahoma, USA (J. de la Fuente)

DOI: http://dx.doi.org/10.3201/eid1812.120778

References

 Michalik J, Stańczak J, Cieniuch S, Racewicz M, Sikora B, Dabert M. Wild boars as hosts of human-pathogenic *Anaplasma phagocytophilum* variants. Emerg Infect Dis. 2012;18:998–1001. http://dx.doi. org/10.3201/eid1806.110997