## Article DOI: https://doi.org/10.3201/eid3005.231383

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# Recurrence, Microevolution, and Spatiotemporal Dynamics of *Legionella pneumophila* Sequence Type 1905, Portugal, 2014–2022

## Appendix 2

## **Materials and Methods**

### Selection of samples and isolates

This study includes all (n = 14) *L. pneumophila* samples and isolates collected after the large outbreak in 2014 that were identified as serogroup 1 and ST1905, in the context of the National Legionnaires' Disease Surveillance Programme, by the National Reference Laboratory for *Legionella*, in Lisbon, Portugal. In detail, it includes: 12 isolates (6 clinical and 6 environmental) and two positive specimens. A representative genome sequence of a ST1905 *L. pneumophila* PtVFX/2014 isolate collected during the 2014 outbreak, which was previously characterized (*1*), was included in this study as a reference. Details about all studied samples are available at the Appendix 1 Table 1 (https://wwwnc.cdc.gov/EID/article/30/5/23-1383-App1.xlsx).

### **DNA extraction and quantification**

Isolation of bacterial DNA from 12 *L. pneumophila* isolates and from 2 *L. pneumophila* PCR-positive culture-negative samples was performed on the EMAG<sup>®</sup> platform (bioMérieux), an automated nucleic acid extraction platform based on magnetic silica technology. DNA was quantified by fluorescence spectroscopy using the Qubit Fluorometer (ThermoFisher Scientific).

## L. pneumophila Sequence-Based Typing (SBT) and Whole Genome Sequencing (WGS)

SBT was performed at the National Reference Laboratory for *Legionella* using the DNA retrieved from pure cultures and uncultured clinical samples using the amplification of the 7 loci

in the SBT scheme, following the protocol of the European Working Group for *Legionella* infections (2). Sequencing libraries of DNA obtained from the *L. pneumophila* isolates (n = 12) was prepared using the Nextera XT DNA Sample Preparation Kit (Illumina), following manufacturer's instructions. Whole Genome Sequencing (WGS) was performed using Illumina MiSeq equipment (2x250 bp), as previously described (*1*).

### Phylogeny and microevolutionary analysis

INNUca v4.2.2 (https://github.com/B-UMMI/INNUca) was applied to perform quality control of reads, draft de novo assembly and contigs quality assessment, and contamination detection. ReporType (3) (<u>https://github.com/insapathogenomics/ReporType</u>) was applied to perform in silico prediction of the L. pneumophila subspecies and serogroup from the draft assemblies. SBT profile ST1905 was also confirmed in silico through BLASTn (also using ReporType) against the SBT allele database available in https://github.com/tseemann/legsta. Snippy v4.6.0 (https://github.com/tseemann/snippy; settings:-mapqual 20-mincov 10-minfrac 0.51-basequal 20) was applied to the INNUca quality processed reads to perform referencebased mapping and SNP/indel analysis using the L. pneumophila PtVFX/2014 draft genome annotation (LORH0000000.1) as reference. Core genome SNP alignments were extracted from the Snippy-derived whole SNP alignment using ReporTree 1.1.2 (4). Whenever a mutation was reported for any of the studied isolates, the mapping files (BAM) of the other isolates were visually inspected in Integrative Genomics Viewer (IGV, https://igv.org/app/) to confirm/exclude its presence. Mapping files were also used to search and confirm the presence of indel events. A maximum phylogenetic likelihood tree was then constructed based on the curated core-genome SNP alignment (excluding SNPs found in ~2.5Kb recombination event found in reference contig 8, positions 8985–08995) using the Tamura 3-parameter evolutionary model in MEGA 11 (https://www.megasoftware.net/), incorporating 1000 random bootstrap replicates to assess node support within the tree.

#### References

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