LETTERS

create a niche for *M. tuberculosis* (10).

Our hypothesis could be refuted or corroborated in several ways, for example, by a case-control study of HIV-negative patients infected with tuberculosis. If this study refutes our hypothesis, the idea that sex hormones play a direct role in the immune response to *M. tuberculosis* would be supported. Such findings might also provide possibilities for drug development. However, if casecontrol studies support our hypothesis, attempts should be made to identify the pathogen.

Nico J.D. Nagelkerke,*† Sake J. de Vlas,‡ Kelly S. MacDonald,§ and Hans L. Rieder¶

*Leiden University Medical Center, Leiden, the Netherlands; †University of Manitoba, Manitoba,Winnipeg, Canada; ‡University Medical Center, Rotterdam, the Netherlands; §Mount Sinai Hospital, Toronto, Ontario, Canada; and ¶International Union Against Tuberculosis and Lung Disease, Paris, France

References

- Rieder HL. Epidemiological basis of tuberculosis control. Paris: International Union Against Tuberculosis and Lung Disease; 1999.
- 2. Dubos RJ, Dubos J. The white plague: tuberculosis, man and society. Camden (NJ): Rutgers University Press; 1952.
- 3. Hamilton JB, Mestler GE. Mortality and survival: comparison of eunuchs with intact men and women in a mentally retarded population. J Gerontol. 1969;24:395–411.
- Bleiker MA, Douma J, Van Geuns HA, Van Joost CRNF, Manten A, Meijer J, et al. Leerboek der tuberculosebestrijding. The Hague: KNCV; 1984.
- 5. Quetel C. Le mal de Naples. Histoire de la syphilis. Paris: Seghers; 1986.
- Frost WH. The age selection of mortality from tuberculosis in successive decades. Am J Epidemiol. 1995;141:4–9.
- Holmes CB, Hausler H, Nunn P. A review of sex differences in the epidemiology of tuberculosis. Int J Tuberc Lung Dis. 1998;2:96–104.
- Barnes PF, Silva C, Otaya M. Testing for human immunodeficiency virus infection in patients with tuberculosis. Am J Respir Crit Care Med. 1996;153:1448–50.
- 9. Stephens TT, Braithwaite R, Cozza S, Robillard A, Arriola KJ. History of prior TB

infection and HIV/AIDS risk behaviours among a sample of male inmates in the USA. Int J STD AIDS. 2003;14:514–8.

 Redpath S, Ghazal P, Gascoigne SR. Hijacking and exploitation of IL-10 by intracellular pathogens. Trends Microbiol. 2001;9:86–92.

Address for correspondence: Nico J.D. Nagelkerke, Department of Medical Statistics, Leiden University Medical Center, P.O. Box 9604, 2300 RC Leiden, the Netherlands; fax: 31-71-5276799; email: n.j.d.nagelkerke@lumc.nl

Leptotrichia amnionii and the Female Reproductive Tract

To the Editor: Detection of new bacteria, human complex microflora, by using 16S rRNA gene amplification and sequencing has been reported (1). 16S rRNA gene amplification and sequencing detected pyosalpinx, caused by *Leptotrichia amnioni*, in a patient whose samples were culture-negative.

This anaerobic gram-negative bacterium has been isolated only once before (2). A 41-year-old woman from the island of Comoros who had been having lower abdominal pain for 6 days was admitted to the emergency department of Hôpital Nord in Marseille. The patient's history included type 2 diabetes mellitus treated by metformin and laparoscopy to explore infertility. On examination, the patient had a pulse rate of 90 beats per min, a blood pressure of 130/80 mm Hg, and a temperature of 38.5°C. Her abdomen was not distended, but diffused lower abdominal tenderness, especially at the right iliac fossa, was present. Blood testing showed a leukocyte count of 7.7x109/L, hemoglobin of 13.1g/dL, and platelet count of 213x109/L. The chemistry

panel showed hyperglycemia (14.1 mmol/L) and elevated C-reactive protein (254 mg/L). Renal and liver function test results were all within normal limits. Serum β-human chorionic gonadotropin was negative. A computed tomographic scan of the abdomen and pelvis showed two septated adnexal masses, a 12x7x5 cm mass on the right and a 6x4x2 cm mass on the left; the patient was referred to the gynecologic surgery department. Gynecologic examination showed greenish, purulent vaginal discharge and a fluctuant mass in the pouch of Douglas. Uterine cervical motion caused pain to the patient. Transabdominal and transvaginal ultrasound scan showed a 10x7x5 cm homogeneous liquid mass in the pouch of Douglas.

The patient was taken to the operating room and prepared for surgery. The gynecologic team performed a laparotomy that showed a 5-cm, left hydrosalpinx and a 10-cm, right tuboovarian abscess adherent to the uterus, sigmoid colon, pelvic sidewall, and pouch of Douglas. The appendix and other viscera were normal. The adhesiolysis led to the rupture of the abscess and discharge of clear greenish pus, a sample of which was sent to the laboratory for culture. Antimicrobial drug treatment was started with intravenous cefazolin, gentamicin, and metronidazole. On the first postoperative day, the patient was afebrile. Oral amoxicillin plus clavulanic acid was administered for 15 days, and oral ciprofloxacin was administered for 20 days. The patient was discharged on day 7 of hospitalization and was well at the follow-up examination 1 month later.

After Gram staining, a sample of the abscess drainage was injected onto Columbia agar with 5% sheep blood (bioMerieux, Marcy l'etoile, France) under 5% CO_2 and anaerobic atmosphere. Antimicrobial susceptibility of the sample was tested by an agar diffusion method (3). A drop of the sample was deposited on an agar plate flooded with a suspension of a antimicrobial susceptible strain of *Micrococcus luteus*. After 24 h of incubation at 37° C, presence of antimicrobial activity in the sample was evident by a visible area of growth inhibition of *M. luteus* around the sample. Procedures for DNA extraction and 16S rRNA gene amplification and sequencing have been detailed (4).

Gram staining of the sample showed numerous polymorphonuclear leukocytes and gram-negative bacteria. Culture of the sample remained sterile after 20 days of incubation, and antimicrobial susceptibility was found. The 16S rRNA gene amplification and sequencing determined a 1,493 nucleotide sequence. This sequence had 99.7% nucleotide similarity with that of L. amnionii (GenBank accession no. AY078425), which corresponded to a difference of 4 nucleotide. The 16S rRNA gene sequence of the detected bacterium was deposited under accession no. AY489565. L. amnionii was previously recovered in anaerobic culture of the amniotic fluid of a woman after intrauterine fetal demise (2). It was isolated on blood and chocolate agar under anaerobic conditions and showed very small gray colonies of <1 mm. This slow-growing bacterium was lost after two subcultures, and no isolate is available for further description (2). In that case and in the case reported here, the patients had uneventful recoveries after an amoxicillin plus clavulanic acid antimicrobial regimen was given. This bacterium and our isolate are related to, but different from, L. sanguinegens.

Leptotrichia is a small genus closely related to Fusobacterium and comprises slow-growing, gram-negative, filamentous, anaerobic bacterial flora of the oral cavity and genital tract (5). Species included in the genus are L. buccalis, L. trevisanii, L. sanguinegens, and L. amnionii (2,6).

All *Leptotrichia* species are extremely fastidious and cannot be grown easily on conventional microbiologic media or by conventional methods. As evidenced by sequences available in the GenBank database, most of the 16S rRNA gene sequences are from cloned DNA from complex flora but from bacterial not isolates. Leptotrichia species has been suspected to play a role in periodontal disease. However, Leptotrichia species have only been associated with serious systemic disease, usually in immunocompromised patients (7,8). Bacteremia caused by L. sanguinegens in pregnant women has also been reported (9). More widespread use of polymerase chain reaction amplification and sequencing of the 16S rRNA gene for identification or detection of fastidious pathogens in humans will likely provide verification of several new pathogens that are now part of normal human flora.

Vijay A.K.B.Gundi,* Raoul Desbriere,† and Bernard La Scola*

*Unité des Rickettsies, Marseille, France; and †Hôpital Nord, Marseille, France

References

- Leys EJ, Lyons SR, Moeschberger ML, Rumpf RW, Griffen AL. Association of *Bacteroides forsythus* and a novel *Bacteroides* phylotype with periodontitis. J Clin Microbiol. 2002;40:821–5.
- 2. Shukla SK, Meier PR, Mitchell PD, Frank DN, Reed KD. *Leptotrichia amnionii* sp. nov., a novel bacterium isolated from the amniotic fluid of a woman after intrauterine fetal demise. J Clin Microbiol. 2002;40:3346–9.
- Zannier A, Drancourt M, Franceschi JP, Aubaniac JM, Raoult D. Interest of the centrifugation lysis method for bacterial isolation from bone and joint specimens. Pathol Biol (Paris). 1991;39:543–6.
- Raoult D, Birg ML, La Scola B, Fournier PE, Enea M, Lepidi H, et al. Cultivation of the bacillus of Whipple's disease. N Engl J Med. 2000;342:620–5.
- Holt JG, Kreig NR, Sneath PH, Staley JT, Williams ST. Bergey's manual of determinative bacteriology. 9th ed. Baltimore: Williams & Wilkins; 1994.

- Collins MD, Hoyles L, Tornqvist E, von Essen R, Falsen E. Characterization of some strains from human clinical sources which resemble "*Leptotrichia sanguinegens*": description of *Sneathia sanguinegens* sp. nov., gen. nov. Syst Appl Microbiol. 2001;24:358–61.
- Weinberger M, Wu T, Rubin M, Gill VJ, Pizzo PA. *Leptotrichia buccalis* bacteremia in patients with cancer: report of four cases and review. Rev Infect Dis. 1991;13:201–6.
- Tee W, Midolo P, Janssen PH, Kerr T, Dyall-Smith ML. Bacteremia due to *Leptotrichia trevisanii* sp. nov. Eur J Clin Microbiol Infect Dis. 2001;20:765–9.
- Hanff PA, Rosol-Donoghue JA, Spiegel CA, Wilson KH, Moore LH. *Leptotrichia sanguinegens* sp. nov., a new agent of postpartum and neonatal bacteremia. Clin Infect Dis. 1995;20(Suppl. 2):S237–9.

Address for correspondence: Bernard La Scola, Faculté de Médecine, Université de la Méditerrannée, 27 Boulevard Jean Moulin, 13385 Marseille Cedex 05, France; fax: 33-04-91-83-03-90; email: bernard.lascola@medecine. univ-mrs.fr

Cholera in Mozambique, Variant of *Vibrio cholerae*

To the Editor: Cholera outbreaks caused by toxigenic Vibrio cholerae serogroup O1 frequently occur in many sub-Saharan African countries. The serogroup O1 is classified into two biotypes, classical and El Tor. The seventh and current pandemic of cholera is caused by the El Tor biotype; the classical biotype is believed to be extinct. The classical and El Tor biotypes of V. cholerae O1 are closely related in their O-antigen biosynthetic genes but differ in other regions of the genome. The genomic structure of the CTX Φ filamentous phage (1), in which the cholera toxin genes are contained, differs between the classical and El Tor biotypes. $CTX^{class}\Phi$ is found in classical strains, $CTX^{ET}\Phi$ is