

parallel with similar efforts in Europe, strategies need to be developed to protect commercial and pet rabbits.

Tracking the spread of RHDV2 in Australia, in competition with existing field strains, highlights the value of Australia's rabbits and their diseases as a model system for emerging infectious diseases. The point releases of both myxoma virus and RHDV into large naive host populations represent a grand experiment in disease emergence and evolution (10), which provides a unique opportunity to study the virulence evolution of emerging pathogens as well as their complex interactions with each other. It is notable that since the release of RHDV in Australia in 1995, strains of 1 viral lineage dominate the viral population nationwide despite hundreds of deliberate releases of the original virus strain for local rabbit control, which strongly suggests it has a major selective advantage (7). That RHDV2 appeared in a wild rabbit is therefore remarkable, particularly because Australian field strains were spreading simultaneously in the same area. Comparing the epidemiology of this strain in Australia to the epidemiology of its well-documented spread in Europe will provide valuable insights into RHDV epidemiology relevant to both continents.

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References

1. Le Gall-Reculé G, Zwingelstein F, Boucher S, Le Normand B, Plassiart G, Portejoie Y, et al. Detection of a new variant of rabbit haemorrhagic disease virus in France. *Vet Rec.* 2011;168:137–8. <http://dx.doi.org/10.1136/vr.d697>
2. Dalton KP, Nieceza I, Balseiro A, Mugerza MA, Rosell JM, Casais R, et al. Variant rabbit hemorrhagic disease virus in young rabbits, Spain. *Emerg Infect Dis.* 2012;18:2009–12. <http://dx.doi.org/10.3201/eid1812.120341>
3. Le Gall-Reculé G, Lavazza A, Marchandeu S, Bertagnoli S, Zwingelstein F, Cavadini P, et al. Emergence of a new lagovirus related to rabbit haemorrhagic disease virus. *Vet Res.* 2013;44:1–13. <http://dx.doi.org/10.1186/1297-9716-44-81>
4. Abrantes J, Lopes AM, Dalton KP, Melo P, Correia JJ, Ramada M, et al. New variant of rabbit hemorrhagic disease virus, Portugal, 2012–2013. *Emerg Infect Dis.* 2013;19:1900–2. <http://dx.doi.org/10.3201/eid1911.130908>
5. Lopes AM, Correia J, Abrantes J, Melo P, Ramada M, Magalhaes MJ, et al. Is the new variant RHDV replacing genotype 1 in Portuguese wild rabbit populations? *Viruses.* 2015;7:27–36. <http://dx.doi.org/10.3390/v7010027>
6. Elsworth P, Cooke BD, Kovaliski J, Sinclair R, Holmes EC, Strive T. Increased virulence of rabbit haemorrhagic disease virus associated with genetic resistance in wild Australian rabbits (*Oryctolagus cuniculus*). *Virology.* 2014;464–5:415–23. <http://dx.doi.org/10.1016/j.virol.2014.06.037>
7. Eden JS, Kovaliski J, Duckworth JA, Swain G, Mahar JE, Strive T, et al. Comparative phylodynamics of rabbit haemorrhagic disease virus (RHDV) in Australia and New Zealand. *J Virol.* 2015;89:9548–58. <http://dx.doi.org/10.1128/JVI.011100-15>
8. Lopes AM, Dalton KP, Magalhaes MJ, Parra F, Esteves PJ, Holmes EC, et al. Full genomic analysis of new variant rabbit hemorrhagic disease virus revealed multiple recombination events. *J Gen Virol.* 2015;96:1309–19. <http://dx.doi.org/10.1099/vir.0.000070>
9. Almeida T, Lopes AM, Magalhães MJ, Neves F, Pinheiro A, Gonçalves D, et al. Tracking the evolution of the G1/RHDVb recombinant strains introduced from the Iberian Peninsula to the Azores islands, Portugal. *Infect Genet Evol.* 2015;34:307–13. <http://dx.doi.org/10.1016/j.meegid.2015.07.010>
10. Di Giallonardo F, Holmes EC. Viral biocontrol: grand experiments in disease emergence and evolution. *Trends Microbiol.* 2015;23:83–90. <http://dx.doi.org/10.1016/j.tim.2014.10.004>

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Characteristics of Traveler with Middle East Respiratory Syndrome, China, 2015

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To the Editor: A traveler returning from the Middle East initiated an outbreak of Middle East respiratory syndrome (MERS) in South Korea in 2015, which resulted in 186 cases and 36 deaths (1–3). We report a case of

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MERS in a 43-year-old man from South Korea who acquired this disease during this outbreak (online Technical Appendix Figure 1, panel A, <http://wwwnc.cdc.gov/EID/article/21/12/15-1232-Techapp1.pdf>) (4).

The National Health and Family Planning Commission of China determined that collection of data for this patient was part of a public health investigation of an emerging outbreak. Therefore, informed consent was not required. This study was approved by the ethical committee of the First Affiliated Hospital of Guangzhou Medical University.

The patient had been receiving thiamazole for 7 years for hyperthyroidism. He had contact with the index case-patient during the outbreak in South Korea on May 16, 2015. On May 25, the patient traveled to Hong Kong and then to Huizhou, China. He was hospitalized in China on May 28 (day 7 of illness). At admission, he had a high fever (temperature 39.5°C) and a dry cough. Chest radiography on day 7 showed mild bilateral ground glass opacities in the lower lung (online Technical Appendix Figure 1, panel B).

The patient was given oseltamivir (150 mg, 2×/day for 2 days) until identified as being infected with Middle East respiratory syndrome coronavirus (MERS-CoV) on day 8 by real-time reverse transcription PCR. He was given ribavirin (2.0 mg on day 8; 0.6 mg 3×/d on days 9–16; and 0.6 mg 2×/d on days 17–19) and 135 µg of peginterferon α-2a by intravenous injection on day 8 (online Technical Appendix Table 2). Thrombocytopenia and a decrease in the hemoglobin level developed, which might have been related to use of ribavirin (online Technical Appendix Table 1).

Chest radiography on June 1 (day 11) showed increased bilateral consolidation of the patient's lower lung (online Technical Appendix Figure 1, panel C). He was given intravenous immunoglobulin, antimicrobial drugs, and thymosin α1. His body temperature returned to normal on day 14 (online Technical Appendix Figure 2). Chest radiography on day 35 showed resolution of bilateral lung infiltrations (online Technical Appendix Figure 1, panel D). He was discharged on day 36.

Viral RNA was detected in sputum and fecal specimens up to day 26 of illness. Virus load in sputum specimens collected on days 11–15 were lower than in specimens obtained on days 16–18 (online Technical Appendix Figure 3, panel A). Swab samples collected on days 13 and 15 from the patient's palm, mobile telephone, blanket, and bed railings, and from his hospital room floor were negative for viral RNA.

Concentrations of proinflammatory cytokines and chemokines (interferon-α, interferon-inducible protein 10, monocyte chemoattractant protein-1, interleukin 6 [IL-6], IL-10, tumor necrosis factor-α, IL-8, macrophage inflammatory protein-α [MIP-1α], MIP-1β, and IL-1β) were determined for serial serum samples. Interferon-α, interferon-inducible protein 10, monokine induced by interferon-γ, IL-6, monocyte

chemoattractant protein-1, and IL-8 were detected on day 11 of illness but levels decreased as the patient clinically improved (online Technical Appendix Figure 3, panel B).

The peginterferon α2 the patient was given on day 8 might have influenced his plasma interferon-α levels (6). However, a previous study also showed increased levels of interferon-α in a patient who survived MERS-CoV infection but not in a person who died of MERS (7). Although MERS-CoV evades induction of innate immune responses by cell types, the virus elicits interferon responses in plasmacytoid dendritic cells *in vitro* (8). Levels of tumor necrosis factor-α, MIP-1α, MIP-1β, IL-10, and IL-1β did not increase in any of these specimens.

Peripheral blood mononuclear cells (PBMCs) obtained on day 24 of illness showed a strong specific T-cell response against MERS-CoV spike protein but not against severe acute respiratory syndrome coronavirus (SARS-CoV) spike protein (online Technical Appendix Figure 3, panel C). PBMCs from persons who were infected with SARS-CoV in 2003, as well as healthy persons, showed low-level T-cell responses against MERS-CoV spike protein, although some persons with a history of SARS still had detectable responses to SARS-CoV spike protein. It was reported that T-cell responses to SARS-CoV were directed against spike and nucleocapsid proteins (9). We did not have sufficient PBMCs to test T-cell responses against nucleocapsid protein.

Results for MERS-CoV antibody were negative at day 11 of illness by MERS-CoV spike pseudotype assay (MERS-S ppNT), microneutralization, 50% plaque reduction neutralization test (PRNT₅₀), and S1 ELISA (EUROIMMUN AG, Lübeck, Germany). The patient showed seroconversion by day 14. MERS-S ppNT and PRNT₅₀ provided earlier evidence of seroconversion (day 15) and higher antibody titers than the microneutralization, (day 18) (online Technical Appendix Figure 3, panel D). Potent T-cell responses were elicited to MERS-CoV spike protein. These responses did not show cross-reactivity with SARS-CoV spike protein.

The MERS-S ppNT, which does not require Biosafety Level 3 containment, had sensitivity equivalent with that of PRNT₅₀, which requires containment. Thus, MERS-S ppNT is a sensitive and specific assay for detecting neutralizing antibody against MERS-CoV. The sensitivity and specificity of this assay have been well-documented with serum samples from dromedary camels and other animals (10).

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References

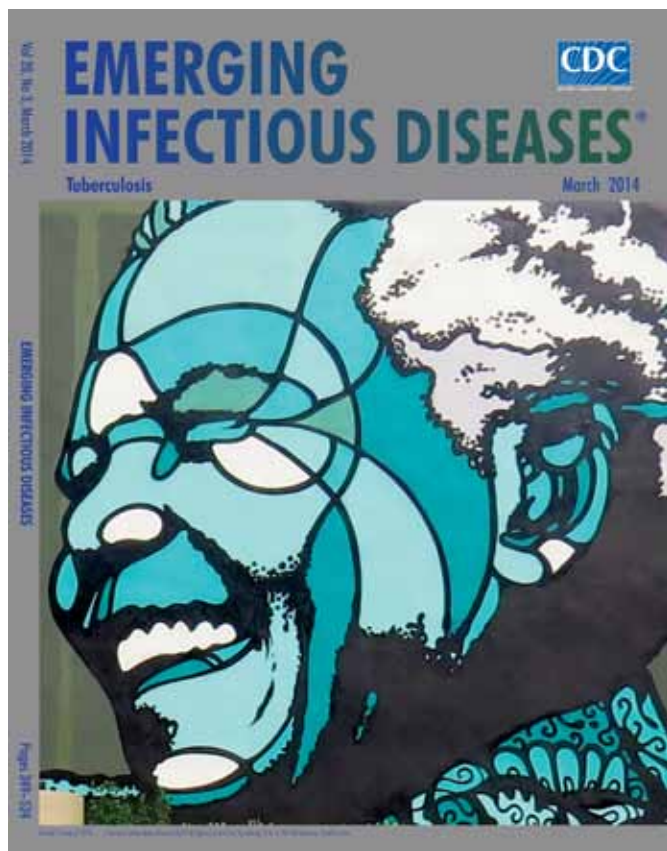
1. Cowling BJ, Park M, Fang VJ, Wu P, Leung GM, Wu JT. Preliminary epidemiological assessment of MERS-CoV outbreak in South Korea, May to June 2015. *Euro Surveill.* 2015;20:pii: 21163.
2. Park HY, Lee EJ, Ryu YW, Kim Y, Kim H, Lee H, et al. Epidemiological investigation of MERS-CoV spread in a single hospital in South Korea, May to June 2015. *Euro Surveill.* 2015; 20:pii: 21169.
3. World Health Organization. Middle East respiratory syndrome coronaviruses (MERS-CoV) [cited 2015 Sep 5]. <http://www.who.int/emergencies/mers-cov/en/>
4. Wu J, Yi L, Zou L, Zhong H, Liang L, Song T, et al. Imported case of MERS-CoV infection identified in China, May 2015: detection and lesson learned. *Euro Surveill.* 2015;20:pii:21158.
5. Memish ZA, Al-Tawfiq JA, Makhdoom HQ, Assiri A, Alhakeem RF, Albarrak A, et al. Respiratory tract samples, viral load, and genome fraction yield in patients with Middle East respiratory syndrome. *J Infect Dis.* 2014;210:1590–4. <http://dx.doi.org/10.1093/infdis/jiu292>
6. El Sabaawy D, El-Haggar S, El-Bahrawy, Waked I, El-Said H. A comparative study of variants of pegylated interferon alpha in treatment of chronic HCV patients. *APMIS.* 2015;123:482–9. <http://dx.doi.org/10.1111/apm.12377>
7. Faure E, Poissy J, Goffard A, Fournier C, Kipnis E, Titecat M, et al. Distinct immune response in two MERS-CoV-infected patients: can we go from bench to bedside? *PLoS ONE.* 2014;9:e88716. <http://dx.doi.org/10.1371/journal.pone.0088716>
8. Scheuplein VA, Seifried J, Malczyk AH, Miller L, Höcker L, Vergara-Alert J, et al. High secretion of interferons by human plasmacytoid dendritic cells upon recognition of Middle East respiratory syndrome coronavirus. *J Virol.* 2015;89:3859–69. <http://dx.doi.org/10.1128/JVI.03607-14>
9. Channappanavar R, Zhao J, Perlman S. T cell-mediated immune response to respiratory coronaviruses. *Immunol Res.* 2014;59:118–28. <http://dx.doi.org/10.1007/s12026-014-8534-z>
10. Perera RA, Wang P, Gomaa MR, El-Shesheny R, Kandeil A, Bagato O, et al. Seroepidemiology for MERS coronavirus using microneutralisation and pseudoparticle virus neutralisation assays reveal a high prevalence of antibody in dromedary camels in Egypt, June 2013. *Euro Surveill.* 2013;18:pii=20574.

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