Variant Creutzfeldt-Jakob disease (vCJD) may be transmissible by blood. To prevent secondary transmission through blood components, several countries have started to exclude as donors persons who have received a blood transfusion. We investigated the effectiveness of this measure by using a dynamic age-structured model. It is the first such model based on epidemiologic data: 1) blood donor activities, 2) a case-control study on CJD, 3) age distribution of recipients, and 4) death of recipients of blood transfusions. The model predicts that an infection like vCJD, which has been introduced into the population by the alimentary route, could not become endemic by transfusion alone and that only <1% of cases would be avoided by excluding from blood donation those persons who have received a transfusion.

Recent studies of variant Creutzfeldt-Jakob disease (vCJD) indicate that this disease is transmissible by blood. One case of probable transfusion-transmitted vCJD infection has been reported, and 1 case of subclinical infection has been detected (1,2). On February 9, 2006, a third case was announced by the UK Health Protection Agency (www.hpa.org.uk/hpa/news/articles/press_releases/2006/060209_cjd.htm). Each of the 3 patients had received a blood transfusion from a donor who subsequently developed clinical vCJD, which indicates that transfusion caused the infection. However, a policy to exclude potential donors who had received a transfusion would not have prevented at least the first 2 cases because the corresponding donors had not received any blood transfusion. Diagnostic tools to detect prions in blood are under development (3), but no routine test for the presence of the infectious agents of vCJD is available. Therefore, the questions arise as to whether an infection like vCJD could become endemic through blood donation alone and to what extent exclusion of potential donors with a history of transfusion would influence the transmission of such an infection (i.e., how many deaths due to the infection could be prevented?). The following mathematical model is the first to address these questions on the basis of epidemiologic data and realistic and epidemiologically justified assumptions.

Methods

Model Structure

Figure 1A shows the transitions of a person through the basic states of potential donor activities and receipt of blood transfusion. After birth a person is in the state of not having received any transfusion and not yet being an active donor (S00). The first index refers to the person’s state as a transfusion recipient; the second index, to the person’s status as a donor. Persons in state S00 can change to state S01 by becoming a donor or to states S100 or S101 by receiving a blood transfusion. The third index indicates whether a person with a transfusion history can actually be identified and excluded from donating blood (deferred) (index 1) or not (index 0). The states S111 and S110 can be reached by either transfusion recipients who start donating blood or active donors who receive a blood transfusion. Blood donors who become inactive are transferred into the states of ex-donors S02 and S12, depending on their transfusion history. Ex-donors can also become transfusion recipients; i.e., they are transferred from S02 to S12. Donor exclusion transfers a certain proportion of transfusion recipients into
the state of ex-donors. For all susceptible states, Figure 1B shows the transitions to the corresponding infected states. Table 1 provides a list of all input parameters together with descriptions and sources. The details of the model with all the numerical parameter estimates and the equations are given in the online Appendix (available from www.cdc.gov/ncidod/EID/13/1/89-app.htm). The computer program is available upon request. This article summarizes the major features of the model, the data sources, and the estimation of the model parameters.

Demography

To simplify the model, we did not attempt to describe the demographics of the population during the next 150 years. Doing so would involve predicting changes in rates of birth, death, and immigration. It is assumed that in the absence of infection, the population is demographically stationary. We assumed a constant inflow of newborns and an age-specific death rate. The latter was estimated as a weighted mean of the age-specific female and male death rates. Because this study was initiated in Germany, we used the corresponding demographic data. To start the simulation in a demographically stationary state, the model was run for 100 years without infection. Thus, the age distribution of the population was identical to the life table of Germany 2002/2004 averaged over both sexes (www.destatis.de/download/d/bevoe/sterbet04.xls).

Modeling Blood Donors

Blood donors in Germany are >18 and <68 years of age. The rates for becoming a new donor and terminating the period as an active donor are age dependent. The corresponding parameters were estimated by using data from 262,071 donors registered with the German Red Cross.

<table>
<thead>
<tr>
<th>Table 1. Summary of input parameters for the model*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Parameters</td>
</tr>
<tr>
<td>Age-specific mortality rates</td>
</tr>
<tr>
<td>Donor recruitment</td>
</tr>
<tr>
<td>Proportion of donors</td>
</tr>
<tr>
<td>Duration as active donor</td>
</tr>
<tr>
<td>Risk of receiving transfusions</td>
</tr>
<tr>
<td>Transfusion-associated risk for death</td>
</tr>
<tr>
<td>Alimentary infection</td>
</tr>
<tr>
<td>Incubation period†</td>
</tr>
<tr>
<td>Donor exclusion</td>
</tr>
</tbody>
</table>

*DRK, German Red Cross; SD, standard deviation; vCJD, variant Creutzfeldt-Jakob disease.
†Time between infection and death, i.e., duration of infection.
(DRK) Blood Service West in Hagen, Germany, including age, sex, age at first donation, number of donations, and date of last donation.

The age-specific prevalence of active donors peaks at ≈24 years of age and subsequently declines monotonically to zero by age 68. The overall prevalence in the population is 3%, i.e., 2.4 million donors in a population of ≈80 million.

Modeling Transfusion Recipients

The model takes into account that persons may receive >1 transfusion throughout their lifetime, but it does not track the number of transfusions received per person. Persons with ≥1 transfusion continue to be at risk for infection from further transfusions. The age-specific risk of receiving a transfusion was estimated from data for all patients hospitalized at the University Hospital in Essen during March 2003. Of 4,867 patients, 1,343 (27.6%) received ≥1 transfusion. The number of persons receiving a blood transfusion in each 5-year age group was divided by the corresponding number of persons in the general population. The observed rates were fitted with a simple model that assumes initially an exponential decline and subsequently a unimodal peak, which is proportional to the density function of the normal distribution. These age-specific ratios were properly scaled to balance the yearly number of transfusions per capita. To limit the complexity of the model, we did not take into account persons in subgroups, such as those with hemophilia, who obtain blood products from pools of donors. Because for medical reasons these subgroups are excluded from donating blood, they cannot contribute to persistence of the infection.

Independence of Receiving and Donating Blood

The events of receiving a blood transfusion and of donating blood are assumed to be independent of each other. This assumption is supported by the results of a case-control study of potential risk factors for CJD, which was coordinated by the Clinical Surveillance Centre for CJD, Department of Neurology in Göttingen, Germany (7). Table 2 shows the joint distribution for the control group of having received and donated blood. According to the Fisher exact test, the p value for the hypothesis of no association is 0.43.

Heterogeneity in the risk of receiving a blood transfusion is modeled by the assumption that only a proportion of the population are at risk, whereas the remaining proportion never receives a transfusion. This assumption was introduced to be consistent with data from the case-control study, in which ≈18% of the population reported having ever received a blood transfusion. Without this assumption, the model would predict that eventually 100% of a cohort would receive a blood transfusion because the average age annual risk of receiving a blood transfusion is about 5%, i.e., ≈4 million in a population of 80 million.

Modeling Transfusion-associated Death Rates

The transfusion-associated death rate has been described in detail by Wallis et al. (4). A good fit to the data assumes that at all ages a certain proportion of transfusion recipients have a higher rate of dying and the remaining proportion has a survival rate that corresponds to that of persons of the same age group in the general population. This age-dependent proportion of transfusion recipients with an increased risk for death is described by a generalized logistic function with a positive value at birth and an asymptote <100% for old age. The transfusion-associated death rate increases linearly with age. The increased death rate appears to be concentrated in the first 2 years after a transfusion. Wallis et al. report that 2,888 patients were observed as long as 7.4 years after transfusions received in June 1994 (4). The sex-specific rates were averaged for the simulation model.

Modeling the Infection

Usually the incubation period refers to the time between the infection and disease. In the context of CJD, however, disease can refer to onset, diagnosis, or death. Like Bacchetti, we also focused on death rates (8–10). The incubation period is assumed to be gamma distributed with a mean duration of 16 years and a standard deviation of 4 years, which conforms to estimates of Valleron et al. and Ghani et al. (5,6). Because of great uncertainty about the length of the incubation time, we also considered a much higher value of 50 years in the absence of the competing risk for death. The coefficient of variation is assumed to be the same, such that the standard deviation is 12.5 years. Because of competing risks, the actual sojourn in the incubation period is 15.3 for an incubation period of 16 years and 34.0 years for an incubation period of 50 years. The proportions of infected persons who would die with disease symptoms are 79% and 37% for the incubation periods of 16 and 50 years, respectively. This means that for an incubation time of 50 years, nearly two thirds would die without disease symptoms. Hereafter we refer to these values of 15 and 50 years as short and long incubation periods.

Table 2. Joint distribution of transfusion history and blood donation

<table>
<thead>
<tr>
<th>Received blood</th>
<th>Donated blood, no. observed (no. expected if events are independent)</th>
<th>Total no. (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>No</td>
<td>No</td>
<td>401 (404)</td>
</tr>
<tr>
<td>Yes</td>
<td>Yes</td>
<td>93 (90)</td>
</tr>
<tr>
<td>Total no. (%)</td>
<td>494 (80)</td>
<td>123 (20)</td>
</tr>
</tbody>
</table>
We distinguish between 2 modes of transmission. Initially, the infection is introduced into the population by the alimentary route. In the United Kingdom the number of infected animals entering the food supply peaked in 1989; most were concentrated within a period of 10 years (11), which we take as the assumed period of alimentary infection. After this period, this mode of transmission was interrupted so that further transmissions are possible only through blood transfusions.

A study to detect the presence of abnormal prion protein in appendix and tonsil tissues has suggested a prevalence of 235 infections per million in the United Kingdom (12). We arbitrarily assumed the prevalence of infections in Germany to be ≈1 order of magnitude lower, yielding a cumulative incidence of 25 per million, which was the value used for the simulations.

We made 2 contrasting assumptions about the infectivity of blood preparations and evaluated the results of these 2 simulations: each transfusion (100% infectivity) or no blood transfusion (0% infectivity) from an infected donor leads to infection of the recipient. In the model the infection probability (probability of receiving blood from an infected donor) is proportional to the proportion of infected donors among all donors. Thus, we can calculate the number of infections from blood transfusions compared with the number of infections from alimentary transmission alone.

Modeling Donor Exclusion

The model distinguishes between persons with and without transfusion history, termed recipients and nonrecipients; these terms are applied to persons whether they have or have not donated blood. The model allows recipients to be excluded from donating blood. In modeling the exclusion of recipients, we took into account that this measure may be imperfect and that a certain proportion of recipients may not be excluded.

Results

For the parameter estimates obtained from the sources described above, the infection cannot become endemic (Figure 2). If we assume no further spread through blood transfusions after 10 years of infections by the alimentary route, the maximum prevalence reached is ≈1,860 (1,434 for nonrecipients plus 426 for recipients) because some of the infected persons die of other causes during the incubation period. If transmission is assumed to be possible through blood transfusions (100% infectivity), then the maximum prevalence among recipients is increased by ≈78 infections after 4 more years for the short incubation period and by 193 infections after 23 more years for the long incubation period.

We assumed that donor exclusion is implemented immediately at the beginning of the alimentary infection risk period, which reduced the original number of 2.55 million donors by ≈20% to a value of 2.05 million donors. Because the model does not account for the stock of blood donations, this reduction in the number of donors must be compensated for with an increased rate of donations per donor to satisfy the demand; i.e., the average number of donations would have to increase from 1.6 to 2 per donor per year. Figure 2A shows that donor exclusion has almost no effect when the incubation period is assumed to be 16 years. The absolute prevalence (i.e., the actual number of infected persons) differs at most by 9. For a long incubation, differences are visible (59 persons at most) but small in view of the long time intervals and the size of the total

Figure 2. Absolute prevalence of infection for an incubation period of 16 (A) and 50 (B) years, for nonrecipients of blood transfusion (solid, black), recipients under the assumption of no infectivity (dashed, gray), of 100% infectivity without donor exclusion (dotted, black), and 100% infectivity with donor exclusion (solid, gray). The prevalence declines after the alimentary route of transmission is interrupted, i.e., after 10 years. Prevalence differs only slightly if the infection probability of a transfusion from an infected donor is increased from 0% to 100%. Donor exclusion produces negligible reductions.
population (Figure 2B). The reason for these small differences is described below.

The cumulative numbers of deaths from the infection are given in Table 3. The numbers are considerably smaller for the long than for the short incubation period because a long incubation period implies more deaths from other causes. The numbers are given separately for cases in patients with and without a history of blood transfusion. The route of infection for nonrecipients is alimentary only, whereas the route of infection for recipients is unclear. If we compare the simulations at 100% and 0% infectivity of blood transfusions, we observe 172 and 224 additional cases for the short and the long incubation periods, respectively. These numbers represent 11% of 1,557 and 31% of 725 cases, which would be expected for 0% infectivity for the short and long incubations periods, respectively. For the short incubation period we expect a higher absolute number of alimentary cases but a smaller proportion of transfusion cases than for the long incubation period. The exclusion of donors would prevent only 15 and 50 cases, i.e., -15 (0.9%) of 1,729 and 50 (5%) of 949, respectively, at the end of the epidemic. The epidemic lasts for ≈50 or ≈150 years for the short and the long incubation periods, respectively.

The predicted yearly incidence of deaths due to vCJD, separated by transfusion history, is shown in Figure 3. The yearly peak incidence of total deaths would be 128 and 29 for the short and the long incubation periods at 23 and 51 years after the beginning of the epidemic, respectively. For 0% infectivity the peak incidence would be only 5 and 3 cases less for the short and long incubation periods, respectively, which implies that the exclusion of donors with a transfusion history does not effectively prevent infection.

Figure 4 shows the predicted yearly incidence of deaths according to the route of infection. The time lags between the peaks of deaths due to alimentary infection and due to transfusion clearly differ and are 9 and 20 years for short and long incubation periods, respectively.

Finally, we considered the absolute prevalence of infected donors according to their history of blood transfusion (Figure 5). Most infected donors do not have a transfusion history, which explains the negligible effect of a policy excluding transfusion recipients from donation.

To determine whether the same model could also predict transition into a positive endemic equilibrium of the infection, we made the unrealistic assumptions that the rates of donor recruitment and donor loss are constant between the ages of 18 and 67 and that the rate of receiving a blood transfusion is constant throughout life. Then the model showed an extremely long time (>2,000 years).

<table>
<thead>
<tr>
<th>Table 3. Cumulative numbers of deaths from variant Creutzfeldt-Jakob disease at the end of the epidemic</th>
</tr>
</thead>
<tbody>
<tr>
<td>Incubation period</td>
</tr>
<tr>
<td>-------------------</td>
</tr>
<tr>
<td>Short</td>
</tr>
<tr>
<td></td>
</tr>
<tr>
<td></td>
</tr>
<tr>
<td>Long</td>
</tr>
<tr>
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</table>
before positive equilibrium would be reached (results not shown).

**Discussion**

Our model is the first attempt to describe in a realistic way the transmission of infections through blood transfusions. In 1994, Velasco-Hernández proposed a model for the spread of Chagas disease by vectors and blood transfusion (13). His model was used by Roberts and Heesterbeek to introduce their new concept to estimate the effort to eradicate an infectious disease (14). Huang and Villasana included transmission through blood transfusion in an AIDS model (15). All these models have in common what Inaba and Sekine state about their extension of Velasco-Hernández’s Chagas model: “...here we assume that blood donors are randomly chosen from the total population, and so there is no screening and the recipients of blood donations are donating blood themselves at the same rate as anybody else. This is an unrealistic assumption, but we will use it.” (16). These models implicitly describe transmission through blood transfusion exactly like person-to-person transmission by droplet infections.

The key innovation in our model is the simultaneous incorporation of 6 functions that all depend explicitly on the age of a person: 1) natural death rate, 2) rate of receiving a blood transfusion, 3) rates of donor recruitment, 4) donor loss, 5) death rate associated with transfusions, and 6) proportion of transfusion recipients at increased risk for death. The age-dependent effects of these processes cannot be ignored. Peak ages of donor activity (=22 years) and of receiving a blood transfusion (=70 years) are quite distinct and 50 years apart. This age pattern does not favor the spread of infection by blood transfusion. Another factor that acts against the infection becoming endemic is the transfusion-associated death rate. The good quality of the follow-up data of nearly 3,000 patients helped to incorporate realistic assumptions about the survival probabilities of transfusion recipients (4). The only data available about the joint distribution of blood donor activity and history of a blood transfusion was the CJD case-control study performed in Göttingen, Germany (7).

![Figure 4. Yearly incidence of deaths caused by alimentary transmission (solid line) and by blood transfusion (dashed line). The 2 peaks differ by 9 and 20 years, depending on the incubation period: 16 (A) and 50 (B) years, respectively.](image)

![Figure 5. Absolute prevalence of infected donors for an incubation period of 16 (A) and 50 (B) years. The solid black curves show the infected donors without transfusion history. These curves are identical for 0% and 100% infectivity and are independent of donor exclusion. The gray curves show infected donors with transfusion history for 100% (solid) and 0% (dashed) infectivity, respectively, without donor exclusion. The dotted black curves show the effect of donor exclusion starting at the beginning of alimentary risk. Most infected donors have no transfusion history and cannot, therefore, be excluded from blood donation.](image)
The length of the incubation period plays a major role in transmission dynamics and hence was subject to a sensitivity analysis. The model does not account for possible changes of infectivity during the incubation period. The model represents a worst-case scenario because it assumes 100% infectivity throughout the period of infection. Even under this extreme assumption, donor exclusion can prevent only 0.9% (or 5%) of the expected deaths, assuming the incubation period has a mean duration of 16 (or 50) years. The main explanation for this surprising result is that most infected donors have been infected by the alimentary route and never received any blood transfusion and, therefore, are not eligible for donor exclusion.

The present simulations have arbitrarily assumed a cumulative incidence of alimentary infection, about 25 per million (2,000 per 80 million). With pessimistic assumptions, the model predicts either 19.5 deaths per million for the short incubation period or 9 deaths per million for the long incubation period in the absence of spread through blood transfusion. This corresponds to at least 9 (36%) of 25 deaths attributable to the infection, which is ≈2 orders of magnitude higher than expected for vCJD in the United Kingdom. As of July 2006, the number of vCJD cases in the United Kingdom was 160. If we assume that the total number of cases will be 200, then our assumption corresponds to about 3.3 cases per million. Thus, at most, 1.4% of infected persons would die from the infection (unless a second wave of vCJD cases with a long incubation period occurs). According to our model, 0.9% of the deaths could be prevented by donor exclusion under the assumption of the short incubation period. In absolute numbers this would be ≈2 cases.

In France, the total number of vCJD cases recorded through July 2006 is 18. Even under the assumption that this number represents only 35% of the total number of cases (17), the absolute expected number of prevented cases would be <1. In 1998, France decided to exclude donors with a transfusion history, primarily to reduce the spread of viruses. The present model could be modified to assess the effectiveness of excluding donors with transfusion history for preventing emerging infections with different modes of transmission and additional epidemiologic states, e.g., latent or immune.

Our worst-case scenario assumptions of the epidemiology might seem similar to the situation in the United Kingdom. In Germany, no case of vCJD has been reported, which indicates that the expected number of cases in Germany is at least 2 orders of magnitude less than that in the United Kingdom. This latter aspect was considered in the interpretation of our model by a working group commissioned by the German Federal Minister of Health, which recommended in April 2006 that persons with a transfusion history not be excluded from donating blood (18). Our analysis enables different countries to perform their own risk assessment and choose a strategy according to the absolute number of cases observed or expected.

The German CID Surveillance study was supported by a grant from the German Ministry of Health (Az 325–4471–02/15 to Inga Zerr and H. A. Kretzschmar). Helpful discussions about previous versions of the model took place with the Working Group Overall Blood Supply Strategy with regard to vCJD, Germany (Chairman R. Seitz).

Dr Dietz is head of the Department of Medical Biometry at the University of Tübingen, Germany. His main interest is the application of mathematical models in the field of infectious diseases, in particular malaria and other parasitic diseases.

References


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Blood Transfusion and Spread of Variant Creutzfeldt-Jakob Disease

Klaus Dietz, Günter Raddatz, Jonathan Wallis, Norbert Müller, Inga Zerr, Hans-Peter Duerr, Hans Lefèvre, Erhard Seifried, Johannes Löwer

Technical Appendix

Modelling the age-specific mortality:

These death rates are described as a sum of three parametric functions which refer to (i) infancy and early childhood, (ii) late childhood and early adulthood, and (iii) the remaining life (a function which corresponds to a Gompertz model of aging with exponentially increasing death rate) (1). This idea of expressing the mortality throughout the whole life analytically is well-known since more than 100 years (Figure S 1) (2).

Figure S 1: Age-specific death rates in the general population. Dots: observed values; line: fitted death rates according to a parametric model involving three functions as described in the sections about demography. Beyond the age of 40 years the observations are well described by a Gompertz model which predicts that the death rates increase exponentially corresponding to a straight line on the logarithmic scale which is used here. The death rates for males and females are averaged in the present model.
The death rate initially declines very fast and reaches a minimum around the age of 10 years. Afterwards there is a unimodal peak which is well described by a lognormal distribution with a peak around 21 years. The corresponding life table, i.e. the probability of surviving a given age yields a life expectancy of 79 years (Figure S 2).

![Figure S 2: Life table for the stationary population used in the present model. This corresponds to the age-distribution in the absence of the infection](image)

The three functions $\mu_k$ describe the age-specific mortality during infancy and early childhood ($k=0$), during late childhood and early adulthood ($k=1$) and during late adulthood ($k=2$).

$$
\mu_0(a) = \mu_0 + \mu_1(a) + \mu_2(a)
$$

$$
\mu_0(a) = \exp(\alpha_0)\beta_0\delta a^{\delta-1}\exp(-\beta_0 a^\delta)
$$

$$
\mu_1(a) = \frac{\beta_1}{\sqrt{2\pi}\sigma} \exp\left[-\frac{1}{2} \left(\frac{\log\frac{a}{A}}{\sigma^2}\right)^2\right]
$$

$$
\mu_2(a) = \beta_2 \exp(\beta_3 a)
$$

The following parameter values were fitted by nonlinear least squares regression to the age-specific mortality of Germany, where the data from both sexes were averaged using the age specific survival function as weights ([http://www.destatis.de/download/d/bevoe/sterbet04.xls](http://www.destatis.de/download/d/bevoe/sterbet04.xls)).
Each year a fixed number of susceptible newborns is added to the population, such that the total size of the population is about 80 million.

**Modelling donor activities**

The yearly rate of becoming an active donor (the donor recruitment rate) is denoted by $\rho(a)$. It is zero below 18 and above 67 years. The age-specific number of first time donors of the German Red Cross (DRK) Blood Service was divided by the corresponding age-specific number of individuals in the population as provided by the Federal Statistical Office (to obtain an age specific rate of becoming a new active donor. This rate has to be multiplied by an age-independent scaling factor which ensures that the calculated total number of active donors corresponds to the observed number of about 2.5 million. The logarithm of the empirical yearly rate is approximated by a polynomial of fifth degree with the following parameters:

$$
\rho(a) = 10^{(r_0 + r_1 a + r_2 a^2 + r_3 a^3 + r_4 a^4 + r_5 a^5)} \quad \text{for } 18 \leq a \leq 67
$$

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>$r_0$</td>
<td>-2.96</td>
</tr>
<tr>
<td>$r_1$</td>
<td>-0.0145</td>
</tr>
<tr>
<td>$r_2$</td>
<td>-0.0015</td>
</tr>
<tr>
<td>$r_3$</td>
<td>0.0000784</td>
</tr>
<tr>
<td>$r_4$</td>
<td>0.0000023</td>
</tr>
<tr>
<td>$r_5$</td>
<td>-0.00000236</td>
</tr>
</tbody>
</table>

The rate of terminating active donation is denoted by $\sigma(a)$. Depending on the age of recruitment we estimate the average time as active donor by fitting an exponential distribution to the age-distribution of active donors. For these age specific average
sojourn times we fit a polynomial of fourth degree. The inverse of these averages we take as the age-specific rate $\sigma(a)$:

$$\sigma(a) = \frac{1}{s_0 + s_1 a + s_2 (a - 42.5)^2 + s_3 (a - 42.5)^3 + s_4 (a - 42.5)^4}$$

with parameters

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>$s_0$</td>
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<tr>
<td>$s_1$</td>
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</tr>
<tr>
<td>$s_2$</td>
<td>-0.00254</td>
</tr>
<tr>
<td>$s_3$</td>
<td>0.000289</td>
</tr>
<tr>
<td>$s_4$</td>
<td>-0.000006</td>
</tr>
</tbody>
</table>

The donor recruitment rate is estimated from the age distribution of first-time donors in our sample (Figure S 3A) and from the age distribution of the population (Figure S 3B). The age-specific ratios of these frequencies are multiplied by a scaling factor in order to obtain a realistic size of the donor population. The recruitment rate is highest at the age of 18 years, declines by more than one order of magnitude up to the age of 28, remains almost constant up to the age of 50 and declines substantially in aged donors (Figure S 3C).
Figure S 3: Estimation of the rate of becoming an active donor. A) Age-distribution of first-time donors, B) Age-distribution of the population of Germany C) observed (dots) and fitted (line) logarithm of the age-specific rate to become an active donor.
Donor loss is estimated from the age distributions of active donors as a function of the youngest age as active donor. An exponential distribution is fitted to each of these distributions for all yearly age classes from 18 to 66 years (Figure S 4A provides an example). The corresponding mean durations are fitted by a polynomial (Figure S 4B).

![Graph A](image)

**Figure S 4A**: The observed distribution of the time since the first donation for 9539 donors who were 22 years old when they started to donate blood (black line). We fit an exponential distribution to this curve and obtain a mean duration of 13.6 years as active donors (grey line).

![Graph B](image)

**Figure S 4B**: A polynomial is fitted to these average durations. The inverse values of these durations are used as the age-specific transition rates to become an ex-donor.

The duration as active donor increases from 11 years in 18 year old donors to 13.7 years in 23 year old donors, and subsequently declines nearly linearly to zero until the maximum age of active donors.
Modelling the age-specific prevalence of active donors

The estimates of donor recruitment and loss of donors produces the age-specific prevalence of active donors (Figure S 5). The prevalence peaks at ≈24 years of age and subsequently declines monotonically to zero by age 67. The overall prevalence in the population is 3%, i.e., 2.4 million donors in a population of ≈80 million.

![Graph showing age-specific prevalence of active donors](image)

Figure S 5: Predicted age-specific prevalence of active blood donors, calculated on the basis of the parameter estimates of the rates of donor recruitment and donor loss.

Modelling the risk of receiving transfusions

The yearly age-specific risk of receiving a transfusion $\varepsilon_b(a)$ among the fraction $b_x$ of the population with a positive transfusion risk is estimated from the age-specific transfusion risk at the University Hospital Essen, Germany during one month. The function has a peak at birth which declines exponentially and another peak at about 65 years of age:

$$\varepsilon_b(a) = \mu_0 + \mu_1 \exp(-\mu_2 a) \frac{H_3}{\sigma} \exp \left[ -\frac{(a - \mu_4)^2}{2\sigma^2} \right].$$

A graphical representation is given by Figure S 6.
Figure S 6: The observed (dots) and fitted (line) age-specific transfusion risk.

The parameter values for the fitted function are given in the following Table:

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>$\mu_0$</td>
<td>0.0155</td>
</tr>
<tr>
<td>$\mu_1$</td>
<td>0.0412</td>
</tr>
<tr>
<td>$\mu_2$</td>
<td>0.259</td>
</tr>
<tr>
<td>$\mu_3$</td>
<td>1.133</td>
</tr>
<tr>
<td>$\sigma$</td>
<td>11.16</td>
</tr>
<tr>
<td>$\mu_4$</td>
<td>65.35</td>
</tr>
</tbody>
</table>

The cumulative transfusion risk is given by:

$$E_B(a) = \int_0^a \varepsilon_B(s) ds .$$

The following expression describes the age-specific risk of transfusion for the total population:

$$\varepsilon(a) = \frac{b\varepsilon_B(a) \exp(-E_B(a))}{1 - b\varepsilon(1 - \exp(-E_B(a)))},$$

which is obtained by calculating the hazard rate for the proportion of individuals without transfusion in the denominator of the previous expression.
Modelling the transfusion associated mortality

The proportion of transfusion recipients with increased mortality is described by the generalised logistic function:

\[ q(a) = p_0 + \frac{p_1 - p_0}{1 + \left( \frac{A_q}{a} \right)^c} \]

Sex-specific fits are presented in Figure S 7.

Figure S 7: Age-and sex-specific parameter values for the proportion \( q(a) \) of recipients with an increased risk of death (males: dotted in black, females: dashed in grey) and the linearly increasing death rates \( \beta(a) \) for those at increased risk of dying (males: solid-black, females: solid-grey).

Parameters for both sexes combined are fitted from original data (published in (3)), and are given in the following Table:

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>( p_0 )</td>
<td>0.170</td>
</tr>
<tr>
<td>( p_1 )</td>
<td>0.4824</td>
</tr>
<tr>
<td>( A_q )</td>
<td>42.1</td>
</tr>
<tr>
<td>( c )</td>
<td>11.8</td>
</tr>
</tbody>
</table>

The generalised logistic function predicts that the proportion of transfusion recipients with increased mortality increases from 17% at birth to about 48% in old age. The point of inflection is at the age of about 42 years. The high value of nearly 12 for the exponent \( c \) indicates a steep slope at the point of inflection. For those individuals with a high transfusion associated death rate this rate increases linearly with age:

\[ \beta(a) = \beta_1 + m_\beta a \]

with the following parameters
This means that at birth the life expectancy for those affected by an increased death rate is about 2.5 years. This value decreases to about 0.5 years for age 80 years.

The cumulative death rate is given by $B(a) = \beta_4 a + m_\beta \frac{a^2}{2}$.

The survival probability for a 3 year old girl is much closer to the survival of the general population compared to a 70 year old man (Figure S 8).

![Figure S 8: The predicted survival curves (grey) for two patients together with the expected survival without transfusion-associated risk (black). A) 3 year old girl, B) 70 year old man.](image-url)
Furthermore, the asymptote is approached more slowly for a 3 year old girl indicating a longer survival. The goodness of fit of the model has been evaluated by dividing the 1284 males and the 1605 females into deciles according to age such that there are about 130 and 160 individuals, respectively, in each age group (Figure S 9).

![Figure S 9: Goodness of model fit for the lowest (A) and the highest (B) sex-specific age deciles of survival after a transfusion together with the expected survival of those without transfusion. The lower curves are for male, the intermediate curves for females. Black: observed, grey: predicted. The upper dashed curves show the expected survival without transfusion (males: dashed in black, females: dotted in grey; in (A) both approximate the value of one and are almost identical). The goodness of fit of this model is further illustrated in Figure S 10:](image-url)
Figure S 10: Observed (grey) and predicted (black) sex-specific survival curves after a blood transfusion (all ages combined). The lower curves are for males; the intermediate curves are for females; the upper curves show the expected survival rates without transfusion (dashed for males, dotted for females; both are almost identical).

Since we model explicitly the transfusion associated death rates we have to decrease the general death rates according to the following expression in order to keep the total survival equal to the observations

$$\mu(a) = \mu_b(a) - \frac{b \times \beta(a) \int_0^a q(s) \epsilon_B(s) \exp\left( -E_B(s) - (B(a) - B(s)) \right) ds}{1 - b \int_0^a q(s) \epsilon_B(s) \exp\left( -E_B(s) \right) \left( 1 - \exp\left( -\left( B(a) - B(s) \right) \right) \right) ds}.$$  

**Modelling the infection**

The sojourn time in the infected stage before an individual dies due to the infection has a gamma distribution with shape parameter $n = 16$ and $\alpha = 0.0625$ per year which corresponds to an average duration of 16 years. This value is reduced to 0.02 to explore the sensitivity of the results with respect to this parameter. The density of the incubation time distribution is not very different from a lognormal distribution with the same mean and standard deviation (Figure S 11).
Figure S 11: Density of the gamma distributed incubation time (grey line) in comparison to a lognormal density with the same mean and variance (black line).

The risk of an infection via the alimentary route is given by the following parameters:

\[ \lambda(t) = \lambda_0 \quad \text{für } 0 \leq t < 10, \]
\[ \lambda(t) = 0 \quad \text{für } t \geq 10. \]

The probability that a transfusion from an infected donor is infective is denoted by the parameter \( f \) which is assigned to the values of zero and one.

**Modelling donor exclusion**

Donor exclusion is modelled by two parameters: coverage \( b \) and a time-dependent exclusion rate \( \gamma(t) \) which allows determining the “velocity” of donor exclusion. By means of the coverage parameter the model may take into account that a donor exclusion program may not succeed to exclude 100% of potential and active donors with transfusion history. The time-dependent parameter \( \gamma(t) \) allows determining the timing of a donor exclusion program in relation to the period during which there is an alimentary infection risk.
Model Equations

All variables $S(t,a)$ and $I(t,a)$ depend both on time $t$ and year $a$, both of which are updated every two months, i.e. we always follow a cohort by increasing simultaneously time and age by two months. All 26 variables are distinguished by appropriate indices of the numbers of susceptible ($S(t,a)$) and infective ($I(t,a)$) individuals.

Explanation of the indices of the 26 variables
- 1st index: without history of transfusion = 0 (non-recipients); with history transfusion = 1 (recipients);
- 2nd index: potential donor = 0; active donor = 1, past donor or excluded from donation = 2;
- 3rd index (for recipients, i.e. 1st index=1, and 2nd index <2): not deferrable=0, deferrable=1;
- 3rd index (for recipients, i.e. 1st index=1, and 2nd index =2): without transfusion associated risk of death = 0; with transfusion associated risk of death =1;
- 3rd index for individuals infected via the alimentary route (1st index = 0): number of the fictitious incubation state ($j=1, \ldots, n$);
- 4th index (for recipients, i.e. 1st index = 1, and 2nd index <2): without transfusion associated risk of death =0; with transfusion associated risk of death =1;
- 4th index (for recipients, i.e. 1st index = 1, and 2nd index =2) for infected individuals: number of the fictitious incubation state ($j=1, \ldots, n$);
- 5th index (for recipients, i.e. 1st index = 1, and 2nd index <2) for infected individuals: number of the fictitious incubation state ($j=1, \ldots, n$)
With this notation, the 26 variables represent proportions in the population with following attributes:

<table>
<thead>
<tr>
<th></th>
<th>1st index</th>
<th>2nd index</th>
<th>3rd index</th>
<th>4th index</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>S(_{00})</td>
<td>non-recipient</td>
<td>no donor</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>S(_{01})</td>
<td>non-recipient</td>
<td>donor</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>S(_{02})</td>
<td>non-recipient</td>
<td>ex-donor</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>S(_{1000})</td>
<td>recipient</td>
<td>no donor</td>
<td>deferrable</td>
</tr>
<tr>
<td>5</td>
<td>S(_{1001})</td>
<td>recipient</td>
<td>no donor</td>
<td>deferrable</td>
</tr>
<tr>
<td>6</td>
<td>S(_{1010})</td>
<td>recipient</td>
<td>no donor</td>
<td>not deferrable</td>
</tr>
<tr>
<td>7</td>
<td>S(_{1011})</td>
<td>recipient</td>
<td>no donor</td>
<td>not deferrable</td>
</tr>
<tr>
<td>8</td>
<td>S(_{1100})</td>
<td>recipient</td>
<td>donor</td>
<td>deferrable</td>
</tr>
<tr>
<td>9</td>
<td>S(_{1101})</td>
<td>recipient</td>
<td>donor</td>
<td>deferrable</td>
</tr>
<tr>
<td>10</td>
<td>S(_{1110})</td>
<td>recipient</td>
<td>donor</td>
<td>not deferrable</td>
</tr>
<tr>
<td>11</td>
<td>S(_{1111})</td>
<td>recipient</td>
<td>donor</td>
<td>not deferrable</td>
</tr>
<tr>
<td>12</td>
<td>S(_{120})</td>
<td>recipient</td>
<td>ex-donor</td>
<td>no extra mortality</td>
</tr>
<tr>
<td>13</td>
<td>S(_{121})</td>
<td>recipient</td>
<td>ex-donor</td>
<td>extra mortality</td>
</tr>
<tr>
<td>14</td>
<td>I(_{00j})</td>
<td>non-recipient</td>
<td>no donor</td>
<td></td>
</tr>
<tr>
<td>15</td>
<td>I(_{01j})</td>
<td>non-recipient</td>
<td>donor</td>
<td></td>
</tr>
<tr>
<td>16</td>
<td>I(_{02j})</td>
<td>non-recipient</td>
<td>ex-donor</td>
<td></td>
</tr>
<tr>
<td>17</td>
<td>I(_{1000j})</td>
<td>recipient</td>
<td>no donor</td>
<td>not deferrable</td>
</tr>
<tr>
<td>18</td>
<td>I(_{1001j})</td>
<td>recipient</td>
<td>no donor</td>
<td>not deferrable</td>
</tr>
<tr>
<td>19</td>
<td>I(_{1010j})</td>
<td>recipient</td>
<td>no donor</td>
<td>deferrable</td>
</tr>
<tr>
<td>20</td>
<td>I(_{1011j})</td>
<td>recipient</td>
<td>no donor</td>
<td>deferrable</td>
</tr>
<tr>
<td>21</td>
<td>I(_{1100j})</td>
<td>recipient</td>
<td>donor</td>
<td>not deferrable</td>
</tr>
<tr>
<td>22</td>
<td>I(_{1101j})</td>
<td>recipient</td>
<td>donor</td>
<td>not deferrable</td>
</tr>
<tr>
<td>23</td>
<td>I(_{1110j})</td>
<td>recipient</td>
<td>donor</td>
<td>deferrable</td>
</tr>
<tr>
<td>24</td>
<td>I(_{1111j})</td>
<td>recipient</td>
<td>donor</td>
<td>deferrable</td>
</tr>
<tr>
<td>25</td>
<td>I(_{120(j-1)})</td>
<td>recipient</td>
<td>ex-donor</td>
<td>no extra mortality</td>
</tr>
<tr>
<td>26</td>
<td>I(_{121j})</td>
<td>recipient</td>
<td>ex-donor</td>
<td>extra mortality</td>
</tr>
</tbody>
</table>
For the indices we use the so called “+”-Notation: if an index is replaced by a “+”, then this means that we have summed all variables with different indices at this position.

The model is illustrated with Figure S 12:

Figure S 12: States and transitions of the epidemiological model. For the transfusion recipients (all states for which the 1st index equals 1) the model distinguishes individuals with and without increased risk of dying. In the model equations this aspect is expressed with the last index of the variables. In this figure this aspect is not included. The death rates associated with each state are also not shown. Parameters: $\nu$: birth rate, $\lambda$: rate of infection during the period of alimentary transmission, $\varepsilon$: rate of receiving blood transfusions, $\rho$: donor recruitment rate, $\sigma$: donor loss rate, $\gamma$: rate of donor exclusion, $b$: proportion of deferrable donors with transfusion history, $p$: proportion of infected donors among all donors, $f$: probability that an infected blood transfusion leads to infection of the recipient.
**Model equations**

Since our step size is two months and some rates are high we use the exponential function in all equations for calculating how many individuals are still in the initial state after two months. The argument of the exponential function is the sum of all rates which describe a transition out of the corresponding variable. This approach avoids negative variables which would arise when more individuals are removed than were present at the beginning of the interval which could happen for large rates. E.g. in the second equation (for the active susceptible donors without transfusion history) we use the theory of competing risks to determine which proportion of the potential donors are added to this category: in the denominator we add all transition rates and in the numerator we select the relevant transition rate out of those from the numerator. The number of individuals in the previous state is multiplied by the probability to survive the previous year and the probability to make a transition which is the complement to the probability to make no transition.

The following expression is the time-dependent prevalence of infected active donors among all active donors:

$$p(t) = \frac{\sum_{a=18}^{67} I_{01}(a,t) + I_{11++}(a,t)}{\sum_{a=18}^{67} I_{01}(a,t) + I_{11++}(a,t) + S_{01}(a,t) + S_{11+}(a,t)}.$$  

If the duration of an infection is short, i.e. the parameter $\alpha$ is high, then it may happen that a newly infected individual at the end of the first iteration step is not only in the first fictitious state of the infection but already in the second state or even in a higher state or with a small probability the individual may even have died during the iteration step. The following expressions determine the probabilities according to a gamma-distribution of finding a newly infected individual in state $j = 1, \ldots, n$. The state $j = n+1$ corresponds to the death of the individual due to the infection:

$$q_j = \exp(-n\alpha) \frac{(n\alpha)^{j-1}}{(j-1)!}, j = 1, \ldots, n,$$

$$q_{n+1} = 1 - \sum_{j=1}^{n} q_j.$$
This material, provided by the authors as a supplement to Blood Transfusion and Spread of Variant Creutzfeldt-Jakob Disease, is not part of Emerging Infectious Diseases contents.

(1) $S_{00}(t + 1, a + 1) = S_{00}(t, a) \exp \left\{ - \frac{\theta(t) + \rho(a) + \epsilon(a) + \mu(a)}{\lambda(t) + \rho(a) + \epsilon(a)} \right\}$

(2) $S_{01}(t + 1, a + 1) = S_{01}(t, a) \exp \left\{ - \frac{\theta(t) + \rho(a) + \mu(a)}{\lambda(t) + \rho(a) + \epsilon(a)} \right\}$

(3) $S_{0a}(t + 1, a + 1) = S_{0a}(t, a) \exp \left\{ - \frac{\theta(t) + \epsilon(a) + \mu(a)}{\lambda(t) + \sigma(a) + \epsilon(a)} \right\}$

(4) $S_{100}(t + 1, a + 1) = S_{100}(t, a) \exp \left\{ - \frac{\theta(t) + f(t)\epsilon(a) + \mu(a)}{\lambda(t) + \rho(a) + \epsilon(a)} \right\}$

(5) $S_{101}(t + 1, a + 1) = S_{101}(t, a) \exp \left\{ - \frac{\theta(t) + f(t)\epsilon(a) + \gamma(t) + \mu(a)}{\lambda(t) + \rho(a) + \epsilon(a)} \right\}$

(6) $S_{110}(t + 1, a + 1) = S_{110}(t, a) \exp \left\{ - \frac{\theta(t) + f(t)\epsilon(a) + \rho(a) + \gamma(t) + \mu(a)}{\lambda(t) + \rho(a) + \epsilon(a)} \right\}$

(7) $S_{111}(t + 1, a + 1) = S_{111}(t, a) \exp \left\{ - \frac{\theta(t) + f(t)\epsilon(a) + \gamma(t) + \mu(a) + \beta(a)}{\lambda(t) + \rho(a) + \epsilon(a)} \right\}$

(8) $S_{100}(t + 1, a + 1) = S_{100}(t, a) \exp \left\{ - \frac{\theta(t) + \rho(a) + \mu(a)}{\lambda(t) + \rho(a) + f(t)\epsilon(a)} \right\}$

(9) $S_{101}(t + 1, a + 1) = S_{101}(t, a) \exp \left\{ - \frac{\theta(t) + \rho(a) + f(t)\epsilon(a) + \beta(a) + \mu(a)}{\lambda(t) + \rho(a) + f(t)\epsilon(a)} \right\}$
This material, provided by the authors as a supplement to Blood Transfusion and Spread of Variant Creutzfeldt-Jakob Disease, is not part of Emerging Infectious Diseases contents.

(10) \[ S_{110}(t + 1, a + 1) = S_{110}(t, a) \exp \left\{ - \left[ \lambda(t) + \sigma(a) + f(t) \varepsilon(a) + \gamma(t) + \mu(a) \right] \right\} + \frac{\rho(a)}{\lambda(t) + \rho(a) + f(t) \varepsilon(a) + \gamma(t)} S_{1010}(t, a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \rho(a) + f(t) \varepsilon(a) + \gamma(t) \right] \right\} \right\} + \frac{\varepsilon(a)(1 - f(t))b(1 - q(a))}{\lambda(t) + \sigma(a) + \varepsilon(a)} S_{01}(t, a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \sigma(a) + \varepsilon(a) \right] \right\} \right\} \]

(11) \[ S_{111}(t + 1, a + 1) = S_{111}(t, a) \exp \left\{ - \left[ \lambda(t) + \sigma(a) + f(t) \varepsilon(a) + \gamma(t) + \mu(a) + \beta(a) \right] \right\} + \frac{\rho(a)}{\lambda(t) + \rho(a) + f(t) \varepsilon(a) + \gamma(t)} S_{1110}(t, a) \exp(-\mu(a) - \beta(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \rho(a) + f(t) \varepsilon(a) + \gamma(t) \right] \right\} \right\} + \frac{\varepsilon(a)(1 - f(t))bq(a)}{\lambda(t) + \sigma(a) + \varepsilon(a)} S_{00}(t, a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \sigma(a) + \varepsilon(a) \right] \right\} \right\} \]

(12) \[ S_{120}(t + 1, a + 1) = S_{120}(t, a) \exp \left\{ - \left[ \lambda(t) + f(t) \varepsilon(a) + \mu(a) \right] \right\} + \frac{\varepsilon(a)(1 - f(t))(1 - q(a))}{\lambda(t) + \varepsilon(a)} S_{1201}(t, a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \varepsilon(a) \right] \right\} \right\} + \frac{\sigma(a)}{\lambda(t) + \sigma(a) + f(t) \varepsilon(a)} S_{1100}(t, a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \sigma(a) + f(t) \varepsilon(a) \right] \right\} \right\} + \frac{\sigma(a) + \gamma(t)}{\lambda(t) + \sigma(a) + f(t) \varepsilon(a) + \gamma(t)} S_{1110}(t, a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \sigma(a) + f(t) \varepsilon(a) + \gamma(t) \right] \right\} \right\} + \frac{\gamma(t)}{\lambda(t) + \rho(a) + f(t) \varepsilon(a) + \gamma(t)} S_{1101}(t, a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \rho(a) + f(t) \varepsilon(a) + \gamma(t) \right] \right\} \right\} \]

(13) \[ S_{121}(t + 1, a + 1) = S_{121}(t, a) \exp \left\{ - \left[ \lambda(t) + f(t) \varepsilon(a) + \mu(a) + \beta(a) \right] \right\} + \frac{\varepsilon(a)(1 - f(t))q(a)}{\lambda(t) + \varepsilon(a)} S_{1210}(t, a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \varepsilon(a) \right] \right\} \right\} + \frac{\sigma(a)}{\lambda(t) + \sigma(a) + f(t) \varepsilon(a)} S_{1101}(t, a) \exp(-\mu(a) - \beta(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \sigma(a) + f(t) \varepsilon(a) \right] \right\} \right\} + \frac{\sigma(a) + \gamma(t)}{\lambda(t) + \sigma(a) + f(t) \varepsilon(a) + \gamma(t)} S_{1111}(t, a) \exp(-\mu(a) - \beta(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \sigma(a) + f(t) \varepsilon(a) + \gamma(t) \right] \right\} \right\} + \frac{\gamma(t)}{\lambda(t) + \rho(a) + f(t) \varepsilon(a) + \gamma(t)} S_{1110}(t, a) \exp(-\mu(a) - \beta(a)) \left\{ 1 - \exp \left\{ - \left[ \lambda(t) + \rho(a) + f(t) \varepsilon(a) + \gamma(t) \right] \right\} \right\} \]
\begin{align}
(14) \quad I_{00j}(t+1,a+1) &= I_{00j}(t,a)\exp \left\{ \frac{n\alpha}{\rho(a)+\epsilon(a)+n\alpha} \right\} + \\
& \frac{\lambda(t)}{\lambda(t)+\rho(a)+\epsilon(a)} I_{00(j-1)}(t,a)\exp(-\mu(a))(1 - \exp \left\{ \frac{\lambda(t)+\rho(a)+\epsilon(a)}{\lambda(t)+\rho(a)+\epsilon(a)+\alpha} \right\}) q_j.
\end{align}

\begin{align}
(15) \quad I_{w,j}(t+1,a+1) &= I_{w,j}(t,a)\exp \left\{ \frac{n\alpha}{\sigma(a)+\epsilon(a)+n\alpha} \right\} + \\
& \frac{\rho(a)}{\sigma(a)+\epsilon(a)+n\alpha} I_{w(0),(j-1)}(t,a)\exp(-\mu(a))(1 - \exp \left\{ \frac{\lambda(t)+\rho(a)+\epsilon(a)}{\lambda(t)+\rho(a)+\epsilon(a)+\alpha} \right\}) q_j.
\end{align}

\begin{align}
(16) \quad I_{a,j}(t+1,a+1) &= I_{a,j}(t,a)\exp \left\{ \frac{n\alpha}{\sigma(a)+\epsilon(a)+n\alpha} \right\} + \\
& \frac{\lambda(t)}{\lambda(t)+\sigma(a)+\epsilon(a)} I_{a(0),(j-1)}(t,a)\exp(-\mu(a))(1 - \exp \left\{ \frac{\lambda(t)+\rho(a)+\epsilon(a)}{\lambda(t)+\rho(a)+\epsilon(a)+\alpha} \right\}) q_j.
\end{align}

\begin{align}
(17) \quad I_{1000j}(t+1,a+1) &= I_{1000j}(t,a)\exp \left\{ \frac{n\alpha}{\rho(a)+\mu(a)+n\alpha} \right\} + \\
& \frac{(1-b)fp(t)\sigma(a)(1-q(a))}{\lambda(t)+\rho(a)+\epsilon(a)} I_{1000(j-1)}(t,a)\exp(-\mu(a))(1 - \exp \left\{ \frac{\lambda(t)+\rho(a)+\epsilon(a)}{\lambda(t)+\rho(a)+\epsilon(a)+\alpha} \right\}) q_j + \\
& \frac{fp(t)\sigma(a)+\lambda(t)}{\lambda(t)+\rho(a)+fp(t)\sigma(a)} S_{1000}(t,a)\exp(-\mu(a))(1 - \exp \left\{ \frac{\lambda(t)+\rho(a)+fp(t)\sigma(a)}{\lambda(t)+\rho(a)+\epsilon(a)+\alpha} \right\}) q_j.
\end{align}

\begin{align}
(18) \quad I_{1001j}(t+1,a+1) &= I_{1001j}(t,a)\exp \left\{ \frac{n\alpha}{\rho(a)+\mu(a)+\beta(a)+n\alpha} \right\} + \\
& \frac{(1-b)fp(t)\sigma(a)q(a)}{\lambda(t)+\rho(a)+\epsilon(a)} S_{1001}(t,a)\exp(-\mu(a))(1 - \exp \left\{ \frac{\lambda(t)+\rho(a)+\epsilon(a)}{\lambda(t)+\rho(a)+\epsilon(a)+\alpha} \right\}) q_j + \\
& \frac{fp(t)\sigma(a)+\lambda(t)}{\lambda(t)+\rho(a)+fp(t)\sigma(a)} S_{1001}(t,a)\exp(-\mu(a))(1 - \exp \left\{ \frac{\lambda(t)+\rho(a)+fp(t)\sigma(a)}{\lambda(t)+\rho(a)+\epsilon(a)+\alpha} \right\}) q_j.
\end{align}
\[
\begin{align*}
(19) \quad & I_{1010+j}(t+1,a+1) = I_{1010+j}(t,a) \exp \left\{ -\left[ \rho(a) + \gamma(t) + \mu(a) + n \alpha \right] \right\} + \\
& \frac{n \alpha}{\rho(a) + \gamma(t) + n \alpha} I_{1010+j-1}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \rho(a) + \gamma(t) + n \alpha \right] \right\} \right\} + \\
& \frac{bfp(t)c(a)(1-q(a))}{\lambda(t) + \rho(a) + \epsilon(a)} S_{00}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \lambda(t) + \rho(a) + \epsilon(a) \right] \right\} \right\} q_j + \\
& \frac{fp(t)c(a) + \lambda(t)}{\lambda(t) + \rho(a) + \epsilon(a)} S_{10}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \lambda(t) + \rho(a) + \epsilon(a) \right] \right\} \right\} q_j + \\
& \frac{c(a)b(1-q(a))}{\rho(a) + \epsilon(a) + n \alpha} I_{10}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \rho(a) + \epsilon(a) + n \alpha \right] \right\} \right\}.
\end{align*}
\]

\[
\begin{align*}
(20) \quad & I_{1011+j}(t+1,a+1) = I_{1011+j}(t,a) \exp \left\{ -\left[ \rho(a) + \gamma(t) + \mu(a) + \beta(a) + n \alpha \right] \right\} + \\
& \frac{n \alpha}{\rho(a) + \gamma(t) + n \alpha} I_{1011+j-1}(t,a) \exp(-\mu(a) - \beta(a)) \left\{ 1 - \exp \left\{ -\left[ \rho(a) + \gamma(t) + n \alpha \right] \right\} \right\} + \\
& \frac{bfp(t)c(a)q(a)}{\lambda(t) + \rho(a) + \epsilon(a)} S_{00}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \lambda(t) + \rho(a) + \epsilon(a) \right] \right\} \right\} q_j + \\
& \frac{fp(t)c(a) + \lambda(t)}{\lambda(t) + \rho(a) + \epsilon(a)} S_{10}(t,a) \exp(-\mu(a) - \beta(a)) \left\{ 1 - \exp \left\{ -\left[ \lambda(t) + \rho(a) + \epsilon(a) \right] \right\} \right\} q_j + \\
& \frac{c(a)bq(a)}{\rho(a) + \epsilon(a) + n \alpha} I_{10}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \rho(a) + \epsilon(a) + n \alpha \right] \right\} \right\}.
\end{align*}
\]

\[
\begin{align*}
(21) \quad & I_{1100+j}(t+1,a+1) = I_{1100+j}(t,a) \exp \left\{ -\left[ \sigma(a) + \mu(a) + n \alpha \right] \right\} + \\
& \frac{n \alpha}{\sigma(a) + n \alpha} I_{1100+j-1}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \sigma(a) + n \alpha \right] \right\} \right\} + \\
& \frac{\rho(a)}{\rho(a) + \sigma(a) + n \alpha} I_{1100+j}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \rho(a) + n \alpha \right] \right\} \right\} + \\
& \frac{(1-b)fp(t)c(a)(1-q(a))}{\lambda(t) + \sigma(a) + \epsilon(a)} S_{01}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \lambda(t) + \sigma(a) + \epsilon(a) \right] \right\} \right\} q_j + \\
& \frac{fp(t)c(a) + \lambda(t)}{\lambda(t) + \sigma(a) + \epsilon(a)} S_{11}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \lambda(t) + \sigma(a) + \epsilon(a) \right] \right\} \right\} q_j + \\
& \frac{c(a)(1-b)(1-q(a))}{\lambda(t) + \sigma(a) + \epsilon(a) + n \alpha} I_{10}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \sigma(a) + \epsilon(a) + n \alpha \right] \right\} \right\}.
\end{align*}
\]

\[
\begin{align*}
(22) \quad & I_{1101+j}(t+1,a+1) = I_{1101+j}(t,a) \exp \left\{ -\left[ \sigma(a) + \mu(a) + \beta(a) + n \alpha \right] \right\} + \\
& \frac{n \alpha}{\sigma(a) + n \alpha} I_{1101+j-1}(t,a) \exp(-\mu(a) - \beta(a)) \left\{ 1 - \exp \left\{ -\left[ \sigma(a) + n \alpha \right] \right\} \right\} + \\
& \frac{\rho(a)}{\rho(a) + n \alpha} I_{1101+j}(t,a) \exp(-\mu(a) - \beta(a)) \left\{ 1 - \exp \left\{ -\left[ \rho(a) + n \alpha \right] \right\} \right\} + \\
& \frac{(1-b)fp(t)c(a)q(a)}{\lambda(t) + \sigma(a) + \epsilon(a)} S_{01}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \lambda(t) + \sigma(a) + \epsilon(a) \right] \right\} \right\} q_j + \\
& \frac{fp(t)c(a) + \lambda(t)}{\lambda(t) + \sigma(a) + \epsilon(a)} S_{11}(t,a) \exp(-\mu(a) - \beta(a)) \left\{ 1 - \exp \left\{ -\left[ \lambda(t) + \sigma(a) + \epsilon(a) \right] \right\} \right\} q_j + \\
& \frac{c(a)(1-b)q(a)}{\sigma(a) + \epsilon(a) + n \alpha} I_{10}(t,a) \exp(-\mu(a)) \left\{ 1 - \exp \left\{ -\left[ \sigma(a) + \epsilon(a) + n \alpha \right] \right\} \right\}.
\end{align*}
\]
This material, provided by the authors as a supplement to Blood Transfusion and Spread of Variant Creutzfeldt-Jakob Disease, is not part of Emerging Infectious Diseases contents.

\[ (23) \quad I_{1101}(t + 1, a + 1) = I_{1110}(t, a)\exp \left\{ - \left[ \sigma(a) + \mu(a) + \gamma(t) + n\alpha \right] \right\} + \frac{n\alpha}{\sigma(a) + \gamma(t) + n\alpha} I_{1100}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \sigma(a) + \gamma(t) + n\alpha \right] \right\} \} + \frac{\rho(a)}{\rho(a) + \gamma(a) + n\alpha} I_{1010}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \rho(a) + \gamma(t) + n\alpha \right] \right\} \} + \frac{bfp(t)\epsilon(a)(1 - q(a))}{\lambda(t) + \sigma(a) + \epsilon(a)} S_{10}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \lambda(t) + \sigma(a) + \epsilon(a) \right] \right\} \} q_j + \frac{\epsilon(a)b(1 - q(a))}{[\sigma(a) + \epsilon(a) + n\alpha]} I_{01}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \sigma(a) + \epsilon(a) + n\alpha \right] \right\} \} \]

\[ (24) \quad I_{1111}(t + 1, a + 1) = I_{1111}(t, a)\exp \left\{ - \left[ \sigma(a) + \mu(a) + \beta(a) + \gamma(t) + n\alpha \right] \right\} + \frac{n\alpha}{\sigma(a) + \gamma(t) + n\alpha} I_{1110}(t, a)\exp(-\mu(a) - \beta(a)) \{ 1 - \exp \left\{ - \left[ \sigma(a) + \gamma(t) + n\alpha \right] \right\} \} + \frac{\rho(a)}{\rho(a) + \gamma(a) + n\alpha} I_{1101}(t, a)\exp(-\mu(a) - \beta(a)) \{ 1 - \exp \left\{ - \left[ \rho(a) + \gamma(t) + n\alpha \right] \right\} \} + \frac{bfp(t)\epsilon(a)(1 - q(a))}{\lambda(t) + \sigma(a) + \epsilon(a)} S_{11}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \lambda(t) + \sigma(a) + \epsilon(a) \right] \right\} \} q_j + \frac{\epsilon(a)b(1 - q(a))}{[\sigma(a) + \epsilon(a) + n\alpha]} I_{10}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \sigma(a) + \epsilon(a) + n\alpha \right] \right\} \} \]

\[ (25) \quad I_{1201}(t + 1, a + 1) = I_{1201}(t, a)\exp \left\{ - \left[ \sigma(a) + n\alpha \right] \right\} + I_{1200}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \gamma(t) + n\alpha \right] \right\} \} + \frac{fp(t)\epsilon(a)(1 - q(a))}{\lambda(t) + \epsilon(a)} S_{20}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \lambda(t) + \epsilon(a) \right] \right\} \} q_j + \frac{fp(t)\epsilon(a)}{\lambda(t) + \epsilon(a)} I_{20}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \lambda(t) + \epsilon(a) \right] \right\} \} q_j + \frac{\gamma(t) + \rho(a) + n\alpha}{\gamma(t) + \sigma(a) + n\alpha} I_{1100}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \gamma(t) + \rho(a) + n\alpha \right] \right\} \} + \frac{\gamma(t) + \sigma(a)}{\gamma(t) + \sigma(a) + n\alpha} I_{1110}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \gamma(t) + \sigma(a) + n\alpha \right] \right\} \} + \frac{\sigma(a)}{[\sigma(a) + n\alpha]} I_{1101}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \sigma(a) + n\alpha \right] \right\} \} + \frac{\epsilon(a)(1 - q(a))}{[\epsilon(a) + n\alpha]} I_{02}(t, a)\exp(-\mu(a)) \{ 1 - \exp \left\{ - \left[ \epsilon(a) + n\alpha \right] \right\} \} \]
In addition to the 26 variables we also calculate the cumulative number of individuals who have died due to the infection. These numbers we do not calculate for the individual ages separately, i.e. we sum all age groups. Since we have 13 types of infected individuals we define those who died in the corresponding 13 categories:
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References (Appendix)