NEWS & NOTES

virus persistence, and flaviviral recombination as a mechanism of flaviviral evolution. In addition, data were presented that illustrated the persistence of, and immune modulation by, alphaviruses, which, in concert, allow the virus to replicate while preventing the host from responding to its benefit.

Other than the classical techniques of preventing infection, little was mentioned about disease control during this symposium. Control must be based on rapid recognition of early cases, subsequent immunization of persons or animals at risk, or immunization of persons or animals with the potential to be at risk, such as travelers, laboratory personnel, and attending clinicians. Attendees learned about diverse methods being used to develop vaccines. Representatives from the World Health Organization explained that organization's plans for responding to disease emergence and for preventing zoonotic diseases from reaching human populations.

New paradigms for field studies of zoonotic diseases are necessary. These approaches must include longitudinal and in-depth investigations of agent, host, habitat, and environment if we are to predict risk and respond in an appropriate manner. At this time, zoonotic disease control comprises prevention and public education and not much more. Progress is being made in rapid diagnosis, production of sophisticated vaccines, and understanding of the molecular mechanisms by which zoonotic viruses persist and cause disease. Most of the papers presented will be published in a special issue of Archives of Virology.

Charles H. Calisher,* Betty Dodet,† and Diane Griffin‡

*Colorado State University, Fort Collins, Colorado, USA: †Fondation Mérieux, Lyon Cedex, France; and ‡Johns Hopkins Bloomberg School of Public Health, Baltimore, Maryland, USA

Address for correspondence: Charles H. Calisher, Arthropod-borne and Infectious Diseases Laboratory, Department of Microbiology, Immunology and Pathology, College of Veterinary Medicine and Biomedical Sciences, Colorado State University, Ft. Collins, CO 80523, USA; fax: (970) 491-8323; email: calisher@cybersafe.net

Conference Summary

New and Reemerging Infectious Diseases

The Sixth Annual Conference on New and Reemerging Infectious Diseases was hosted April 24–25, 2003, by the Center for Zoonoses Research and the College of Veterinary Medicine, University of Illinois at Urbana-Champaign (UIUC). The conference featured seven speakers and 27 poster presentations.

Smallpox

Bertram L. Jacobs (Arizona State University, Tempe, AZ) opened the conference with a presentation on smallpox, one of the most devastating diseases known to humankind. Smallpox was eradicated from the wild in the 1970s, although the potential use of Variola virus as a bioterrorism agent makes it still of great concern. Dr. Jacobs described Vaccinia viruses deficient in E3L, a regulator of the cellular antiviral response and noted their potential for the production of improved vaccines. He also showed that double-stranded (ds)RNA- and ZDNA binding proteins had a role in poxvirus pathogenesis. In the poster section, Joanna Shisler (University of Illinois at Urbana-Champaign [UIUC], Urbana) reported that the modified virus, Ankara, activates nuclear factor kB through the mitogen-activated protein kinase, extracellular signal-regulated kinase (MEK)/ERK extracellular signal-regulated kinase (ERK) pathway, possibly facilitating the host immune response. This virus was used to vaccinate 100,000 people, with no reported complications, at the end of the global smallpox vaccination campaign led by the World Health Organization in the 1970s.

West Nile Virus and Geographic Information Systems

Since it was first detected in New York City in 1999, West Nile virus (WNV) has spread from coast to coast and has been found in 43 states from Maine to California. Stephen C. Guptill (U.S. Geological Survey,

Reston, VA) reported that the U.S. Geological Survey is working with the Centers for Disease Control and Prevention (CDC) to learn the current geographic extent of WNV. This will allow us to understand how it moves between birds, mosquitoes, and humans and to better predict future outbreaks. A collaborative 3-year research project is being conducted on lands administered by the U.S. Fish and Wildlife Service, the National Park Service, and other federal lands, and on state, local, and private lands along the Atlantic and Mississippi flyways. This study tests sampled migratory and local wild birds to detect WNV and identify possible avian carriers. Over 10,000 birds of more than 150 species have been captured, sampled, and released at 20 federal sites and 3 other sites in 12 states during the spring and fall bird migration seasons of 2001 and 2002. A parallel study, conducted with CDC, is examining the distribution and number of mosquito species in relation to land cover, weather conditions, and avian deaths. Systematic mosquito surveillance (weekly collections at seven

sites) is being conducted year-round in St. Tammany Parish in Louisiana, complementing avian collections at the Bogue Chitto and Big Branch National Wildlife Refuges in the parish. Finally, WNV surveillance data from CDC is being studied to determine the spatial and temporal relationships between disease outbreaks in birds and animals and human illness. Information from these analyses will guide the creation of predictive models of disease risk. These surveillance systems provide the basic information on the "geography" of the virus. Combining these data with information about avian migratory patterns, landscape characteristics, and weather conditions, over space and time, will provide the foundation for developing spatial analytical and forecasting models to assess the risk for human illness. In related work, presented at the poster session, Marylin Ruiz (UIUC, Urbana) reported the efforts of the College of Veterinary Medicine Geographic Information System and Spatial Analysis Laboratory, in collaboration with the Illinois Department of Public Health, and the Illinois Department of Agriculture in the mapping and analysis of the WNV outbreak in Illinois. (Illinois was the state hit the hardest by the epidemic in 2002.) Geographic information systems in conjunction with fine resolution satellite data and spatial statistics are also useful to investigate the distribution of other diseases, for example, schistosomiasis (Julie A. Clennon, UIUC, Urbana).

Animal Models of Infectious Diseases

Streptococcal pathogens continue to evade concerted efforts to decipher clear-cut virulence mechanisms, although numerous genes have been implicated in pathogenesis. Melody N. Neely (Wayne State University, Detroit, MI) reported the development of a unique animal model, the zebrafish (*Danio rerio*), to characterize specific virulence mechanisms used within various tissues in vivo. Her group is using this model host to study infection by two streptococcal species that represent two forms of streptococcal disease: a natural pathogen of fish and humans, Streptococcus initiae, and a humanspecific pathogen, S. pyogenes. S. initiae primarily causes a fatal systemic disease in the zebrafish after intramuscular injection, with pathologic changes similar to those seen in human infections caused by S. agalactiae and S. pneumoniae. The fatal infection by S. pyogenes causes a locally spreading necrotic disease confined to the muscle with pathologic features similar to those observed in a human infection of necrotizing fasciitis. By studying pathogens that are virulent for both fish and humans and that mediate disease states in the zebrafish identical to those found in human streptococcal infections, common virulence strategies shared by a number of gram-positive pathogens can be identified. Using several genetic strategies with the two streptococcal strains, Dr. Neely's group is currently conducting specific screens in the zebrafish to 1) identify and characterize cell membrane proteins that interact with the host in vivo to cause specific disease states; 2) identify genes required for growth in vivo, as well as progressive stages of infection; 3) identify genes that are only expressed while in the host along with tissue specificity of the encoded proteins; and 4) analyze responses of the host that affect progression of disease.

Foodborne Diseases

Shigella boydii is a food pathogen that was implicated in a 1999 foodborne outbreak involving contaminated bean salad that contained fresh parsley and cilantro. Hans Blaschek and collaborators (UIUC, Urbana) reported the high tolerance of this bacterium to acidic pH, and the presence and formation of biofilms in cilantro and parsley samples treated with produce wash and water. Cells in biofilms are known to be more resistant to antibiotics and disinfectants, and biofilm formation may explain the decreased efficacy of produce wash on parsley and cilantro samples.

Intracellular Parasites

The trypanosomatid protozoan Leishmania synthesizes a variety of glycosylphosphatidylinositol (GPI GPI)-anchored molecules on its surface. Sorting out their individual functions has been difficult and in some cases controversial as they share structural domains and generally have been tested outside the context of key components such as the major glycolipid lipohosphoglycan (LPG) and phosphoglycans (PGs); ether lipids, which constitute approximately 20% of the membrane lipids and most GPI anchors; and others. Stephen M. Beverley and his collaborators (Washington University, St. Louis, MO) have studied their role in the context of parasite knockouts and "add-back" controls, probing their role in diverse aspects of parasitism such as entry, inhibition of phagolysosomal fusion and host-signal transduction, and pathogenesis. All of these molecules play critical roles in virulence, although sometimes in unanticipated ways. PGs in particular are required for parasite persistence but not acute pathology, therefore defining a new class of parasite genes important to transmission. Ted Hackstadt (National Institute of Allergy and Infectious Diseases, National Institutes of Health, Hamilton, MT) reported that, unlike the majority of most intracellular parasites, which block maturation of endosomes to lysosomes at discrete stages and then replicate within those vacuoles, chlamydiae appear to dissociate themselves from the endocytic pathway shortly after internalization by actively modifying the vacuole to become fusogenic with sphingomyelin-containing exocytic vesicles. Interaction with a secretory pathway appears to provide a pathogenic mechanism that allows chlamydiae to establish themselves in a site not destined to fuse with lysosomes. Fusion with Golgi-derived vesicles provides a likely source of cellular lipids for the growth of the inclusion membrane as it expands to accommodate the multiplying parasites.

Kasturi Haldar (Northwestern University, Chicago, IL) presented studies on the malaria parasite, focusing on protein trafficking, gene expression, and drug development. Her group has studied vacuolar trafficking of host raft proteins and parasite virulence determinants (by tagging genes with GFP and expressing them by transfection) for their consequences on malarial entry into the red blood cell, virulence secretion systems, and apicoplast biogenesis. The apicoplast is a newly identified residual plastid acquired by secondary endosymbiosis that has attracted attention for its evolutionary novelty and its potential as a drug target. Temporal regulation of plasmodial genes may be important for proteintargeting in cells. In this context, Dr. Haldar's group is examining the role of unique promoter elements and chromatin in regulating expression of secretory determinants such as the histidine-rich proteins and adherence antigens. Whole genome scanning approaches (microarrays and other functional approaches) are being used in combination with informatics to develop novel lipid-linked targets for development. Studies drug on Salmonella are currently examining effectors of the SPI-2 system (Salmonella Pathogenicity Island-2 system) for their effects on sterol recruitment, metabolism, and bacterial virulence in the mouse model. The requirement for nonsterol precursors in protecting infected cells from death by apoptotis, necrosis, or both, is also being investigated. Finally, microarrays are used to identify subsets of *S. typhimurium* virulence determinants required for lipid-linked intracellular bacterial replication.

Toxoplasma gondii is a major cause of birth defects and infections in immunocompromised persons. Like of all members the phylum Apicomplexa, T. gondii is an obligate intracellular organism. Micronemes and rhoptries are specialized secretory organelles of the Apicomplexa, whose contents are thought to be essential for successful invasion of host cells. Kami Kim (Albert Einstein College of Medicine, Bronx, NY) reported identifying two subtilisin-like serine proteinases from T. gondii, TgSUB1 and TgSUB2, which are necessary for successful invasion. Serine proteinase inhibitors have been reported to block host cell invasion by both T. gondii and the related apicomplexan parasite, P. falciparum. Disruption of TgSUB2 was unsuccessful, which implies that TgSUB2 is an essential gene. Both TgSUB1 and TgSUB2 undergo autocatalytic processing as they traffic through the secretory pathway. TgSUB1 is a microneme protein, whereas TgSUB2 localizes to rhoptries and associates with rhoptry protein ROP1, a potential substrate. Mutational analysis suggests that TgSUB2 is a rhoptry protein mat-Processing of secretory urase. organelle contents appears to be ubiquitous among the Apicomplexa. Since subtilases are found in genomes of all the Apicomplexa sequenced to date, may represent a novel they chemotherapeutic target. Transcriptional regulatory pathways in apicomplexan parasites are understudied and may contain novel drug targets. William Sullivan and his collaborators (Indiana University School of Medicine, Indianapolis, IN) identified and mapped a gene in T. gondii that encodes a homologue of chromatin remodeling factors that uses ATP to promote a more favorable environment for transcription. They also

described a novel histone acetyltransferase, which contains a unique 820amino acid *N*-terminal extension of unknown function.

Parasite Organelles

Acidocalcisomes are novel calcium-containing acidic organelles present in unicellular eukaryotes. Several posters from researchers in the groups of Silvia Moreno and Roberto Docampo (UIUC, Urbana) highlighted recent work on these organelles. Andrea Montalvetti described a functional aquaporin (water channel) found in the organelles of T. cruzi, the etiologic agent of Chagas disease, and Peter Rohloff showed that this protein is translocated to the contractile vacuole upon hypo-osmotic stress. Joanna Cox and Shuhong Luo demonstrated that a Ca2+-ATPase is localized to acidocalcisomes and the plasma membrane of Trypanosoma brucei, the etiologic agent of African sleeping sickness, and is essential for cell growth, as demonstrated by RNA interference experiments. Manfredo Seufferheld presented evidence of the presence acidocalcisomes in the bacterium Rodospirillum rubrum. Felix Ruiz showed that acidocalcisomes of Plasmodium falciparum, one of the etiologic agents of malaria, do not differ significantly from the organelles in other apicomplexan parasites, whereas the platelet-dense granules possess several characteristics common to acidocalcisomes. T. brucei possesses a Ptype proton ATPase with homology to fungal and plant H+-ATPases but absent in mammalian cells; this proton constitutes a formidable target for chemotherapy (Shuhong Luo, UIUC, Urbana).

Chemotherapeutic Targets

A number of poster presentations identified novel chemotherapeutic targets for infectious diseases. Steve Grimme (UIUC, Urbana), who received an award for the best poster presentation, demonstrated that a GPI mannosyltransferase (CaSMP3) was essential for the human pathogenic fungus Candida albicans and therefore was validated as a potential target for chemotherapy. M. Laura Salto (UIUC, Urbana) showed that inositolphosphoceramide, a lipid used in anchoring several proteins to the plasma membrane of T. cruzi and absent in the host, is remodeled during the differentiation of different stages of the parasite. Novel lipid modified phosphoinositide phospholipases C were reported in T. cruzi (Michael Okura, UIUC, Urbana) and T. brucei (Jianmin Fang, UIUC, Urbana). The

enzymes are apparently involved in differentiation of the parasites and are different from their host counterparts. Recently, bisphosphonates have been proposed as novel antiparasitic drugs, and Yan Ling (UIUC, Urbana) reported the cloning and characterization of Toxoplasma gondii farnesyl pyrophosphate synthase, the target of these compounds. Tryptophan metabolism in mosquitoes and the separation and identification of mosquito chrorion proteins through two-dimensional electrophoresis and mass spectrometry will provide useful targets against these vectors of several infectious diseases, as reported by Jianyong Li and his collaborators (UIUC, Urbana).

Roberto Docampo*

*University of Illinois at Urbana-Champaign, Urbana, Illinois, USA

Address for correspondence: Roberto Docampo, Laboratory of Molecular Parasitology, Department of Pathobiology and Center for Zoonoses Research, University of Illinois at Urbana-Champaign, 2001 South Lincoln Avenue, Urbana, IL 61802, USA; email: rodoc@uiuc.edu

The Yellow Book

The Division of Global Migration and Quarantine, National Center for Infectious Diseases, Centers for Disease Control and Prevention, announces the availability of the 2003-2004 edition of Health Information for International Travel (The Yellow Book). Features of this edition include new recommendations for malaria chemoprophylaxis; updated, expanded sections on injury during travel, motion sickness, altitude sickness, and travelers with disabilities; revised vaccine recommendations; changes in recommendations for insect repellents; a new chapter focusing on recommendations for children; new text on scuba diving safety and high-risk travelers; and improved maps and indexing.

The Yellow Book will also be available on CD-ROM later in the year. To order the book or the CD-ROM, contact the Public Health Foundation at 1-877-252-1200 or http://bookstore.phf.org.