

- polymerase chain reaction. [Erratum in: Euro Surveill. 2012;17: pii/20288]. Euro Surveill. 2012;17:20285.
7. van Elden LJ, Nijhuis M, Schipper P, Schuurman R, van Loon AM. Simultaneous detection of influenza viruses A and B using real-time quantitative PCR. *J Clin Microbiol.* 2001;39:196–200. <http://dx.doi.org/10.1128/JCM.39.1.196-200.2001>
 8. Duchamp MB, Casalegno JS, Gillet Y, Frobert E, Bernard E, Escuret V, et al. Pandemic A(H1N1)2009 influenza virus detection by real time RT-PCR: is viral quantification useful? *Clin Microbiol Infect.* 2010;16:317–21. <http://dx.doi.org/10.1111/j.1469-0691.2010.03169.x>
 9. World Health Organization. Health conditions for travellers to Saudi Arabia for the pilgrimage to Mecca (Hajj). *Wkly Epidemiol Rec.* 2013;88:343–7.
 10. Haworth E, Barasheed O, Memish ZA, Rashid H, Booy R. Prevention of influenza at Hajj: applications for mass gatherings. *J R Soc Med.* 2013;106:215–23. <http://dx.doi.org/10.1258/jrsm.2012.120170>

Address for correspondence: Philippe Gautret, Aix Marseille Université, URMITE, 27 Bd Jean Moulin, 13005 Marseille, France; email: philippe.gautret@club-internet.fr

St. Louis Encephalitis Virus Infection in Woman, Peru

To the Editor: St. Louis encephalitis virus (SLEV) is a flavivirus that can asymptotically infect humans or cause clinically apparent disease that manifests with fever, headache, nausea, and vomiting (1). More severe disease with meningoencephalitic involvement may result in stiff neck, alteration in consciousness, gait disturbance, and other focal neurologic deficits. Heightened levels of human disease are often associated with increased abundance of *Culex* spp. mosquitoes and the summer season.

SLEV was first reported in South America in 1960, when it was isolated from pools of *Sabethes bellisarioi* mosquitoes and *Gigantolaelaps mites* in Pará, Brazil (2). SLEV was later recovered from humans in Argentina (1963) (3) and Brazil (1978) (4). Sporadic infections and large outbreaks occurred over ensuing decades, although no isolates in humans have been reported in other South American countries.

Serologic indication of SLEV circulation in Peru was first obtained from hemagglutination-inhibition and neutralization tests of samples collected in 1965 from residents of eastern Peru (5). Later, SLEV was isolated from mosquitoes (6,7), and SLEV antibody was detected in serum specimens from humans by plaque reduction neutralization tests (6). We report the isolation of SLEV from a person in Peru and describe a unique collection method, using oropharyngeal swab specimens, for detecting this virus.

In March 2006, a 50-year-old woman with a 1-day history of fever, sore throat, cough, malaise, myalgia, and headache sought treatment at her local health center in Quistococha, Peru, in the Amazon Basin (3°49'40" N; 73°19'6" E). The woman's recent travel was limited to a 70-km radius from this town. Because influenza was suspected, an oropharyngeal swab specimen was collected as part of an influenza-like illness surveillance project, which had been approved by the US Naval Medical Research Center Institutional Review Board and endorsed by the Peruvian Ministry of Health. No blood specimen was obtained because blood was not collected in this respiratory infection-focused protocol. The swab specimen was inoculated onto Madin-Darby canine kidney cells; no cytopathic effect was observed, and the culture was negative for influenza virus and for other respiratory viruses amenable to culture (e.g., adenovirus and parainfluenza virus).

Nearly 6 years later, as part of a retrospective study of previously negative respiratory specimens, the sample was reevaluated for arboviral infection. Universal transport medium (Copan Diagnostics Inc., Murrieta, CA, USA), containing the swab specimen, was inoculated onto Vero 76 cells; cytopathic effects were revealed on day 7. The cells were harvested, and an indirect immunofluorescence assay was performed by using a panel of mouse polyclonal hyperimmune ascitic fluid specific to a variety of flaviviruses. The initial screening tests indicated reactivity to yellow fever virus, dengue virus, and SLEV. Subsequent immunofluorescence assay analyses using monoclonal antibodies against yellow fever virus and all 4 dengue virus serotypes were negative.

Viral RNA was recovered from the Vero culture supernatant and amplified by conventional reverse transcription PCR/nested PCR with generic flavivirus primers against the nonstructural 5 coding region, which confirmed that the isolate was a flavivirus. Real-time reverse transcription PCR with specific SLEV primers confirmed SLEV.

A total of 10,850 bp, almost the full genome sequence of the virus, were sequenced (GenBank accession no. KF589299), and 10,236 nt from these sequences were compared with other SLEV sequences in GenBank. The strain showed 98.4% similarity with a SLEV strain isolated from a bird in Brazil in 1973, 98.4% similarity with an SLEV strain isolated from mosquitoes in Peru in 1975, and 97.9% similarity with an SLEV strain isolated from mosquitoes in the United States in 2003. The US strain may have been carried by migratory birds from Latin America (8). Phylogenetic analysis by the neighbor-joining method with 1,000 bootstraps replicates identified the isolate as genotype V, subgenotype A, which grouped with the strains obtained in Brazil, Peru, and the United States (Figure) and

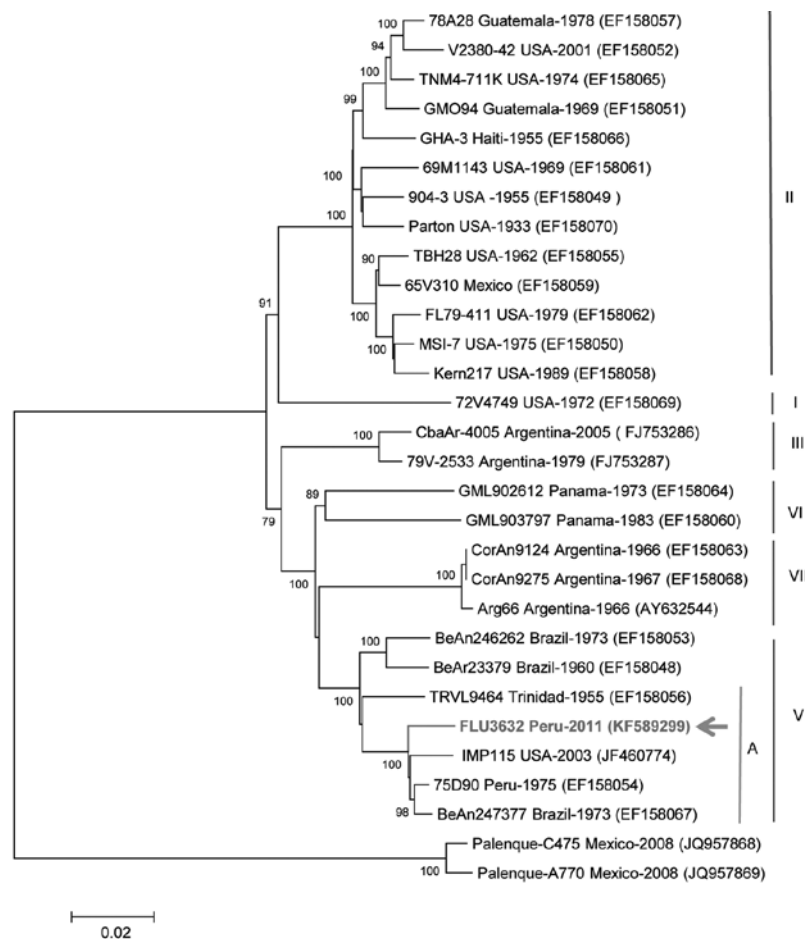


Figure. Phylogenetic analysis from the initially sequenced 10,850-nt region of the St. Louis encephalitis virus (SLEV) genome, isolated from a woman in Peru, 2006. The sequence possessed only 92.8% homology with the NS5 gene region of the sole preexisting SLEV in the laboratory, a genotype II strain similar to TBH28 USA. The Peruvian SLEV sequence described in this case (FLU3632, arrow) groups with Brazil (1975), Peru (1973), and USA (2003) strains, inside the genotype V, subgenotype A. The evolutionary history was inferred by using the neighbor-joining method based on the Kimura 2-parameter model. The entire open reading frame sequence of the Peruvian SLEV isolate was determined in this study by using the Illumina HiSeq 1000 system (Illumina, Inc., San Diego, CA, USA) and assigned GenBank accession no. KF589299. Sequences were analyzed and assembled using the SeqMan Lasergene V.5 and then compared with sequences deposited in GenBank. Multiple sequence alignments were performed by using ClustalX Version 2.0.10 (Conway Institute, University College, Dublin, Ireland; www.clustal.org/) and BioEdit Version 7.0.9.0. (Ibis Biosciences, Carlsbad, CA, USA) Genetic divergence was determined by using MEGA Version 5.02 (www.megasoftware.net/). Scale bar indicates nucleotide substitutions per site.

with an SLEV genotype V strain obtained in Trinidad.

Our findings bolster previous serologic investigations, adding Peru to the South American countries reporting this virus in humans. This discovery is not surprising because *Culex* spp. mosquitoes, the main vectors of SLEV in Brazil and Argentina, have been shown to carry SLEV in Peru (1,7).

Although SLEV disease in humans is usually confirmed by testing blood or cerebrospinal fluid, the isolate described here was recovered from an oropharyngeal swab specimen. The presence of this virus in human respiratory samples was previously suggested by a study in which nasal wash samples from SLEV-infected persons were injected intranasally into mice, which

induced immunity to a subsequent intracerebral challenge with SLEV (9). Although this circumstance is uncommon, other arboviruses have also been obtained from the upper respiratory tract, including dengue virus from nasal and throat swab specimens (10).

Our findings have many implications. First, SLEV may cause human disease in a wider area of South America than was previously known. In addition, SLEV can cause influenza-like illness and may elude identification unless specific assays are used. Finally, the upper respiratory tract may offer a less invasive way of recovering SLEV isolates than by lumbar puncture or even by drawing blood, although further investigations are needed.

Acknowledgments

We thank Juan Sulca and Cristopher Cruz for their laboratory support.

This study was supported by the US Department of Defense Global Emerging Infections Surveillance and Response System, a division of the Armed Forces Health Surveillance Center Work Unit No. 847705.82000.25GB.B0016 and by the National Institutes of Health contract HH-SN272201000040I/HHSN27200004/D04. F.W.S. was supported by the T32 Training grant (AI007536) in Emerging and Re-emerging Infectious Diseases. The sponsors had no role in this study other than providing funding.

The authors have declared that no competing interests exist. The corresponding author had full access to all data in the study and final responsibility for the decision to submit this publication.

Vidal Felices, Julia S. Ampuero, Carolina Guevara, Edna R. Caceda, Jorge Gomez, Felix W. Santiago-Maldonado, Patricia V. Aguilar, and Eric S. Halsey

Author affiliations: US Naval Medical Research Unit No. 6, Lima, Peru (V. Felices, J. Ampuero, C. Guevara, E. Caceda,

E. Halsey); Dirección General de Epidemiología-Ministerio de Salud, Lima (J. Gomez); and University of Texas Medical Branch, Galveston, Texas, USA (F. Santiago-Maldonado, P. Aguilar)

DOI: <http://dx.doi.org/10.3201/eid2004.131735>

References

1. Reisen WK. Epidemiology of St. Louis encephalitis virus. *Adv Virus Res.* 2003;61:139–83. [http://dx.doi.org/10.1016/S0065-3527\(03\)61004-3](http://dx.doi.org/10.1016/S0065-3527(03)61004-3)
2. Causey OR, Shope RE, Theiler M. Isolation of St. Louis encephalitis virus from arthropods in Pará, Brazil. *Am J Trop Med Hyg.* 1964;13:449.
3. Mettler NE, Casals J. Isolation of St. Louis encephalitis virus from man in Argentina. *Acta Virol.* 1971;15:148–54.
4. Pinheiro FP, LeDuc JW, Travassos da Rosa AP, Leite OF. Isolation of St. Louis encephalitis virus from a patient in Belem, Brazil. *Am J Trop Med Hyg.* 1981;30:145–8.
5. Buckley SM, Davis JL III, Madalenoitia J, Flores W, Casals J. Arbovirus neutralization tests with Peruvian sera in Vero cell cultures. *Bull World Health Organ.* 1972;46:451–5.
6. Scherer WF, Madalenoitia J, Menesis O, Acosta M. Study of VE virus and isolation of SLE, EE, group C, and GUAMA group arboviruses in the Amazon region of Peru, 1975. *Bull Pan Am Health Organ.* 1979;13:272–84.
7. Turell MJ, O'Guinn ML, Jones JW, Sardelis MR, Dohm DJ, Watts DM, et al. Isolation of viruses from mosquitoes (Diptera: Culicidae) collected in the Amazon Basin region of Peru. *J Med Entomol.* 2005;42:891–8. [http://dx.doi.org/10.1603/0022-2585\(2005\)042\[0891:IOVFMJ\]2.0.CO;2](http://dx.doi.org/10.1603/0022-2585(2005)042[0891:IOVFMJ]2.0.CO;2)
8. Brault AC, Green E, Maharaj P, Noiwan-gklang W, Armijos V, Wheeler S, et al. Increased diagnostic capacity for the detection of California mosquito-borne viruses. In: Lanzaro GC, editor. *Mosquito control research—annual report 2006.* Oakland (CA): University of California; 2006. p. 11–19.
9. Sulkin SE, Harford CG, Bronfenbrenner J. Immunization of mice by intranasal instillation of nasopharyngeal washings from cases of St. Louis encephalitis. *Exp Biol Med.* 1939;41:327–9. <http://dx.doi.org/10.3181/00379727-41-10662P>
10. Tavakoli NP, Tobin EH, Wong SJ, Dupuis AP II, Glasheen B, Kramer LD, et al. Identification of dengue virus in respiratory specimens from a patient who had recently traveled from a region where dengue virus infection is endemic. *J Clin Microbiol.* 2007;45:1523–7. <http://dx.doi.org/10.1128/JCM.00209-07>

Address for correspondence: Vidal Felices, US Naval Medical Research Unit No. 6, 3230 Lima Place, Washington, DC 20521-3230; email: vidal.felices@med.navy.mil

Whole-Genome Sequencing for Risk Assessment of Long-term Shiga Toxin-producing *Escherichia coli*

To the Editor: Long-term carriage of Shiga toxin-producing *Escherichia coli* (STEC) can greatly affect the social and work lives of infected patients. We describe the use of whole-genome sequencing to assess the risk from long-term STEC carriage in a patient who had been denied surgery because of the infection.

On August 18, 2013, a 64-year-old woman reporting to be a carrier of STEC since March 2013 contacted the University Medical Center Lübeck, Lubeck, Germany, seeking decolonization therapy that had been provided to long-term STEC carriers during the 2011 STEC O104:H4 outbreak (1). STEC had initially been identified in the patient during an episode of watery diarrhea. She currently had gonarthrosis grade III, indicating the need for a total knee endoprosthesis; however, the responsible orthopedic department had denied surgery because of the potential risk for development of STEC-associated hemolytic uremic syndrome (HUS) caused by the perioperative use of antimicrobial drug prophylaxis. The patient was also rejected for surgery at another orthopedic clinic. Because of this STEC-associated restriction, the patient requested decolonization therapy.

Before responding to the request, we asked the patient to provide a fecal sample for STEC strain typing. A sample provided on August 22, 2013, was confirmed positive for STEC by culturing an STEC strain on MacConkey agar (bioMérieux, Marcy l'Etoile, France) that did not grow on selective agar (CHROMagar STEC, Mast Diagnostika, Reinhold, Germany) optimized for the detection of classical enterohemorrhagic *E. coli* strains. Total DNA was extracted from the isolate, and a sequencing library was generated by using the Nextera XT Sample Preparation Kit (Illumina, San Diego, CA, USA). Sequencing was performed (MiSeq Benchtop Sequencer, Illumina) in 2 batches of paired 250-bp sequencing runs. Sequencing reads were further analyzed by using the CLC Genomics Workbench software package (CLC bio, Aarhus, Denmark). De novo assembly resulted in 120 contigs with an average length of 44,331 bp (N50 = 126,317 bp). A predefined dataset of 2,456 sequences was aligned with the generated contigs in a single step by using BLAST (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) to determine the alleles and subtypes of genes usually used for *E. coli* and STEC strain typing and for seropathotype detection.

Presence of a Shiga toxin subtype 1a with >99.9% and 100% identity to the *stx1aA* and *stx1aB* subunit genes, respectively (GenBank accession no M19473.1), was confirmed. The STEC strain carried genes with high homology to the O91 antigen-encoding operon (GenBank accession no. AY035396.1) and the H14-flagellin gene (GenBank accession no. AY249998.1). This observation was confirmed by a 100% sequence identity of a 643-bp fragment of the *gnd* gene of the sequenced strain with that of a *gnd* reference sequence of STEC O91:H14 (www.corefacility.ca/ecoli_typer). These 2 sequences are different from the *gnd* sequence of reference strain STEC O91:H21. In vitro multilocus sequence typing (2) identified