Transmission of New Bovine Prion to Mice

Thierry G.M. Baron, Anne-Gaëlle Biacabe, Anna Bencsik, and Jan P.M. Langeveld

We previously reported that cattle were affected by a prion disorder that differed from bovine spongiform encephalopathy (BSE) by showing distinct molecular features of disease-associated protease-resistant prion protein (PrPres). We show that intracerebral injection of such isolates into C57BL/6 mice produces a disease with preservation of PrPres molecular features distinct from BSE.

Until recently, transmissible spongiform encephalopathy (TSE) in cattle was believed to be caused by a single strain of infectious agent identified at the beginning of a foodborne epidemic of bovine spongiform encephalopathy (BSE). Characterization of the infectious agent associated with BSE showed unique features. These include defined incubation periods and distribution of brain lesions after transmission to wild-type mice, not only directly from cattle, but also after natural or experimentally induced cross-species transmission (1,2). The uniform features of the disease in cattle have also been shown by analysis of the distribution of neurodegenerative brain lesions at different places during the BSE epidemic (3,4).

Western blot analyses of protease-resistant prion protein (PrPres) accumulating in the brains of animals and humans with BSE have demonstrated specific molecular features. These include a low molecular mass of unglycosylated PrPres with high proportions of diglycosylated PrPres (5,6). However, recent studies reported cases of prion abnormalities in cattle with different PrPres features (7,8). Three cattle isolates from France have been reported, characterized by a higher apparent molecular mass of unglycosylated PrPres (H-type isolates) and decreased levels of diglycosylated PrPres when compared with BSE isolates (7). In addition, only PrPres from H-type isolates were labeled by monoclonal antibody P4 with defined PrPres N terminus epitope specificity, in contrast with PrPres from BSE isolates, which suggests a different cleavage by proteinase K of the disease-associated protein (9).

Twenty years after identification of the BSE epidemic in cattle, the origin of the BSE agent remains controversial (10,11). Researchers have often considered the most likely source to be a recycled infectious agent derived from prion-associated diseases found in other species, such as scrapie in sheep and goats. The recent description of unusual phenotypes of bovine prion diseases distinct from BSE is therefore puzzling (7). This situation has been reinforced by a second bovine amyloidotic spongiform encephalopathy found in cattle in Italy (8). However, whether such cases of bovine prion disorders were transmissible, and to what extent the infectious agent caused specific features distinct from BSE, have not been demonstrated.

The Study

Experimental groups of 20 (4- to 6-week old) C57BL/6 female mice (Charles River, L’Arbresle, France) were injected intracerebrally with 20 µL of 10% (weight/volume) homogenates per mouse prepared from brain stem samples of 3 cattle TSE isolates. Two of the isolates were characterized, as previously described (7), by a higher molecular mass of unglycosylated PrPres (H-type isolates) and labeling with P4 monoclonal antibody (Table). A typical cattle BSE isolate was also analyzed. Mice were housed and cared for in an appropriate biohazard prevention area (A3) according to European (directive 86/609/EEC) and French ethical committee (decree 87–848) guidelines. Mice were checked at least weekly for neurologic clinical signs and were killed when they exhibited signs of distress or confirmed evolution of clinical signs. The whole brain of every second mouse was frozen and stored at –80°C before Western blot analysis. The other brains were fixed in 4% paraformaldehyde for other histopathologic studies.

Frozen mouse brain tissues and fixed brain tissues were examined by Western blot analysis and immunohistochemical tests as previously described (12,13). PrPres extracted from half of whole brain was detected with monoclonal antibodies Sha31 (1:10 from TeSeE sheep/goat Western blot, Bio-Rad, Hercules, CA, USA) (14) and (340 ng/mL) (15). These antibodies are directed against the 144-WEDRYRE-151 and 88-WGQGG-92 murine amino acid PrP sequences, respectively. Antibody 12B2, which has an N-terminal specificity similar to that of monoclonal antibody P4, shows poor binding to BSE-derived PrPres, but unlike P4, binds with high affinity to prion protein from most mammalian species, including mice and cattle. Bound antibodies were detected by using enhanced enzymatic chemiluminescence (Amersham, Little Chalfont, UK) or Supersignal (Pierce, Rockford, IL, USA) and visualized either on film (Biomax, Eastman Kodak, Rochester, NY, USA) or directly in an image analysis system (Versadoc, Bio-Rad). Molecular masses of PrPres glycoforms were determined as the average of the center positions of the bands from at least 3 repeated electrophoretic
procedures, as measured by comparison with a biotinylated marker (B2787, Sigma, Saint Louis, MO, USA) included on each gel. Immunologic reactivities of antibodies 12B2 and Sha31 were compared in Western blots run in parallel with the same samples with both antibodies.

After intracerebral injection of cattle brain samples into C57BL/6 mice, disease was observed in mice with the 2 H-type isolates, as well as with the BSE sample. Survival periods of mice and results of PrPres detection among mice analyzed by Western blot are shown in the Table.

Western blot analysis of PrPres from H-type–infected mouse brains in comparison with BSE-infected mice is shown in Figure 1. All positive mice in the same experimental group showed the same Western blot pattern. This pattern showed higher molecular mass PrPres glycoforms in mice infected with H-type isolates than in mice infected with a typical BSE agent (1.1- to 1.5-Da difference in the unglycosylated PrPres (Figure 1A). Studies of PrPres protease cleavage showed that only the PrPres of mice infected with H-type isolates was recognized by antibody 12B2 (Figure 1B). This finding is in contrast to the result obtained with monoclonal antibody Sha31 directed against an epitope in the central region of the protein, which showed that the 12B2 epitope was preserved in H-type–infected mice. Thus, the molecular features of H-type cattle isolates, which are distinct from those of the BSE agent, were maintained after development of disease in mice.

Histopathologic analysis showed vacuolar lesions in the thalamus (Figure 2A) that were absent from the hypothalamus, cochlear nucleus, and superior collicules. These 3 neuroanatomic sites were severely affected in C57BL/6 mouse brain after primary passage of the BSE agent, as we and others have reported (1). Abnormal PrP was detected only in amyloid plaques (Figure 2B), in contrast to what was reported after BSE transmission in C57BL/6 mice (1).

Conclusions

Our data show that the recently identified bovine H-type isolates involve an infectious agent that can induce development of a disease across a species barrier, while maintaining the specific associated PrPres molecular signature. This evidence in favor of a new bovine prion strain in cattle suggests that BSE is not the only transmissible prion disease in cattle. The origin of such cases has not been determined (7). These cases suggest either the existence of alternative origins of such diseases in cattle or phenotypic changes of PrPres after infection with the BSE agent. However, based on analysis of molecular features of prion diseases in cattle, this situation is similar to that in humans (5), in which different subtypes of sporadic Creutzfeldt-Jakob disease agents are found.

### Table. Cattle sources of transmissible spongiform encephalopathy (TSE) used for experimental infections of C57BL/6 mice and transmission results

<table>
<thead>
<tr>
<th>Cattle TSE isolate</th>
<th>Age, y</th>
<th>Breed</th>
<th>Molecular type</th>
<th>Survival periods (d) in C57BL/6 mice (mean ± SD)</th>
<th>Western blot results†</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>8</td>
<td>Charolais</td>
<td>H</td>
<td>702 ± 117</td>
<td>8/9</td>
</tr>
<tr>
<td>2</td>
<td>12</td>
<td>Crossbreed</td>
<td>H</td>
<td>652 ± 85</td>
<td>10/10</td>
</tr>
<tr>
<td>3</td>
<td>4</td>
<td>Prim’Holstein</td>
<td>Typical</td>
<td>511± 89</td>
<td>8/9</td>
</tr>
</tbody>
</table>

*SD, standard deviation. †No. mice positive for disease-associated prion protein/no. mice analyzed.
Acknowledgments

We thank Jérémy Verchère and Dominique Canal for excellent technical assistance, Emilie Antier and Clément Lavigne for performing animal experiments, and Karel Riepema, Esther de Jong, and Jorg Jacobs for production and characterization of monoclonal antibody 12B2.

This study was supported by the Agence Française de Sécurité Sanitaire des Aliments, the Neuroprion Network of Excellence (FOOD-CT-2004-506579) (EUROSTRAINS project), the Dutch Ministry of Agriculture, Environmental Management and Food (8041869000), and NeuroPrion (FOOD-CT-2004-506579)(STOPPrions project).

Dr Baron is head of the Unité Agents Transmissibles Non Conventionnels, Agence Française de Sécurité Sanitaire des Aliments, in Lyon. His research focuses on diagnosis of prion diseases of ruminants and characterization of the disease-associated prion protein and infectious agents, with particular emphasis on atypical forms of these diseases.

References


Address for correspondence: Thierry G.M. Baron, Unité Agents Transmissibles Non Conventionnels, Agence Française de Sécurité Sanitaire des Aliments, 31 Ave Tony Garnier, 69364 Lyon CEDEX 07, France; email: t.baron@lyon.afssa.fr