

Yong-Zhen Zhang,\* Cheng-Long Xiong,\* Dong-Lou Xiao,\* Ren-Jie Jiang,† Zhao-Xiao Wang,‡ Ling-Zhu Zhang,§ and Zhen F. Fu¶

\*Chinese Center for Disease Control and Prevention, Beijing, China; †Yancheng Municipal Center for Disease Control and Prevention, Yanchen, China; ‡Guizhou Center for Disease Control and Prevention, Guiyang, China; §Guangxi Center for Disease Control and Prevention, Nanning, China; and ¶University of Georgia, Athens, Georgia, USA

## References

1. Wang XJ, Huang JT. Epidemiology. In: Yu YX, editor. Rabies and rabies vaccine. Beijing: Chinese Medicine Technology Press; 2001. p. 127–44.
2. Gylys L, Chomel BB, Gardner IA. Epidemiological surveillance of rabies in Lithuania from 1986 to 1996. *Rev Sci Tech*. 1998;17:691–8.
3. Centers for Disease Control and Prevention. Human rabies prevention—United States, 1999; recommendations of the Advisory Committee on Immunization Practice (ACIP). *MMWR Recomm Rep*. 1999;48(RR-1):1–21.
4. Chhabra M, Ichhpujani RL, Tewari KN, Lal S. Human rabies in Delhi. *Indian J Pediatr*. 2004;71:217–20.
5. Hu JY, Xu TQ, Wu ZM, Si Y, Zhao LL, Shen RH, et al. A study on the epidemiological characteristics and the preventive measures of rabies in Shanghai [article in Chinese]. *Chin J Epidemiol*. 2001;22:11–13.
6. Du F, Huang MT, Liang FP, Luo HM, Wen JH, Chen DR, et al. Investigation on rabies virus in dogs in Guangdong [article in Chinese]. *Chin J Zoonoses*. 1992;8:39–41.
7. Fekadu M, Shaddock JH, Chandler FW, Baer GM. Rabies virus in the tonsils of a carrier dog. *Arch Virol*. 1983;78:37–47.

Address for correspondence: Yong-Zhen Zhang, Institute for Infectious Disease Control and Prevention, Chinese CDC, Liuzi 5, Changping 102206, Beijing, 102206, China; fax: 86-10-61739457; email: yongzhenzhang@sohu.com



## Resistant *Salmonella* Virchow in Quail Products

**To the Editor:** *Salmonella* spp. resistant to multiple antimicrobial agents have emerged worldwide in recent years, but clinical relevance varies with the agent to which resistance evolves. Fluoroquinolones are often the drug of choice to treat gastrointestinal infections in humans, and resistance to this class of antimicrobial agents is associated with increased illness and death (1). Resistance to fluoroquinolones has emerged worldwide during the last decade. *Salmonella* isolates resistant to oxyiminocephalosporins because they produce extended-spectrum  $\beta$ -lactamases (ESBLs) have emerged worldwide since 1992. This emergence has caused concern since cephalosporins are drugs of choice to treat salmonellosis in children, to whom fluoroquinolones must not be administered because of toxicity issues. In Denmark, the first ESBL-producing isolate of animal origin from a *Salmonella enterica* serovar Heidelberg isolated from a boar imported from Canada in 2003 was reported (2), but such isolates have not previously been reported in food products.

On October 15, 2003, the Danish Institute for Veterinary Research, the national reference laboratory, received 3 *Salmonella* isolates found in quails imported from France. *Salmonella* isolates found at any importer's laboratory in Denmark are submitted to the reference laboratory for further analyses. The quails were in the importer's storage room at the time of sampling; sampling was performed routinely by the importer's own laboratory. At the reference laboratory, the isolates were serotyped as *S. enterica* serovar Virchow and found resistant to ampicillin, ceftio-

fur, cephalothin, nalidixic acid, and tetracycline and with reduced susceptibility to ciprofloxacin (MICs >0.125  $\mu$ g/mL) (3). Polymerase chain reaction detection and sequencing (4) showed that the  $\beta$ -lactam resistance was mediated by *bla*<sub>CTX-M-9</sub>. Pulsed-field gel electrophoresis was performed by using *Xba*I and *Bln*I as restriction enzymes according to the PulseNet protocol (5), and all 3 isolates had the same profile.

On October 23, the importer was informed of the laboratory's findings and the increased risk associated with salmonella isolates simultaneously resistant to quinolones and cephalosporins. Based on this information, the importer withdrew the product from the supermarkets on October 24. Recently, *S. enterica* Virchow with *bla*<sub>CTX-M-9</sub> has also been reported in poultry, poultry products, and humans in France (6), as well as humans in Spain (7) and the United Kingdom (8). The isolates from France were also resistant to nalidixic acid; the isolates we have obtained from fresh quails imported from France are possibly related to these isolates.

The global food-products trade is expected to increase in the future. Thus, attempts to improve food safety must emphasize detection of antimicrobial drug-resistant bacteria in imported food products. Furthermore, international agreements that limit contamination with drug-resistant bacteria and resistance genes at the primary production site are necessary to ensure consumer safety (9). International agreements must be based on antimicrobial-resistance data and early reports of emerging problems. Recently, the World Health Organization (WHO) launched the Global Salm Surv program (10) to isolate and identify antimicrobial resistance to *Salmonella* globally.

Many national and international rules, as well as marketing and consumer factors, regulate the interna-

tional trade of food products and live animals. Large international corporations may also affect international trade. For example, McDonald's Corporation has issued a global policy for antimicrobial drug use in food animals that specifies requirements for their food product suppliers. Local groceries or supermarkets may also impose their own standards nationally. We are aware of only 1 product withdrawal related to antimicrobial resistance, the quail imported from France.

No international standards exist for managing food safety problems related to antimicrobial resistance. However, in 2003 the Food and Agriculture Organization of the United Nations, WHO, and the World Organisation for Animal Health jointly hosted a workshop with a panel of experts to scientifically assess resistance risks related to nonhuman use of antimicrobial drugs (9). The panel's purpose was also to provide recommendations to the Codex Alimentarius Commission for future risk management of antimicrobial drug resistance (9). Imposing restrictions on products with combinations of resistance, such as simultaneous resistance to quinolones and cephalosporins in *Salmonella*, as reported in this study, would be a good first step towards managing antimicrobial drug-resistance risks.

**Frank M. Aarestrup,\*  
Henrik Hasman,\*  
and Lars Bogø Jensen\***

\*Danish Institute for Food and Veterinary Research, Copenhagen V, Denmark

## References

1. Helms M, Simonsen J, Mølbak K. Quinolone resistance is associated with increased risk of invasive illness or death during infection with *Salmonella* serotype Typhimurium. *J Infect Dis.* 2004;190:1652-4.
2. Aarestrup FM, Hasman H, Olsen I, Sørensen G. International spread of *bla*(CMY-2)-mediated cephalosporin resistance in a multiresistant *Salmonella enterica* serovar Heidelberg isolate stemming from the importation of a boar by Denmark from Canada. *Antimicrob Agents Chemother.* 2004;48:1916-7.
3. Aarestrup FM, Lertworapreecha M, Evans MC, Bangtrakulnonth A, Chalermchaikit T, Hendriksen RS, et al. Antimicrobial susceptibility and occurrence of resistance genes among *Salmonella enterica* serovar Weltevreden from different countries. *J Antimicrob Chemother.* 2003;52:715-8.
4. Hasman H, Mevius D, Veldman K, Olesen I, Aarestrup FM.  $\beta$ -Lactamases among extended-spectrum beta-lactamase (ESBL) resistant *Salmonella* from poultry, poultry products and human patients in The Netherlands. *J Antimicrob Chemother.* 2005;56:115-21.
5. Ribot EM, Wierzbica RZ, Angulo FJ, Barrett TJ. *Salmonella enterica* serotype Typhimurium DT104 isolated from humans, United States, 1985, 1990, and 1995. *Emerg Infect Dis.* 2002;8:387-91.
6. Simarro E, Navarro F, Ruiz J, Miro E, Gomez J, Mirelis B. *Salmonella enterica* serovar Virchow with CTX-M-like beta-lactamase in Spain. *J Clin Microbiol.* 2000;38:4676-8.
7. Weill FX, Lailler R, Praud K, Kerouanton A, Fabre L, Brisabois A, et al. Emergence of extended-spectrum-beta-lactamase (CTX-M-9)-producing multiresistant strains of *Salmonella enterica* serotype Virchow in poultry and humans in France. *J Clin Microbiol.* 2004;42:5767-73.
8. Batchelor M, Hopkins K, Threlfall EJ, Clifton-Hadley FA, Stallwood AD, Davies RH, et al. *bla*(CTX-M) genes in clinical *Salmonella* isolates recovered from humans in England and Wales from 1992 to 2003. *Antimicrob Agents Chemother.* 2005;49:1319-22.
9. World Health Organization. Geneva: 1st Joint FAO/OIE/WHO expert workshop on non-human antimicrobial usage and antimicrobial resistance: scientific assessment. Geneva: The Organization; 2003 [cited 1 Mar 2005]. Available from [http://www.who.int/foodsafety/publications/micro/en/a\\_mr.pdf](http://www.who.int/foodsafety/publications/micro/en/a_mr.pdf)
10. World Health Organization. Global Salm Surv (GSS). Geneva: The Organization; 2004 [cited 1 Mar 2005]. Available from <http://www.who.int/salmsurv/en/>

Address for correspondence: Frank M. Aarestrup, Danish Institute for Food and Veterinary Research, Bülowsvej 27, DK-1790 Copenhagen V, Denmark; fax: 45-72-34-60-01; email: faa@dfvf.dk

## Vancomycin-resistant *Enterococcus faecium* Clone in Swine, Europe

**To the Editor:** The use of antimicrobial agents for growth promotion (AGP) in food-producing animals has been extensively debated because of the risk of establishing a reservoir of antimicrobial resistance genes or antimicrobial-resistant organisms of potential relevance for human health. This concern has motivated the progressive ban of the use of different AGP in the European Union, which began in 1997 with avoparcin and will end in 2006 (1). Worldwide trade of living animals for food production or breeding and of meat products enables multidrug-resistant pathogens to spread across national borders.

Intercontinental dissemination of antimicrobial-resistant bacteria associated with food animals has been described for particular clones such as *Salmonella enterica* Typhimurium DT104 or *Escherichia coli* O157:H7 and for transferable genetic elements such as the genomic island SG1 or the streptococcal plasmid pRE25 (2). Vancomycin-resistant enterococci (VRE) in European farms were initially associated with the intensive use of avoparcin; however, the persistence of VRE in food animal environments after years of avoparcin withdrawal indicates that coselection by further antimicrobial or other agents, increased fitness of strains, and mobile genetic elements cannot be ruled out (1-3).

A specific clone was recently detected among vancomycin-resistant *E. faecium* (VREF) isolated from different swine farms in Denmark and Switzerland and from a healthy Danish woman without antimicrobial drug exposure who ate pork, chicken, and beef (4,5). Since Portugal and