# Francisella tularensis Bacteria Associated with Feline Tularemia in the United States

# Marilynn A. Larson, Paul D. Fey, Steven H. Hinrichs, and Peter C. Iwen

Tularemia in the United States was examined by reviewing 106 *Francisella tularensis* isolates, mostly from Nebraska, collected during 1998–2012: 48% of Nebraska cases were cat-associated; 7/8 human cases were caused by subtype A.I. A vaccine is needed to reduce feline-associated tularemia, and cat owners should protect against bites/scratches and limit their pet's outdoor access.

**F**rancisella tularensis, a Tier 1 select agent, is one of the most pathogenic bacteria known and the etiologic agent of the zoonotic disease tularemia (1,2). Although the various subspecies of *F. tularensis* share considerable genomic content (>97% nt identity), they exhibit different degrees of virulence (3). *F. tularensis* subspecies *tularen*sis (also known as type A) and subspecies *holarctica* (also known as type B) infections can be life-threatening if untreated (1). Type A is further subdivided into subtypes A.I and A.II; clade A.I contains strains that are associated with considerably higher death rates among humans than are the other members of this pathogenic species (4). Type B infections occur throughout the Northern Hemisphere, whereas type A infections occur primarily in North America (3).

Fatal and nonfatal cases of tularemia in domestic cats (*Felis catus*) have been reported, as has the transmission of this disease from cats to humans (5–7). Previous serologic surveys from several geographic regions determined that 12% of the domestic cats examined had antibodies to *F. tularensis* (8). Cases of feline-associated tularemia in humans continue to appear in the literature. However, the relative contribution of this source of *F. tularensis* transmission to humans is unknown and may be underrecognized. This study was conducted to examine the relative proportion

Author affiliations: University of Nebraska Medical Center, Omaha, Nebraska, USA

DOI: http://dx.doi.org/10.3201/eid2012.131101

and characteristics of feline-associated *F. tularensis* isolates within the repository at the University of Nebraska Medical Center (UNMC).

# The Study

A review was conducted of the 106 wild-type F. tularensis isolates voluntarily deposited in the UNMC collection during 1998–2012. These isolates were obtained from infected humans, animals, or ticks predominantly residing in Nebraska; however, isolates from several other regions were also included. Wild-type F. tularensis isolates had been transferred to UNMC from other locations, according to requirements of the national Select Agent Program (2). None of these isolates had been solicited from veterinary or environmental reference laboratories. Species identity for locally detected wild-type F. tularensis isolates to which humans had been exposed was confirmed by the Nebraska Public Health Laboratory in Omaha. Viable culture material was manipulated by authorized persons following select agent-approved Biosafety Level 3 criteria. For genotyping, we used the PCR-based differential insertion sequence amplification (DISA) method with the CR10 C+L+S primer set and pulsed-field gel electrophoresis (PFGE) of PmeI-digested Francisella spp. DNA, as previously described (9).

Of the 106 wild-type *F. tularensis* isolates in the repository, 54 (51%) were from humans for whom the source of exposure was unknown or undocumented, 29 (27%) were from cats (21) or from humans (8) with tularemia linked to infected cats, 5 (5%) were from humans with tularemia linked to infected ticks, 1 (1%) was from a human with tularemia linked to an infected rabbit, 16 (15%) were from animals with tularemia, and 1 (1%) was from an unknown host. Of the 29 isolates derived from feline-associated cases, 28 were associated with domestic cats and 1 involved a feral cat (Figure). Eight cases of human tularemia occurred through a cat bite: 7 of these cases involved adults and 1 involved a 6-year-old child. In 2 cases, the person was bitten while taking a rabbit from a cat. None of the 8 humans died.

We also analyzed a subset of the 29 *F. tularensis* isolates obtained from cats or humans bitten by cats. All but 2 of the 29 isolates were available for genotyping. DISA and PFGE results showed that 1 type B and 26 subtype A.I strains were responsible for the 27 cases of tularemia (Figure). Of the 8 cat bite–associated human tularemia cases, 1 was caused by a type B and 7 by subtype A.I strains. None of the feline-associated tularemia cases were caused by a subtype A.II strain.

PFGE demonstrated that the feline-derived *F. tular*ensis A.I strains could be further divided into 4 subpopulations. For strains in 1 of these subpopulations, DISA results showed chromosomal polymorphisms in which a 1.2-kbp or a 1.5-kbp amplicon was produced. This finding is consistent with 2 unique origins for the strains, even though they were obtained from the same geographic area.

Subset analysis of Nebraska cases showed that 48% (24/50) of the wild-type *F. tularensis* isolates were felineassociated, and most of those 24 isolates (71%, 17/24) were associated with tularemia cases that occurred in 1 city in eastern Nebraska. The highest number of tularemia cases in Nebraska was reported in 2012, a year noted for extreme heat and drought conditions with warm weather beginning in early January and lasting through December. No felineassociated tularemia cases in Nebraska were reported during 1998–2004; however, 18 were reported during 2005–2011.

## Conclusions

Two different molecular methods demonstrated that *F. tularensis* subtype A.I was responsible for most of the tularenia cases in cats (96%, 26/27); 1 of the 27 cases was caused by a type B strain. This finding is consistent with subtype A.I strains being found predominantly east of the 100th meridian in the United States, whereas A.II strains appear to be restricted to the west of this meridian

80 85 95 100	Ge Strain I	eographic location	Host (specimen) i	Year solated	Infected source	Genotype	Amplicon size, kbp
	NE-081405	NE	Human (hand)	2005	Domestic cat	A.I	1.5
	NE-120312B	NE	Cat (lymph node)	2012	Domestic cat	A.I	1.5
	NE-120312A	NE	Cat (lymph node)	2012	Domestic cat	A.I	1.5
	SCHU S4	OH	Human (unknown)	1941	Unknown	A.I	1.2
	OK-FelineNC	OK	Cat (unknown)	Unknown	Domestic cat	A.I	1.2
	NE-081205	NE	Human (hand)	2005	Domestic cat	A.I	1.2
	NE-050707	NE	Cat (brain)	2007	Domestic cat	A.I	1.2
	NE-073007	NE	Human (finger)	2007	Domestic cat	A.I	1.2
	NE-092107	NE	Cat (lymph node)	2007	Domestic cat	A.I	1.2
	NE-120707	NE	Cat (liver and spleen)	2007	Domestic cat	A.I	1.2
	NE-060309	NE	Cat (unknown)	2009	Feral cat	A.I	1.2
	NE-050611	NE	Cat (unknown)	2011	Domestic cat	A.I	1.2
	NE-090712	NE	Human (unknown)	2012	Domestic cat	A.I	1.2
	NE-102412	NE	Cat (spleen and lymph node)	2012	Domestic cat	A.I	1.2
	OK-98041035	OK	Cat (unknown)	1998	Domestic cat	A.I	1.2
	OK-00101504	OK	Cat (unknown)	2000	Domestic cat	A.I	1.2
	NC-54558-01	NC	Cat (lung)	2001	Domestic cat	A.I	1.2
	NC-554559-01	NC	Cat (lymph node)	2001	Domestic cat	A.I	1.2
	NE-102105	NE	Cat (blood)	2005	Domestic cat	A.I	1.2
	NE-121905	NE	Human (hand)	2005	Domestic cat	A.I	1.2
A.I	NE-062008	NE	Human (hand)	2008	Domestic cat	A.I	1.5
	NE-071408	NE	Cat (throat)	2008	Domestic cat	A.I	1.5
	NE-083010	NE	Cat (unknown)	2010	Domestic cat	A.I	1.2
	NE-041912	NE	Cat (unknown)	2012	Domestic cat	A.I	1.2
	NE-101912	NE	Cat (salivary gland)	2012	Domestic cat	A.I	1.2
	NE-110607	NE	Human (hand)	2007	Domestic cat	A.I	1.2
	NE-061608	NE	Cat (liver and spleen)	2008	Domestic cat	A.I	1.2
A.II	WY96-3418	WY	Human	1996	Unknown	A.II	1.3
	ATCC 29684	Russia	Vole	Unknown	Unknown	в	2.1
В	NE-061705	NE	Human (hand)	2005	Domestic cat	В	2.1

Figure. Molecular genotyping of *Francisella tularensis* isolates obtained from infected cats or humans bitten by an infected cat, United States, 1998–2012. Genotyping was performed by using pulsed-field gel electrophoresis (PFGE) and the PCR-based differential insertion sequence amplification (DISA) assay. A dendrogram of PFGE patterns obtained with *Pmel*-digested *F. tularensis* isolates is shown on the left; the scale bar at the top indicates distance in relative units. The genotype-specific amplicon lengths obtained with the DISA CR10 C+L+S primer set are shown on the right. Migration profiles of the *Pmel* restriction fragments from *F. tularensis* chromosomal DNA were normalized to *Smal*-digested *Staphylococcus aureus* NCTC 8325 by using BioNumerics software (Applied Maths, Inc., Austin, TX, USA). Cluster analysis was performed by using the Dice correlation coefficient and the unweighted pair group mathematical average (UPGMA) clustering algorithm in the BioNumerics software. The DISA CR10 C+S primer pair identified the subtype A.I strains, whereas the CR10 C+L primer pair differentiated the subtype A.II and type B strains by the size of the amplicon produced. Underlining indicates *F. tularensis* strains SCHU S4 (subtype A.I), WY96–3418 (subtype A.II), and ATCC 29684 (type B), which were included as references. *F. tularensis* isolates NE-062508 and NE-073009 were no longer viable after storage and therefore not genotyped; these pathogens were isolated from domestic cats from eastern Nebraska (NE) in 2008 and 2009, respectively. ATCC, American Type Culture Collection; NC, North Carolina; OH, Ohio; OK, Oklahoma; WY, Wyoming.

## DISPATCHES

(4). More importantly, this study confirms that the highly virulent A.I strains are frequent causes of tularemia in cats and humans, a finding that is in agreement with those in a previous report (4), and that the type B clade can cause disease in these hosts.

Although cats are a well-known vector of F. tularensis, the strikingly high percentage of disease associated with cats in Nebraska was unexpected (48% in Nebraska vs. 27% in the UNMC repository). Of note, the highest number of feline-derived tularemia cases occurred in a city known to have multiple feral cat colonies and during a year with extended warm weather and extreme drought conditions. Warmer temperatures can accelerate the tick life cycle, augmenting the potential for transmission of vector-borne diseases (10,11), and droughts can contribute to impaired host immunity, thereby increasing the likelihood for the transmission of zoonotic pathogens at the limited sources of water (12). Therefore, although the natural reservoir of F. tularensis is unknown, we speculate that these environmental conditions exacerbated the spread of tularemia by increasing the abundance of ticks harboring and transmitting F. tularensis to outdoor animals. Nevertheless, the high relative contribution of cats to disease may be due to underrecognition of an existing problem, a new emerging threat caused by environmental factors, and/or reporting or other biases.

A 2012 systematic review reported that free-ranging domestic cats are probably responsible for the greatest percentage of bird and mammal deaths (13). This information and the findings from the current study support the supposition that although cats are considered incidental hosts for F. tularensis, their inherent predatory behavior increases the likelihood that they will acquire tularemia through the consumption of infected animals and exposure to contaminated ticks, animals, or water harboring this pathogen. A vaccine for tularemia is not available in the United States, and the overall prognosis for infected persons is poor if antibiotic drug treatment is not administered early in the infection process. Therefore, cat owners should protect themselves against potential bites and/or scratches and consider limiting outdoor access for domestic cats. In addition, because tularemia is often initially misdiagnosed (14), confirmatory testing by public health laboratories is important and will provide critical surveillance information regarding F. tularensis in the environment.

The high percentage of feline-associated tularemia cases reported in this study demonstrates that outdoor domestic cats are frequent vectors of F. *tularensis*, particularly in areas where the bacterium is endemic. The cases reported in this study were primarily from urban areas with nearby access to veterinarians and with cat owners/care-takers motivated to treat infected animals; therefore, our findings may not accurately estimate the prevalence of this

pathogen within the feral and rural domestic cat populations. These findings do emphasize the importance of taking appropriate measures to reduce the transmission of feline-associated tularemia to humans and the need for a vaccine that protects against the highly virulent pathogen *F. tularensis*.

#### Acknowledgments

We thank the Nebraska Public Health Laboratory for providing the bacterial isolates.

This study was funded in part by the Department of Pathology and Microbiology at the University of Nebraska Medical Center.

Dr Larson is an assistant professor in the Pathology and Microbiology Department at the University of Nebraska Medical Center in Omaha. Her research interests include the development of diagnostics and therapeutics for pathogen identification and treatment, respectively, as well as determining the molecular mechanisms that allow pathogens to replicate and persist.

#### References

- Sjöstedt A. Tularemia: history, epidemiology, pathogen physiology, and clinical manifestations. Ann N Y Acad Sci. 2007;1105:1–29. http://dx.doi.org/10.1196/annals.1409.009
- Centers for Disease Control and Prevention, Department of Health and Human Services (HHS). Possession, use, and transfer of select agents and toxins; final rule. Fed Regist. 2012;77:61083–115.
- Pechous RD, McCarthy TR, Zahrt TC. Working toward the future: insights into *Francisella tularensis* pathogenesis and vaccine development. Microbiol Mol Biol Rev. 2009;73:684–711. http://dx.doi. org/10.1128/MMBR.00028-09
- Kugeler KJ, Mead PS, Janusz AM, Staples JE, Kubota KA, Chalcraft LG, et al. Molecular epidemiology of *Francisella tularensis* in the United States. Clin Infect Dis. 2009;48:863–70. http://dx.doi.org/10.1086/597261
- Arav-Boger R. Cat-bite tularemia in a seventeen-year-old girl treated with ciprofloxacin. Pediatr Infect Dis J. 2000;19:583–4. http://dx.doi.org/10.1097/00006454-200006000-00024
- Capellan J, Fong IW. Tularemia from a cat bite: case report and review of feline-associated tularemia. Clin Infect Dis. 1993;16:472–5. http://dx.doi.org/10.1093/clind/16.4.472
- Liles WC, Burger RJ. Tularemia from domestic cats. West J Med. 1993;158:619–22.
- Magnarelli L, Levy S, Koski R. Detection of antibodies to *Francisella tularensis* in cats. Res Vet Sci. 2007;82:22–6. http://dx.doi. org/10.1016/j.rvsc.2006.06.003
- Larson MA, Fey PD, Bartling AM, Iwen PC, Dempsey MP, Francesconi SC, et al. *Francisella tularensis* molecular typing using differential insertion sequence amplification. J Clin Microbiol. 2011;49:2786–97. http://dx.doi.org/10.1128/JCM.00033-11
- Tabachnick WJ. Challenges in predicting climate and environmental effects on vector-borne disease episystems in a changing world. J Exp Biol. 2010;213:946–54. http://dx.doi.org/10.1242/jeb.037564
- Estrada-Peña A, Gray JS, Kahl O, Lane RS, Nijhof AM. Research on the ecology of ticks and tick-borne pathogens—methodological principles and caveats. Front Cell Infect Microbiol. 2013;3:29. http://dx.doi.org/10.3389/fcimb.2013.00029

- 12 Altizer S, Dobson A, Hosseini P, Hudson P, Pascual M, Rohani P. Seasonality and the dynamics of infectious diseases. Ecol Lett. 2006;9:467–84. http://dx.doi.org/10.1111/j.1461-0248.2005.00879.x
- Loss SR, Will T, Marra PP. The impact of free-ranging domestic cats on wildlife of the United States. Nat Commun. 2013;4:1396. http://dx.doi.org/10.1038/ncomms2380
- 14. Weber IB, Turabelidze G, Patrick S, Griffith KS, Kugeler KJ, Mead PS. Clinical recognition and management of tularemia in

Missouri: a retrospective records review of 121 cases. Clin Infect Dis. 2012;55:1283–90. http://dx.doi.org/10.1093/cid/cis706

Address for correspondence: Marilynn A. Larson, Department of Pathology and Microbiology, University of Nebraska Medical Center, 985900 Nebraska Medical Center, Omaha, NE 68198-5900, USA; email: malarson@unmc.edu

