Echinococcus multilocularis in Estonia

To the Editor: Alveolar echinococcosis (AE) caused by Echinococcus multilocularis is one of the most important emerging zoonosis in Europe. The fatality rate is >90% in untreated patients (1). In Europe, the distribution range of the zoonotic tapeworm E. multilocularis has expanded over the last few decades, and the parasite attracts increasing awareness as a public health issue (2–5). In 2003, AE was added to the list of zoonoses to be monitored in the member states of the European Union, according to Directive 2003/99/EC.

This is the first report of E. multilocularis in Estonia, which extends its northern distribution in Europe. Results of examinations of 17 red foxes shot in the eastern (Võnnu and Räpina) and western (Hiiumaa) districts of Estonia from February to December 2003 were included in this study. We examined the intestinal tracts by the sedimentation and counting technique as described (1). Echinococcus adult stages were found in 5 foxes (29.4%). Two foxes, infected with 3 and 5 adult worms, were from the Räpina district; 2 foxes, infected with 66 and 133 worms, were from the Võnnu district; and 1 fox, infected with the highest number of worms (927), was from the Hiiumaa District. The worms were retrieved, counted, washed, and stored in 90% ethanol until DNA purification. The parasites were identified as E. multilocularis, based on the most important morphometric parameters of adult stages (length of worms, number of proglottids, terminal proglottids in percentage of total worm length, position of genital pore, and form of uterus) (2).

To confirm the taxonomic status of the worms, polymerase chain reaction (PCR) was conducted, followed by restriction fragment length polymorphism (RFLP) analysis and direct sequencing of a portion of the NADH dehydrogenase subunit I (ND1) gene of the mtDNA. A total of 6 specimens of E. multilocularis were used for genetic analysis. Total genomic DNA was extracted with the High Pure PCR Template Preparation Kit (Roche Molecular Biochemical, Mannheim, Germany) according to manufacturer’s instructions. PCR-RFLP was performed as described by Gonzalez et al. (6). The RFLP pattern of E. multilocularis isolates differed from that of E. granulosus. Diagnostic cleavage at the locus Eg9 of E. multilocularis with the enzyme CfoI is able to distinguish E. multilocularis and its closest relative E. granulosus (Figure, lanes 3 and 4 vs. lane 10). All 6 specimens of E. multilocularis produced identical results. A 426-bp fragment of the mitochondrial ND1 gene was amplified with the primers NDfor2-AGTTTTGTAAGGTCCTAATA and NDrev2-CCCACTAACTAATTCCCTTTTC using the BD Advantage 2 PCR Kit (Becton Dickinson Biosciences, Franklin Lakes, NJ, USA) as described (7). DNA cycle sequencing was performed using the DYEnamic ET Terminator Cycle Sequencing Kit (Amersham Pharmacia Biotech, Piscataway, NJ, USA). Sequences were resolved on an ABI PRISM 377 automated DNA sequencer (Applied Biosystems, Foster City, CA, USA).

All analyzed E. multilocularis specimens had identical sequences. The ND1 sequence of E. multilocularis from Estonia was submitted to GenBank under accession no. AY855918. The nucleotide sequences obtained were compared with those in the GenBank sequence database. The sequence of the Estonian isolate was identical with other E. multilocularis sequences deposited under accession nos. AJ32907, AJ32908, AJ32909, and AJ32910 from Poland (7) and AY389984 from China (Yang JK et al., unpub. data), and differed considerably from the sequences of the most closely related species, E. granulosus. For phylogenetic analysis, the ND1 sequences of 7 E. multilocularis, 24 E. granulosus, 1 Taenia solium, 1 E. vogeli, and 1 E. oligarthrus isolates were included and MrBayes 3.04b (8) was used for the Bayesian estimation of phylogeny, applying the GTR+I+G substitution model that best fitted the data (determined with Modeltest 3.06) (9). Searches were conducted with

![Figure](image-url)
LETTERS

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References


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Instructions for Emerging Infectious Diseases Authors

Letters. Letters commenting on recent articles as well as letters reporting cases, outbreaks, or original research are welcome. Letters commenting on articles should contain no more than 300 words and 5 references; they are more likely to be published if submitted within 4 weeks of the original article’s publication. Letters reporting cases, outbreaks, or original research should contain no more than 800 words and 10 references. They may have 1 figure or table and should not be divided into sections. All letters should contain material not previously published and include a word count.

Influenza Virus Infection in Racing Greyhounds

To the Editor: Influenza is globally the most economically important respiratory disease in humans, pigs, horses, and fowl (1). Influenza virus is known for its continuous genetic and antigenic changes, which impeded effective influenza control (1,2). More importantly, emergence of a new subtype by genetic reassortment or interspecies transmission is of great concern for preventing influenza epidemics and pandemics (1). Recently, influenza outbreaks have occurred in species (feline and canine) that historically do not carry influenza virus (3,4), which alerted both regulatory and scientific communities to expansion of the host range of influenza virus. We report an outbreak of respiratory disease by influenza virus infection in Iowa racing greyhounds after influenza outbreaks in Florida in 2004.

In mid-April, an influx of racing greyhounds into Iowa greyhound tracks resulted in outbreaks of respiratory disease within the track compounds. The disease was characterized by rapid onset of fever and cough, rapid respiration, and hemorrhagic nasal discharge. The illness rate was almost 100% in both race-track compounds, although the death rate was <5%. Most affected dogs recovered, yet many died of hemorrhagic pneumonia. Therapeutic administration of broad-spectrum antimicrobial drugs reduced the severity of the disease but could not control it.

Tissue samples from 4 animals that died of severe pneumonia were submitted to the Iowa State University Veterinary Diagnostic Laboratory. The animals represented 2 different racing tracks located in eastern and western Iowa. On gross examination, lungs exhibited extensive red to red-