## LETTERS

the severity of the gastrointestinal symptoms (4). Moreover, it can mimic the endoscopic and histopathological features of WD (8).

In this case, the positive PASstaining, the weak positivity of immunochemical staining for *T.* whipplei, and the false-positive results for 1 PCR temporarily delayed diagnosis. False-positive PCR results have been mainly reported when molecular diagnosis for *T. whipplei* was based on 16S rRNA PCR (9). Thus, positivity of a first PCR should be confirmed by using a second PCR with another target (10).

Bacteria responsible for lymph node enlargement are rarely isolated by culture. Molecular methods performed on lymph node biopsy specimens are useful diagnostic tools, but the common single molecular approach using 16S rRNA PCR lacks sensitivity, which delayed diagnosis for this patient (3). To address this issue, simultaneously to performing 16S rRNA PCR, we followed a strategy of systematic qPCR for lymph node specimens that targeted Bartonella spp., F. tularensis, T. whipplei, and Mycobacterium spp. (3). This report confirms the power of this systematic molecular approach, which enabled us to identify a rare bacterial agent scarcely reported for transplant patients.

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### References

- Fenollar F, Laouira S, Lepidi H, Rolain JM, Raoult D. Value of *Tropheryma whipplei* quantitative PCR assay for the diagnosis of Whipple's disease— usefulness of saliva and stool specimens for first line screening. Clin Infect Dis. 2008;47:659– 67. http://dx.doi.org/10.1086/590559
- Lepidi H, Fenollar F, Gerolami R, Mege JL, Bonzi MF, Chappuis M, et al. Whipple's disease: immunospecific and quantitative immunohistochemical study of intestinal biopsy specimens. Hum Pathol. 2003;34:589–96. http://dx.doi. org/10.1016/S0046-8177(03)00126-6
- Angelakis E, Roux V, Raoult D, Rolain JM. Real-time PCR strategy and detection of bacterial agents of lymphadenitis. Eur J Clin Microbiol Infect Dis. 2009;28:1363– 8. http://dx.doi.org/10.1007/s10096-009-0793-6
- Charles P, Lortholary O, Dechartres A, Doustdar F, Viard JP, Lecuit M, et al. *Mycobacterium genavense* infections: a retrospective multicenter study between 1996 and 2007 in France. Medicine. 2011;90:223–30. http://dx.doi. org/10.1097/MD.0b013e318225ab89
- Doggett JS, Strasfeld L. Disseminated Mycobacterium genavense with pulmonary nodules in a kidney transplant recipient: case report and review of the literature. Transpl Infect Dis. 2011;13:38–43. http://dx.doi.org/10.1111/j.1399-3062. 2010.00545.x
- Nurmohamed S, Weenink A, Moeniralam H, Visser C, Bemelman F. Hyperammonemia in generalized *Mycobacterium genavense* infection after renal transplantation. Am J Transplant. 2007;7:722–3. http://dx.doi.org/10.1111/j.1600-6143.2006.01680.x
- de Lastours V, Guillemain R, Mainardi JL, Aubert A, Chevalier P, Lefort A, et al. Early diagnosis of disseminated *Mycobacterium genavense* infection. Emerg Infect Dis. 2008;14:346–7. http://dx.doi. org/10.3201/eid1402.070901

- Albrecht H, Rusch-Gerdes S, Stellbrink HJ, Greten H, Jackle S. Disseminated *Mycobacterium genavense* infection as a cause of pseudo-Whipple's disease and sclerosing cholangitis. Clin Infect Dis. 1997;25:742–3. http://dx.doi. org/10.1086/516941
- Fenollar F, Raoult D. Whipple's disease. Clin Diagn Lab Immunol. 2001;8:1–8.
- Fenollar F, Fournier PE, Robert C, Raoult D. Use of genome selected repeated sequences increases the sensitivity of PCR detection of *Tropheryma whipplei*. J Clin Microbiol. 2004;42:401–3. http://dx.doi. org/10.1128/JCM.42.1.401-403.2004

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# Murine Typhus in Drug Detoxification Facility, Yunnan Province, China, 2010

To the Editor: An outbreak of murine typhus caused by Rickettsia typhi was confirmed among persons attending a 51-acre drug detoxification program 2.5 km from Ruili City in Yunnan Province, People's Republic of China. Ruili City, with an average altitude of 1,381 km, is located in southwestern China near the Myanmar border (Figure). At the time of the outbreak, the detoxification program had 1,264 inpatients and 96 staff members. The facility is divided into sections A (women), B, C, and D. Residents of each section are housed in a 4-story building; each floor contains 9 rooms (2 m<sup>2</sup> per person). During September 4–21, 2010, a total of 76 of the 430 residents of section B were reported with fever of unknown

cause. All patients were men 19–38 years of age who worked in clothing manufacture at the facility and were receiving treatment for drug addiction. Before the outbreak, rats and stray cats were frequently observed in a cafeteria in section B. No persons with similar illness were observed in the other 3 sections.

To investigate the outbreak, gathered information about we demographics; past medical histories; exposures to vectors, such as ticks, mites, fleas, and lice; and symptoms. Patients frequently reported headache, dizziness, diffuse myalgia, high fever (>39°C), and shivers but did not report a rash or eschar. No patients remembered a flea or louse bite, but they frequently reported seeing rats in the area. The Chinese Center for Disease Control and Prevention (China CDC) Institutional Review Board approved the investigation.

Two milliliters of blood was collected from each consenting patient. Separated serum and the remaining blood clots were stored at -70°C and transferred to the Department of Rickettsiology, National Institute of Communicable Disease Control and Prevention, China CDC, for testing. Specimens were tested by indirect immunofluorescence assay (1) to detect specific IgM and IgG against 10 common rickettsiae: Rickettsia prowazekii, *R*. typhi, heilongjiangensis, Orientia R.

tsutsugamushi types Karp and Kato, Coxiella burnetii, Bartonella henselae, and B. quintana, Ehrlichia chaffeensis, and Anaplasma phagocytophilum. Antigens were prepared by placing the rickettsial stains in L929 cells and HL60 or and DH82 cells, respectively; collecting the culture when Gimenez stain or Wright staining showed positive results; ultrasonically crushing the culture; and purifying the bacteria by density ultracentrifugation. Positive control serum was prepared by inoculating rabbits with the above standard rickettsiae strains.

We collected 76 serum samples from patients a median of 4 days (range 1-9 days) after illness onset. Thirty-five (40%) were IgM positive for *R. typhi* (titer >40, maximum titer 160) and 29 (38%) were IgG positive for *R. typhi* (titer  $\geq$  80, maximum titer 320). No samples were positive for the other 8 rickettsial antigens, except for 10 (13%) that had weak reactions for R. prowazakii (titer 40). Twelve convalescent-phase serum samples (median interval between acute and convalescent phases 187 days [range 181–192 days]) were IgG positive for R. typhi (titer >80) and 4 had 4-fold increases in titer; 2 reached titers of 1,280 and 2,560.

DNA was extracted from acutephase samples by using a QIAGEN DNA extraction kit (Hilden, Germany) and tested by real-time PCR that targeted the *groEL* gene of R. prowazekii and R. typhi (2). Twelve (16%) of the 76 samples were positive. To differentiate between R. prowazekii and R. typhi, we used a previously developed nested PCR targeting the groEL gene of R. prowazekii and R. typhi (3) and found the expected 218-bp fragments in 11 patients. BLAST analysis (http://blast. ncbi.nlm.nih.gov/Blast.cgi) showed that these sequences (200 bp) were 100% homologous with that of R. typhi strain Wilmington (GenBank accession no. AF017197).

Initially, patients were treated with antiviral drugs and Chinese herbal medicine for suspected influenza. Subsequently, murine typhus was suspected and doxycycline was administered. All patients recovered fully.

Yunnan Province's subtropical geographic and climate characteristics are advantageous to the vectors of rickettsial diseases, such as murine typhus, scrub typhus, spotted fever, and Q fever (4-6). Three national murine typhus outbreaks involving >10,000 cases each have been reported since 1949, and each involved Yunnan Province (7). In the 1970s, an outbreak of louse-borne typhus occurred in northeastern Yunnan Province (4); since then, louse-borne typhus has been rarely reported. Murine typhus was reported from Baoshan City, east of Ruili City, in 2010. However, the currently reported murine typhus

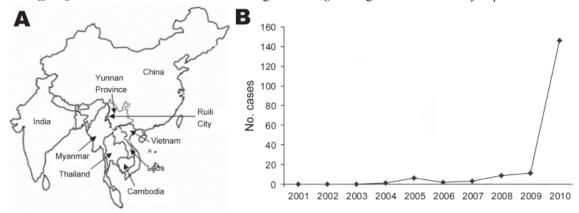


Figure. A) Location of Ruili City, Yunnan Province, People's Republic of China (97°51′–98°02′E, 23°38′–24°14′S; altitude 1,381 m). B) Number of murine typhus cases reported from Ruili City Center for Disease Control and Prevention during 2001–2010.

outbreak in Ruili City near the China-Myanmar border was the largest outbreak in China during the previous decade. None of the 76 patients had rash, a finding similar to that reported in previous outbreaks in Myanmar, Thailand, and other Southeast Asia regions (8-10). In addition to the 76 cases reported here, 70 additional sporadic cases of murine typhus were reported to the Ruili CDC in 2010. We conclude that murine typhus should be considered in cases of unexplained fever with nonspecific clinical manifestations in southern Yunnan Province.

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### References

- Eremeeva ME, Balayeva NM, Raoult D. Serological response of patients suffering from primary and recrudescent typhus: comparison of complement fixation reaction, Weil-Felix test, microimmunofluorescence, and immunoblotting. Clin Diagn Lab Immunol. 1994;1:318–24.
- Wang YY, Liang CW, He J. Establishment of real-time PCR assay for typhus group rickettsia *groEL* genes and clinical case detection [in Chinese]. Dis Surveill. 2011;26:8–11.
- Luan MC, Yu DZ, Tang L, Zhang LJ. Identification of *Orientia tsutsugamushi*, spotted fever group and typhus group rickettsia by duplex and nested PCR methods. Asian Pac J Trop Med. 2008;1:1–8.
- Zhang HL. Research progress on epidemiology of rickettsia disease in Yunnan, China [in Chinese]. Endemic Dis Bull. 2001;16:86–8.
- Zhang LJ, Li XM, Zhang DR, Zhang JS, Di Y, Luan MC, et al. Molecular epidemic survey on co-prevalence of scrub typhus and marine typhus in Yuxi City, Yunnan Province of China. Chin Med J (Engl). 2007;120:1314–8.
- Zhang HL, Yang H, Chao WC. Spotted fever group rickettsia DNA was detected in wild rodent and tick in Dali, Yunnan Province, China [in Chinese]. Chin J Vector Eiol & Control. 2004;15:461–2.
- Fan MY. In historical experience of preventive medicine in new China [in Chinese]. In: Zheng G, editor. Typhus. Beijing: People's Hygiene Publishing House; 1988. p. 145–53.
- Parola P, Miller RS, McDaniel P, Telford SR III, Rolain JM, Wongsrichanalai C, et al. Emerging rickettsioses of the Thai–Myanmar border. Emerg Infect Dis. 2003;9:592–5. http://dx.doi.org/10.3201/ eid0905.020511
- Dumler JS, Taylor JP, Walker DH. Clinical and laboratory features of murine typhus in south Texas, 1980 through 1987. JAMA. 1991;266:1365–70. http://dx.doi. org/10.1001/jama.1991.03470100057033
- Silpapojakul K, Chayakul P, Krisanapan S. Murine typhus in Thailand: clinical features, diagnosis and treatment. Q J Med. 1993;86:43–7.

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# Carpal Tunnel Syndrome with Paracoccidioidomycosis

To the Editor: Paracoccidioidomycosis, a systemic mycosis caused by Paracoccidioides brasiliensis, is endemic to rural areas of Latin America (1). Persons are infected early in life by inhaling the fungus propagules, which reach the lower airway and cause primary complex (2). The most common clinical manifestation of paracoccidioidomycosis, which occurs with the chronic multifocal form, is characterized by pulmonary and extrapulmonary (e.g., skin, central nervous system, osteoarticular system) involvement, which occurs after a prolonged latency period (2). Carpal tunnel syndrome (CTS) is seldom associated with pyogenic agents (3), Mycobacterium tuberculosis (4), or fungal agents (5). Few reports have described paracoccidioidomycosis immunosuppressed patients in (6). We report a rare case of flexor tenosynovitis and severe CTS in the context of reactivated, chronic paracoccidioidomycosis infection.

63-year-old white male А agricultural worker from São Paulo, Brazil, reported insidious and progressive pain, numbness, and tingling in his right hand and fingers, which began in April 2009. His medical history included symmetric polyarthritis of hands, ankles, and knees, which had been diagnosed elsewhere as seronegative rheumatoid arthritis in 2006. At that point, he also had chronic cough; a computed tomographic (CT) scan of the chest showed small nodules and mild interstitial fibrosis, and sputum specimens were negative for fungi or mycobacteria by microscopy. For treatment, he received prednisone, leflunomide, meloxicam, and methotrexate. Hydroxychloroquine was added in March 2010 because

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