Venezuelan equine encephalitis virus (VEEV) is an emerging pathogen of equids and humans, but infection of its rodent reservoir hosts has received little study. To determine whether responses to infection vary among geographic populations, we inoculated 3 populations of cotton rats with 2 enzootic VEEV strains (Co97-0054 [enzootic ID subtype] and 68U201 [enzootic IE subtype]). The 3 populations were offspring from wild-caught cotton rats collected in a VEE-enzootic area of south Florida, USA; wild-caught cotton rats from a non–VEE-enzootic area of Texas, USA; and commercially available (Harlan) colony-reared cotton rats from a non–VEE-enzootic region. Although each population had similar early viremia titers, no detectable disease developed in the VEE-sympatric Florida animals, but severe disease and death affected the Texas and Harlan animals. Our findings suggest that the geographic origins of cotton rats are important determinants of the outcome of VEE infection and reservoir potential of these rodents.

Vertebrate reservoir hosts play an important role in maintenance and dissemination of zoonotic pathogens. For arthropodborne viruses (arboviruses), infected hosts generally show little or no disease, which presumably reflects long-term selection for host resistance and possibly virus attenuation (1,2). Understanding how pathogens affect their reservoir hosts as well as how the reservoir affects fitness and replication of the pathogen could enable better prediction of emergence, reemergence, or extinction of zoonotic pathogens in response to environmental changes. For example, changes in a reservoir host’s habitats and ecology, due to anthropogenic or natural causes, may affect pathogen transmission to humans and domestic animals. A better understanding of reservoir biology and of pathogen-reservoir interactions, particularly their mechanisms of disease resistance, could also facilitate the development of treatment and control strategies for humans and domestic animals (3,4).

Venezuelan equine encephalitis virus (VEEV), a member of the family Togaviridae, genus Alphavirus (5), was first isolated and characterized in 1938 (6,7) and affects humans and equids in the Americas (8,9). VEEV strains are classified into 2 epidemiologic groups: enzootic and epizootic. Enzootic strains (subtype I, varieties D and E, as well as related species in the VEE complex comprising subtypes II–VI) have been regularly isolated in lowland tropical forests and swamps in Florida, Mexico, and Central and South America. These enzootic viruses generally circulate between Culex (Melanoconion) spp. mosquito vectors and rodent reservoirs and are usually avirulent for and incapable of amplification in equids (8). In contrast, epizootic (and epidemic) VEEV strains (subtype I, variety AB and variety C), have been responsible for all major equine outbreaks that have involved other mosquito vectors, primarily Aedes and Psorophora spp. Epizootics and epidemics have occurred from southern North America to northern South America, and the VEEV strains involved have caused debilitating neurologic disease with high fatality rates in equids (9–11). In humans, who are tangential hosts during endemic and epizootic cycles, severe febrile illness can become life threatening. Although <1% of infected humans die, permanent neurologic sequelae can occur in survivors, particularly young children (8,12,13).

Serosurveys have found VEEV antibodies in many species of small mammals (14–18). However, spiny rats (Proechimys spp., family Echimyidae) and cotton rats (Sigmodon spp., family Cricetidae) have been most often implicated as principal reservoir hosts for enzootic strains, based on seroprevalence and experimental infections demonstrating viremia adequate in titer and duration to infect enzootic
mosquito vectors (19–22). Their geographic distributions are different, but overlapping: Proechimys spp. are found mainly in Panama, northern Peru, Bolivia, Paraguay, and Brazil; Sigmodon spp. are found mainly from southern North America to northern parts of Venezuela, Colombia, and Peru (23).

Studies supporting the role of cotton rats as reservoir hosts for enzootic VEEV have investigated viremia and antibody responses as well as horizontal transmission in laboratory settings (19–21, 24, 25). Howard et al. reported illness and death in cotton rats after infection with a Texas epizootic subtype IB strain (21). The cause of death was linked more to experimental manipulation than to virus infection. Another study that examined clinical and histopathologic manifestations after infection of rats with Everglades virus (EVEV; subtype II in the VEE complex) reported that although viremia developed and the virus replicated in a wide variety of organs, only 2% died (25).

The southern United States has 12 native subspecies of cotton rats (26), which differ by as much as 5% in their cytochrome b DNA sequence (27). To determine whether responses to infection vary among these geographic populations, we studied infection with 2 different subtypes of enzootic VEEV in 3 populations of cotton rats.

Materials and Methods

Animals

Three subspecies of Sigmodon hispidus (cotton rat) were used in this study: the Harlan colony, the Texas population, and the Florida population. The Harlan colony consisted of 6- to 8-week-old female rats purchased from Harlan (Indianapolis, IN, USA) from an established colony. Because the exact geographic origin was unknown, cytochrome B mitochondrial gene sequences were amplified by PCR and compared phylogenetically with those from cotton rats from various locations in North America (27). The sequences from the Harlan colony grouped with those of animals collected in east Texas, Louisiana, and Tennessee but were outside of the clade from southern Florida (data not shown), which indicated that these rats originated from a nonenzootic region. (Florida is the only VEE complex alphavirus–enzootic region in the United States, aside from the Rocky Mountains, which are outside of Sigmodon distribution [8,23]). The Texas population consisted of 4- to 12-month-old wild-caught male and female S. hispidus berlandieri trapped in Galveston Island State Park, Texas (29.27°N, 94.83°W) (25). The Florida population consisted of 3- to 21-week-old male and female cotton rats (S. hispidus spadicipygus) trapped in southern Florida (25) and used to rear F1 rats for experimental studies. Before inoculation, all rats were tested for antibodies against VEEV, EVEV, and Eastern equine encephalitis virus (EEEV) and acclimated for 3 days in a Biosafety Level 3 animal facility. All experiments included ≥2 animals as negative controls. All studies were approved by the University of Texas Medical Branch Animal Care and Use Committee.

Viruses and Infections

VEEV strain Co97-0054 (enzootic ID subtype), isolated in Colombia in 1997 from a sentinel hamster, and strain 68U201 (enzootic IE), isolated in Guatemala from a sentinel hamster and derived from a cDNA clone (28), were used for experimental infections. These strains were selected because they had low passage histories and represent the 2 major enzootic VEEV subtypes; strain Co97-0054 has also been used for experimental infection of spiny rats (22). Cotton rats were inoculated subcutaneously in the left footpad or left thigh with 3–4 log10 PFU of virus, a dose consistent with the maximum amount of VEEV in mosquito saliva (29). To determine whether an increase in virus inoculum could change the outcome of the disease, 4 Florida rats were also inoculated with 5–6 log10 PFU of VEEV strain 68U201. All animals were observed for signs of illness once a day for 15 days.

To determine the neurovirulence of VEEV in the Florida population, 3 rats (2 months of age) were inoculated intracranially with 3 log10 PFU/mL of strain 68U201. Blood samples were collected 24 h after inoculation, and the rats were observed for signs of disease.

Viremia Assays

Blood samples were collected from the retroorbital sinus for ≤10 days after inoculation. Serum samples were diluted 1:10 in Eagle minimal essential medium supplemented with 20% fetal bovine serum, gentamicin, and L-glutamine and stored at −80°C. Viremia titers were determined by plaque assay with Vero cells (30).

Antibody Assays

Plaque reduction (80%) neutralization (PRNT) and hemagglutination inhibition (HI) tests were performed (30). To detect specific VEEV immunoglobulin M (IgM) antibodies, an IgM-capture ELISA was performed (31). Briefly, microplates were coated with anti-rat IgM, diluted 1:500 in carbonate-bicarbonate buffer pH 9.6 (Kirkegaard and Perry Laboratories, Gaithersburg, MD, USA), and incubated at 4°C for at least 16 h. Subsequently, the following were sequentially added: test serum, mouse immune ascitic fluid prepared against VEEV antigens, anti-mouse conjugate (Kirkegaard and Perry), and ABTS (2,2′-azino-bis[3-ethylbenzthiazoline-6-sulphonic acid])–peroxidase substrate (Kirkegaard and Perry). Test serum samples were diluted at 1:40 in 0.5% bovine serum albumin in phosphate-buffered saline at pH 7.4, and the plates were read by using a spectrophotometer with a 405-nm wavelength filter.
cut-off value was calculated as the mean optical density (OD) of negative control samples plus 3 standard deviations, or 0.200. Linear regression, the Student t test, and analysis of variance were used to analyze data.

Results

Clinical Responses and Survival

Inoculation of the Florida rats with 3 log_{10} PFU of VEEV strain 68U201 (IE) or Co97-0054 (ID) resulted in no detectable signs of illness and survival rates of 100% and 87.5%, respectively (Figure 1). One Florida rat inoculated with strain Co97-0054 died on day 10 postinoculation without exhibiting any signs of illness. These findings contrasted with the results of VEEV infections of the Texas and Harlan populations. Although the Harlan rats were inoculated with only the subtype ID strain, signs of severe illness developed in all of the Harlan and Texas rats beginning on day 5. Signs included ruffled coats, lack of grooming, lethargy, and for many, signs of encephalitis (incoordination and instability when walking and erratic movements of the head and limbs), dehydration (measured by lack of skin turgor), and anorexia. Most animals that died before day 5 postinoculation showed no prior signs of illness. The average survival time for the Texas population was 6.8 days after inoculation with the subtype ID strain and 8.2 days with the IE strain; for the Harlan colony, it was 5 days after inoculation with the subtype ID VEEV. None of the animals that survived past day 15 died. The 2 sham-inoculated and the 2 noninoculated rats survived without signs of disease.

Neurovirulence in Florida Cotton Rats

To determine whether the absence of disease in the Florida population was due to the inability of the virus to penetrate the central nervous system, 3 rats were inoculated intracranially with 3 log_{10} PFU/mL of subtype IE VEEV. Viremia titers at 24 h postinoculation were 6.3, 6.2, and 6.8 log_{10} PFU/mL (mean 6.5). By day 3 postinoculation, all of these rats started showing signs of illness, including ruffling of the fur and lack of movement. By day 9 postinoculation, 1 rat was dead and the other 2 exhibited instability and difficulty in walking, uncoordinated and erratic movements of the head and limbs, dehydration, and anorexia; these animals were euthanized because of the severity of disease. Histopathologic examination of the brains showed clear signs of encephalitis, focal meningoencephalitis (Figure 2, panels A, B) and associated perivascular mononuclear cell infiltration (Figure 2, panel C), and neuropathia, which led to the conclusion that the cause of death was from the viral infection and not from the injection or manipulation of the animals.

Dose Dependence

To determine whether the Florida population’s apparent resistance to VEE after peripheral infection was dose-dependent, 8 additional animals were inoculated with 5 or 6 log_{10} PFU (4 animals per dose). In each population, 3 (75%) of the rats survived infection (data not shown). One rat inoculated with 6 log_{10} PFU of virus succumbed to disease on day 3, whereas another animal inoculated with 5 log_{10} PFU of virus died on day 11, which suggests that the dose could have affected disease progression. None of the other inoculated animals showed any clinical signs of illness.
Viremia Titer

Viremia profiles for the Florida rats were similar after inoculation (3 log_{10} PFU) with either subtype ID or IE VEEV (p>0.05). Peak viremia titers of 6.2 and 5.4 log_{10} PFU/mL, respectively, occurred at 24–48 h postinoculation, then became undetectable by days 4–5 postinoculation (Figure 3). The Texas population inoculated with 3–4 log_{10} PFU of subtype IE virus and 3 log_{10} PFU of subtype ID VEEV had similar viremia profiles, with peak titers of 6.1 and 6.6 log_{10} PFU/mL, respectively, at 24–48 h.

We found in the Florida population a correlation between age and peak viremia titers. Younger animals inoculated with the IE virus had higher peak titers on days 1 and 2 postinoculation than did older animals (Figure 4, Table 1). In addition, we observed a significant difference in mean viremia titers on day 2 between 3- and 8-week-old animals and on day 3 between 5- and 8-week-old animals (p<0.05).

Differences in Virus Titers among Cotton Rat Populations

No differences in viremia profile were observed between the Texas and Harlan rats, for which VEEV infection with subtype ID was generally fatal (p>0.05). Viremia in the Texas and Harlan rats peaked between 24 and 48 h postinoculation, with mean titers of ≈6.7 log_{10} PFU/mL, and was undetectable by day 8 postinoculation. In contrast, significant differences were observed in peak viremia titers between the Florida and Texas rats inoculated with either the ID or IE VEEV subtype (p<0.05). In all cases, Texas rats produced higher titers (24 h postinoculation) and had a longer duration of viremia than the Florida rats, in which no disease was apparent (Figure 3, Tables 2, 3). Similar results occurred when Florida and Harlan rats inoculated with subtype ID were compared.

Antibody Responses

To determine whether the difference in disease outcome in Florida versus Texas rats was due to a difference in antibody responses, serum was tested by PRNT. In the Texas population inoculated with subtype IE VEEV, low titers of neutralizing antibodies (NAb) were produced by days 5–6 postinoculation (mean titer 20), and mean titers never exceeded 40 by day 10 (Figure 5, panel A); however, detectable NAb were not produced in all animals that died. In contrast, some Florida rats inoculated with the same virus strain had detectable NAb titers by day 4, and all had detectable titers by day 6. NAb titers were significantly lower in the Texas rats on days 5, 6, and 7 (p = 0.02, 0.04, 0.04, respectively).

When Texas rats were inoculated with the subtype ID VEEV strain, even lower NAb titers were produced, despite the development of higher viremia titers during the later stages, compared with those inoculated with subtype IE. Peak NAb titers occurred on day 8 postinoculation (mean 25), and 2 of the surviving animals had no detectable NAb titers (<19) late during infection (day 32). In contrast, in all Florida rats, detectable NAb developed by day 4 and peaked on day 6 (mean 57), when they were significantly higher (p = 0.004).

To determine the duration of the antibody response in the Florida population, we measured NAb titers in 2 rats for 7 months postinoculation. Titers peaked at 4 months and then gradually decreased to the detection limit by 7 months, when the experiment was terminated (Figure 5, panel B).

Because NAb were not detected in all infected animals, serum samples were further tested by HI and IgM ELISA. All rats had detectable HI antibodies by days 5–6 postinoculation. Although the titers were relatively low (<200) on
Table 1. Mean viremia titers of Florida cotton rats inoculated subcutaneously with Venezuelan equine encephalitis virus, subtype IE*

<table>
<thead>
<tr>
<th>Day postinfection</th>
<th>Age, wk</th>
<th>Florida, IE</th>
<th>Florida, ID</th>
<th>Texas, IE</th>
<th>Texas, ID</th>
<th>Harlan, ID</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>3 (n = 2)</td>
<td>6.7 ± 0.2</td>
<td>5.9 ± 0.2</td>
<td>6.3 ± 0.2</td>
<td>6.1 ± 0.2</td>
<td>6.8 ± 0.9</td>
</tr>
<tr>
<td>2</td>
<td>5 (n = 2)</td>
<td>5.2 ± 0.2</td>
<td>4.9 ± 0.2</td>
<td>6.4 ± 0.1</td>
<td>6.6 ± 0.2</td>
<td>5.8 ± 0.9</td>
</tr>
<tr>
<td>3</td>
<td></td>
<td>3.7 ± 0.4</td>
<td>4.1 ± 0.2</td>
<td>5.6 ± 0.2</td>
<td>6.3 ± 0.2</td>
<td>6.5 ± 0.5</td>
</tr>
<tr>
<td>4</td>
<td></td>
<td>&lt;1.9</td>
<td>2.3 ± 0.2</td>
<td>3.8 ± 0.3</td>
<td>5.6 ± 0.2</td>
<td>4.7 ± 0.8</td>
</tr>
<tr>
<td>5</td>
<td></td>
<td>&lt;1.9</td>
<td>&lt;1.9</td>
<td>2.6 ± 0.3</td>
<td>5.0 ± 0.5</td>
<td>3.9 ± 0.1</td>
</tr>
<tr>
<td>6</td>
<td></td>
<td>&lt;1.9</td>
<td>&lt;1.9</td>
<td>2.2 ± 0.2</td>
<td>3.6 ± 0.7</td>
<td>3.9 ± 0.2</td>
</tr>
<tr>
<td>7</td>
<td></td>
<td>&lt;1.9</td>
<td>&lt;1.9</td>
<td>&lt;1.9</td>
<td>&lt;1.9</td>
<td>2.0 ± 0.0</td>
</tr>
<tr>
<td>8</td>
<td></td>
<td>&lt;1.9</td>
<td>&lt;1.9</td>
<td>&lt;1.9</td>
<td>&lt;1.9</td>
<td>2.1 ± 0.3</td>
</tr>
</tbody>
</table>

*Titers are expressed as log10 PFU/mL ± standard error; 1.9 log10 PFU/mL is the limit of detection of the plaque assay.

Table 2. Mean viremia titers of Florida, Texas, and Harlan cotton rats inoculated subcutaneously with Venezuelan equine encephalitis virus, subtype ID or IE*

<table>
<thead>
<tr>
<th>Day postinfection</th>
<th>Cotton rat population and virus subtype</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Florida, IE</td>
</tr>
<tr>
<td>1</td>
<td>6.1 ± 0.2</td>
</tr>
<tr>
<td>2</td>
<td>5.2 ± 0.2</td>
</tr>
<tr>
<td>3</td>
<td>3.7 ± 0.4</td>
</tr>
<tr>
<td>4</td>
<td>&lt;1.9</td>
</tr>
<tr>
<td>5</td>
<td>&lt;1.9</td>
</tr>
<tr>
<td>6</td>
<td>&lt;1.9</td>
</tr>
<tr>
<td>7</td>
<td>&lt;1.9</td>
</tr>
<tr>
<td>8</td>
<td>&lt;1.9</td>
</tr>
</tbody>
</table>

*Titers are expressed as log10 PFU/mL ± standard error; 1.9 log10 PFU/mL is the limit of detection of the plaque assay.
other zoonotic viruses, which presumably reflects long-term selection for resistance (1,2). This resistance is little studied and poorly understood, yet it might provide insight into improved treatments for arbovirus infections in humans. Our findings may also have implications for VEEV ecology, especially in the event of virus introduction into a nonenzootic region, as occurred during the 1971 Texas VEEV epizootic (38,39). During such a scenario, virus-induced deaths might deplete cotton rat populations, depending on the VEEV transmission levels and the length of the outbreak.

Viremia and Immunologic Responses

The differences in viremia profiles exhibited by the VEE-sympatric versus VEE-allopatric cotton rat populations could explain the different disease outcomes. Although peaks of viremia titers were similar for both subspecies, durations of viremia differed. The prolonged viremia observed in the Texas and Harlan animals may reflect a poor or delayed innate or adaptive immune response, which led to sustained viral replication and death. This could be an indirect effect of replication in lymphatic tissues leading to immunosuppression. Although peak viremia titers appeared to be age-dependent in the Florida population inoculated with the subtype IE VEEV strain, disease outcomes between age groups, which might reflect maturation of the immune system, did not differ significantly. Antibody titers in both populations of cotton rats after inoculation with either virus strain were relatively low.

Table 3. Statistical comparisons (p values) among viremia titers in Texas and Florida cotton rats inoculated with Venezuelan equine encephalitis virus, subtype IE or ID*

<table>
<thead>
<tr>
<th>Day postinfection</th>
<th>Texas vs. Florida, subtype IE</th>
<th>Texas vs. Florida, subtype ID</th>
<th>Subtype IE vs. ID, Texas</th>
<th>Subtype IE vs. ID, Florida</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>0.123</td>
<td>0.018</td>
<td>0.590</td>
<td>0.078</td>
</tr>
<tr>
<td>2</td>
<td>$5.0 \times 10^{-4}$</td>
<td>$4.69 \times 10^{-5}$</td>
<td>0.506</td>
<td>0.078</td>
</tr>
<tr>
<td>3</td>
<td>$9.75 \times 10^{-7}$</td>
<td>$2.72 \times 10^{-6}$</td>
<td>0.506</td>
<td>0.215</td>
</tr>
<tr>
<td>4</td>
<td>$2.82 \times 10^{-10}$</td>
<td>$3.66 \times 10^{-8}$</td>
<td>0.348</td>
<td>0.751</td>
</tr>
<tr>
<td>5</td>
<td>$1.31 \times 10^{-9}$</td>
<td>$5.71 \times 10^{-5}$</td>
<td>0.058</td>
<td>1</td>
</tr>
<tr>
<td>6</td>
<td>0.051</td>
<td>0.010</td>
<td>0.123</td>
<td>–</td>
</tr>
</tbody>
</table>

*Numbers in **boldface** indicate statistically significant differences (analysis of variance).

Figure 5. Antibody responses in cotton rats from Florida and Texas. A) Neutralizing antibody (NAb) titers in Florida group (n = 3–11) and Texas group (n = 1–17) inoculated with subtypes IE or ID Venezuelan equine encephalitis virus (VEEV). B) Long-term NAb titers in Florida rats infected with subtype IE VEEV (n = 2). C) Hemagglutination inhibition (HI) antibody titers for Florida (n = 2–10) and Texas (n = 1–16) rats inoculated with subtype IE VEEV. D) Immunoglobulin M antibody titers for Florida and Texas rats infected with subtype IE VEEV (n = 2). OD, optical density.
These findings contrast with results of a previously published study of EVEV infection of cotton rats from the same 2 geographic regions (25). In that study, both sub-species of rats survived infection, exhibited similar peak viremia titers, and had high antibody titers within 9 days postinoculation; it was suggested that the innate immune response was involved. EVEV, enzootic in South Florida, is less virulent in laboratory rodents than in most other viruses in the VEE complex, including the subtypes we used (19,40). Presumably due to this lack of virulence, Florida and Texas strains of cotton rats tested produced protective antibodies and survived EVEV infection (25).

Our data from the same 2 rat populations but inoculated with more virulent strains of VEEV present a different picture. Although the innate immune response may be involved as well, antibody detection correlated with the disappearance of viremia. The capability of Florida cotton rats to produce antibodies against VEEV early may allow them to better control replication and survive. Antibodies against VEEV persist for at least 6 months in laboratory-infected cotton rats (25). The average lifespan of cotton rats in nature is estimated to be 6–8 months (41).

To identify protective mechanisms in the Florida population, additional studies focusing more on the innate immune responses of enzootic and nonenzootic cotton rat populations are needed. This could be approached by cross-breeding the Texas and the Florida rats and studying the offspring. Elucidation of protective mechanisms may be useful for developing new strategies for treating human or equine infections.

Acknowledgments

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Dr Carrara is a postdoctoral fellow in the Department of Pathology, University of Texas Medical Branch. Her research focuses on the interactions between reservoir rodent hosts and VEEV.

References


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