recommended breakpoint, we designed MHC1 with a colistin concentration of only 1 μ g/mL to minimize false-negative results. However, some colistin-susceptible organisms might grow on MHC1 (<5% in our study), resulting in the low PCR-positive rate for *mcr-1* among isolates.

Exact epidemiology of the *mcr-1* gene is unknown, indicating a need to conduct accurate surveillance of the gene's prevalence in humans. Additional mechanisms unique to the *mcr-1* gene may contribute to colistin resistance, suggested by the wide variation in colistin MICs among *mcr-1*–carrying *Enterobacteriaceae*.

This study was partially supported by the Health and Medical Research Fund, Food and Health Bureau, Hong Kong Special Administrative Region Government (reference nos. HKM-15-M10 and HKM-15-M12).

References

- Liu YY, Wang Y, Walsh TR, Yi LX, Zhang R, Spencer J, et al. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. Lancet Infect Dis. 2016;16:161–8. http://dx.doi.org/10.1016/S1473-3099(15)00424-7
- Jorgensen JH, Pfaller MA, Carroll KC, Funke G, Landry ML, Richter SS, et al., editors. Manual of clinical microbiology, 11th ed. Washington: ASM Press; 2015.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. 25th edition. M100–S25. Wayne (PA): The Institute; 2015.
- Cheng VC, Chan JF, Wong SC, Chen JH, Tai JW, Yan MK, et al. Proactive infection control measures to prevent nosocomial transmission of carbapenem-resistant *Enterobacteriaceae* in a non-endemic area. Chin Med J (Engl). 2013;126:4504–9.
- Agriculture, Fisheries & Conservation Department. Hong Kong Special Administrative Region. Hong Kong: the facts. 2015 Feb [cited 2016 Jan 15]. http://www.gov.hk/en/about/abouthk/factsheets/ docs/agriculture.pdf
- Census and Statistics Department. Hong Kong Special Administrative Region. Hong Kong merchandise trade statistics imports. 2015 Apr [cited 2016 Jan 15]. http://www.statistics.gov.hk/ pub/B10200012015MM04B0100.pdf
- Falgenhauer L, Waezsada S-E, Yao Y, Imirzalioglu C, Käsbohrer A, Roesler U, et al.; RESET consortium. Colistin resistance gene *mcr-1* in extended-spectrum β-lactamase-producing and carbapenemaseproducing Gram-negative bacteria in Germany. Lancet Infect Dis. 2016;16:282–3. http://dx.doi.org/10.1016/S1473-3099(16)00009-8
- 8. Malhotra-Kumar S, Xavier BB, Das AJ, Lammens C, Butaye P, Goossens H. Colistin resistance gene *mcr-1* harboured on a multidrug resistant plasmid. Lancet Infect Dis. 2016;16:283–4. http://dx.doi.org/10.1016/S1473-3099(16)00012-8
- Tse H, Yuen KY. Dissemination of the mcr-1 colistin resistance gene. Lancet Infect Dis. 2016;16:145–6. http://dx.doi.org/10.1016/ S1473-3099(15)00532-0
- European Committee on Antimicrobial Susceptibility Testing.
 Breakpoint tables for interpretation of MICs and zone diameters:
 version 6.0. 2016 Jan 1 [cited 2016 Mar 15]. http://www.eucast.
 org/fileadmin/src/media/PDFs/EUCAST_files/Breakpoint_
 tables/v 6.0 Breakpoint table.pdf

Address for correspondence: Kwok-Yung Yuen, Carol Yu Centre for Infection, The University of Hong Kong, Queen Mary Hospital, Pokfulam Rd, Pokfulam, Hong Kong, China; email: kyyuen@hku.hk

Cryptococcus gattii Meningitis Complicated by Listeria monocytogenes Infection

Robert G. Deiss, Michael Bolaris, Angel Wang, Scott G. Filler

Author affiliations: Uniformed Services University of the Health Sciences, Bethesda, Maryland, USA (R.G. Deiss); Henry M. Jackson Foundation for the Advancement of Military Medicine, Bethesda (R.G. Deiss): Naval Medical Center of San Diego, San Diego, California, USA (R.G. Deiss); Harbor-UCLA Medical Center, Los Angeles, California, USA (M. Bolaris, A. Wang, S.G. Filler); David Geffen School of Medicine at UCLA, Los Angeles (S.G. Filler)

DOI: http://dx.doi.org/10.3201/eid2209.160142

To the Editor: Among immunocompetent persons with cryptococcal disease, infection with a second organism is rare; all reported cases have involved concomitant mycobacterial infections (1). Immunosuppression is not a necessary precondition for infection with *Cryptococcus gattii* (2), and among immunocompetent persons, *C. gattii* infection confers high mortality rates: up to 24% according to a large case series (3). In addition, cryptococcomas are frequently observed in patients with *C. gattii*, as opposed to *C. neoformans*, infection, commonly necessitating longer courses of treatment. We report a fatal case of *C. gattii* and *Listeria monocytogenes* co-infection in an immunocompetent woman with cryptococcomas.

The patient was a previously healthy 23-year-old Hispanic woman who was hospitalized in 2009 after weeks of headache and recent-onset diplopia. Lumbar puncture revealed elevated opening pressure of 52 cm H₂O; elevated leukocytes (1,030 cells/µL: 31% neutrophils, 55% lymphocytes, 14% monocytes); elevated protein concentration (117 g/L); and decreased glucose concentration (30 mg/dL). Cerebrospinal fluid (CSF) cryptococcal antigen (CrAg) titer was 1:64, and culture grew C. gattii. HIV antibody test result was negative. Magnetic resonance imaging of the brain demonstrated scattered enhancing round lesions within the cerebrum and cerebellum, consistent with cryptococcomas. The patient was prescribed intravenous amphotericin B (1 mg/kg/d) and intravenous flucytosine (2 g/6 h) (Table); after 5 days of therapy, culture of a repeat lumbar puncture sample was negative. The patient was then given intravenous liposomal amphotericin at 7 mg/kg, and after a 14-day induction period she was discharged with instructions to take fluconazole orally (400 mg $2\times/d$) and to continue amphotericin B infusions $(3\times/d)$ wk) (Table).

Table. Clinical events, management, and parameters for patient with Cryptococcus gattii meningitis complicated by Listeria

monocytogenes infection*

		Opening	Leukocyte			CrAg	
Clinical event		pressure, cm	count,	Protein,	Glucose,	titer	Culture result
(day)	Therapy (days)	H₂O (day)	cells/μL (day)	g/L (day)	g/L (day)	(day)	(day)
Days 1–15:	AMB 1 mg/kg (1-4); 5FC 2 g	52 (1), 12	1,030 (1),	117 (1),	30 (1), 29	1:64 (1)	Cryptococcus
induction therapy	q6h (1-14); L-AMB 7/mg/kg (5-	(12)	123 (12)	104 (12)	(12)	` ,	gattii (1),
	14)	` ,	` ,	, ,	, ,		negative (12)
Days 16-30:	L-AMB 7/mg/kg M,W,F (15-22);	46 (23)	111 (23)	81 (23)	34 (23)	1:8 (23)	Negative (23)
discharge,	FLZ 400 mg q12h (15-22); L-						
outpatient	AMB 5 mg/kg (23-30); FLZ 600						
infusion,	mg q12h: (23–30); 5FC 3 g q6h						
readmission	(23–30)						
Days 31–45:	L-AMB 5 mg/kg (31–45); FLZ	44 (38)	17 (38)	66 (38)	64 (38)	NA	Negative (38)
inpatient therapy	600 mg q12h (31–45); 5FC 3 g						
	q6h (31–45)						
Days 46–60:	L-AMB 5 mg/kg (46–60); FLZ	35 (48)	18 (48)	25 (48)	85 (48)	NA	Negative (48)
inpatient therapy	600 mg q12h (46–60); DEX 2						
	mg q6h (46–60)						
Days 61–75:	L-AMB 5 mg/kg (61–65); FLZ	13 (63)	8 (63)	28 (63)	91 (63)	1:4 (63)	Negative (63)
discharge and	600 mg q12h (61–75); DEX 2						
outpatient infusion	mg q8h (61–75); L-AMB						
D =0.00	7/mg/kg M,W,F (66–75)	(00)	1 0 10 (00)	0=0 (00)	4= (00)	4.4.00	
Days 76–83:	L-AMB 7/mg/kg M,W,F (76–79);	>55 (80)	1,010 (80)	258 (80)	17 (80)	1:4 (80)	Listeria
readmission/coma	DEX 2 mg q12h (76–79); FLZ						monocytogenes
(80); death (83)	600 mg q12h (76–83); CRO 2						(80)
	gm q12h (80–83); AMP 2 gm						
	q4h (80–83); TMP/SMX 320–						
	1,600 mg (2 double-strength						
	tablets) g8h (80–83)						

^{*5}FC, flucytosine; AMB, amphotericin B; AMP, ampicillin; CrAg, cryptococcal antigen; CRO, ceftriaxone; DEX, dexamethasone; F, Friday; FLZ, fluconazole; L-AMB, liposomal amphotericin; M, Monday; NA, not available; q, every; TMP/SMX, trimethoprim/sulfamethoxazole; W, Wednesday.

One week after hospital discharge, the patient experienced recurrent headache and low-grade fever and was readmitted. Repeat lumbar puncture indicated an opening pressure of 46 cm H₂O but improvement of all other clinical parameters (Table). CSF CrAg titer was 1:8 and culture was negative. Repeat brain magnetic resonance images revealed no hydrocephalus, minimal edema, and decreased size and number of cryptococcomas. She was again given amphotericin B (5 mg/kg/d) and intravenous flucytosine (3 g/6 h) and fluconazole (600 mg/12 h). Placement of a ventricular-peritoneal shunt was deferred, and the patient required frequent lumbar punctures to relieve elevated intracranial pressure. After 3 weeks of therapy, she began taking oral dexamethasone (2 mg 4×/d) to reduce intracranial pressure and control symptoms consistent with immune reconstitution inflammatory syndrome. After 30 days of antifungal therapy during this second hospitalization, she experienced symptomatic improvement and was discharged with amphoteric B (5 mg/kg to be infused $3\times$ /wk), fluconazole (600 mg $2\times/d$), and dexamethasone (tapering dosage).

Two weeks later (11 weeks after initial admission), she returned to the hospital with worsening headache and fever. Lumbar puncture demonstrated a leukocyte count of 1,010 cells/μL (74% neutrophils, 12% lymphocytes, 14% monocytes), glucose 17 mg/dL, protein 258 g/L, and an opening pressure of >55 cm H₂O. CSF culture grew *L. monocytogenes*. The patient was prescribed ceftriaxone, ampicil-

lin, and trimethoprim/sulfamethoxazole. Shortly after the lumbar puncture, she experienced status epilepticus and became comatose. Despite emergent craniotomy for relief of intracranial pressure, she remained comatose for several days; subsequently, supportive care was withdrawn and the patient died shortly thereafter.

This case highlights the difficulties of managing severe cryptococcal disease. This patient experienced headache over 3 months and symptom relapse during 10 weeks of anticryptococcal therapy. As was done in this case, practice guidelines recommend a longer duration of polyene antimycotic induction for patients with cryptococcomas than for those without (4), and longer courses of therapy are commonly described for infections caused by C. gattii than for those caused by C. neoformans (5). Corticosteroids are commonly used to treat immune reconstitution inflammatory syndrome associated with cryptococcal meningitis (6), although recently, they have been associated with adverse outcomes (7). As indicated by this case, corticosteroids remain a risk factor for secondary infection with several pathogens, including *Listeria*. No epidemiologic exposure to *Listeria* was identified for this patient.

C. gattii infection has been reported in 8 states, including California (3); we have found the pathogen in the soil south of Los Angeles, California, particularly in association with Canary Island pines and sweet gum trees (8). Some patients with C. gattii infection have autoantibodies to

granulocyte–macrophage (GM) colony-stimulating factor (9). Although these autoantibodies have not been reported in patients with *Listeria* infections, susceptibility to infection caused by this bacterium is increased in GM–colony-stimulating factor –/– mice (10). Autoantibodies against GM–colony-stimulating factor or perhaps other cytokines might have impaired the patient's host defense against these organisms; unfortunately, our report is limited by lack of serum for further testing.

This case demonstrates the difficulties of managing patients with *C. gattii* infection and an unusual co-infection with *L. monocytogenes*. Initiation of corticosteroids for the management of severe cryptococcal disease should be undertaken with caution. The differential diagnosis for worsening cryptococcal disease should include acute or subacute bacterial meningitis, particularly when the patient is receiving corticosteroids for the management of immune reconstitution inflammatory syndrome or associated complications.

Acknowledgments

We thank the patient's family for granting permission to publish this case.

References

- Musabende M, Mukabatsinda C, Riviello ED, Ogbuagu O. Concurrent cryptococcal meningitis and disseminated tuberculosis occurring in an immunocompetent male. BMJ Case Rep. 2016. pii:bcr2015213380. http://dx.doi.org/10.1136/bcr-2015-213380
- Chen SC; Australasian Society for Infectious Diseases (ASID)
 Mycoses Interest Group. Cryptococcosis in Australasia and the
 treatment of cryptococcal and other fungal infections with
 liposomal amphotericin B. J Antimicrob Chemother. 2002;49 Suppl
 1:57–61. http://dx.doi.org/10.1093/jac/49.suppl_1.57
- Harris JR, Lockhart SR, Sondermeyer G, Vugia DJ, Crist MB, D'Angelo MT, et al. *Cryptococcus gattii* infections in multiple states outside the US Pacific Northwest. Emerg Infect Dis. 2013;19:1620–6.. http://dx.doi.org/10.3201/eid1910.130441
- Perfect JR, Dismukes WE, Dromer F, Goldman DL, Graybill JR, Hamill RJ, et al. Clinical practice guidelines for the management of cryptococcal disease: 2010 update by the Infectious Diseases Society of America. Clin Infect Dis. 2010;50:291–322. http://dx.doi.org/10.1086/649858
- Franco-Paredes C, Womack T, Bohlmeyer T, Sellers B, Hays A, Patel K, et al. Management of *Cryptococcus gattii* meningoencephalitis. Lancet Infect Dis. 2015;15:348–55. http://dx.doi.org/10.1016/S1473-3099(14)70945-4
- Phillips P, Chapman K, Sharp M, Harrison P, Vortel J, Steiner T, et al. Dexamethasone in *Cryptococcus gattii* central nervous system infection. Clin Infect Dis. 2009;49:591–5. http://dx.doi.org/10.1086/603554
- Beardsley J, Wolbers M, Kibengo FM, Ggayi A-BM, Kamali A, Cuc NTK, et al.. Adjunctive dexamethasone in HIV-associated cryptococcal meningitis. N Engl J Med. 2016;374:542–54. http://dx.doi.org/10.1056/NEJMoa1509024
- Springer DJ, Billmyre RB, Filler EE, Voelz K, Pursall R, Mieczkowski PA, et al. *Cryptococcus gattii* VGIII isolates causing infections in HIV/AIDS patients in southern California: identification of the local environmental source as arboreal.

- PLoS Pathog. 2014; 10:e1004285. http://dx.doi.org/10.1371/journal.ppat.1004285
- Saijo T, Chen J, Chen SC-A, Rosen LB, Yi J, Sorrell TC, et al. Anti-granulocyte-macrophage colony-stimulating factor autoantibodies are a risk factor for central nervous system infection by *Cryptococcus gattii* in otherwise immunocompetent patients. MBio. 2014;5:e00912–4. http://dx.doi.org/10.1128/mBio.00912-14
- Zhan Y, Lieschke GJ, Grail D, Dunn AR, Cheers C. Essential roles for granulocyte-macrophage colony-stimulating factor (GM-CSF) and G-CSF in the sustained hematopoietic response of *Listeria* monocytogenes-infected mice. Blood. 1998;91:863–9.

Address for correspondence: Robert G. Deiss, Division of Infectious Diseases, Naval Medical Center of San Diego, 34800 Bob Wilson Dr, San Diego, CA 92134, USA; email: robert.g.deiss.ctr@mail.mil

Melioidosis in Travelers Returning from Vietnam to France

Jérôme Gauthier,¹ Patrick Gérôme,¹ Maryline Defez,¹ Fabienne Neulat-Ripoll, Barbara Foucher, Thierry Vitry, Lionel Crevon, Eric Valade, François M. Thibault, Fabrice V. Biot

Author affiliations: Hôpital d'Instruction des Armées Desgenettes, Lyon, France (J. Gauthier, P. Gérôme, B. Foucher, T. Vitry, L. Crevon); Centre Hospitalier Universitaire, Grenoble, France (M. Defez); Institut de Recherche Biomédicale des Armées, Brétigny-sur-Orge, France (F. Neulat-Ripoll, E. Valade, F.V. Biot); Ecole du Val-de-Grâce, Paris, France (E. Valade); Direction Centrale du Service de Santé des Armées, Paris (F.M. Thibault)

DOI: http://dx.doi.org/10.3201/eid2209.160169

To the Editor: Melioidosis, a potentially fatal infectious disease, occurs predominantly across much of Asia and in northern Australia because of the soil and water bacterium *Burkholderia pseudomallei* (1). We report 2 related cases of suppurative cervical lymphadenitis, an unusual adult presentation of melioidosis, in 2 men who returned to France from Vietnam on the same trip (2).

Patient 1, a 28-year-old previously healthy man, was admitted to our hospital in Lyon, France, in October 2013 for the evaluation of a palpable neck mass, which had been growing steadily for the previous 2 months. Examination of the head and neck revealed a fluctuant, tender mass located in the inferior angle of the right side of the mandible, mimicking lymph node tuberculosis. Ultrasonographic investigation confirmed a level II enlarged cervical lymph node

¹These authors contributed equally to this article.