
Population-Attributable Risk Estimates for Risk Factors Associated with *Campylobacter* Infection, Australia

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In 2001–2002, a multicenter, prospective case-control study involving 1,714 participants ≥ 5 years of age was conducted in Australia to identify risk factors for *Campylobacter* infection. Adjusted population-attributable risks (PARs) were derived for each independent risk factor contained within the final multivariable logistic regression model. Estimated PARs were combined with adjusted (for the ≥ 5 years of age eligibility criterion) notifiable disease surveillance data to estimate annual Australian *Campylobacter* case numbers attributable to each risk factor. Simulated distributions of “credible values” were then generated to model the uncertainty associated with each case number estimate. Among foodborne risk factors, an estimated 50,500 (95% credible interval 10,000–105,500) cases of *Campylobacter* infection in persons ≥ 5 years of age could be directly attributed each year to consumption of chicken in Australia. Our statistical technique could be applied more widely to other communicable diseases that are subject to routine surveillance.

Foodborne gastroenteritis is a major public health concern in many countries, including Australia. A recent study estimated that 5.4 million cases (95% credible interval [CrI] 4.0–6.9 million), 15,000 hospitalizations (95% CrI 11,000–18,000), and 80 deaths (95% CrI 40–120) annually are caused by foodborne gastroenteritis in Australia (1). Norovirus, enteropathogenic *Escherichia coli*, *Salmonella* spp., and *Campylobacter* spp. accounted for 88% of the estimated 1.5 million (95% CrI 1.0–1.9 million) cases of foodborne disease caused by known pathogens.

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Among known foodborne pathogens, *Campylobacter* spp. are the most frequently reported enteric pathogens in Australia (2). The incidence of *Campylobacter* infection steadily increased from 1991 through 2001 but has been relatively stable since. In 2005, >15,000 cases were reported in Australia, a crude rate of 113.0/100,000 population. However, because of underreporting, $\approx 223,000$ *Campylobacter* infections are estimated to occur annually; $\approx 75\%$ of these are foodborne (3). Most of these infections are sporadic.

Case-control studies have identified a range of different risk factors for infection; consumption of chicken is the most frequently reported (4–9). Some of these studies report population-attributable fractions associated with independent risk factors, but no estimates of the total magnitude of infection caused by chicken or other risk factors have yet been reported. Using a multicentered, prospective case-control study, we aimed to develop a multivariable logistic regression model that identified independent foodborne and nonfoodborne risk factors for *Campylobacter* infection for this sample (7) and calculate population-attributable risk (PAR) proportions. These PARs were then combined with annual *Campylobacter* infection surveillance data to estimate the total number of infections (with associated CrIs) among persons ≥ 5 years of age attributable to specific risk factors that occur in the community each year in Australia.

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Methods

Study Design and Population

From September 2001 through August 2002, a multicenter, prospective case-control study was conducted across 5 of the 8 states and territories in Australia to identify risk factors for *Campylobacter* infection in persons ≥ 5 years of age. These jurisdictions were those with legislation that required physicians and laboratories to notify health departments about patients infected with *Campylobacter*. At the time of this study, the population of the 5 states combined was ≈ 12 million, and the total population of Australia was ≈ 19 million.

Case-Patients and Controls

A case-patient was defined as a person ≥ 5 years of age reported with a culture-positive stool result for *Campylobacter* infection and a recent history of acute diarrhea, who was not part of an outbreak investigation unless identified as the index patient. Controls were sourced from a national control bank and frequency matched to case-patients by age groups in each state. The age groups were selected on the basis of potential variation in risk factors due to different behavior at different ages. Age groups were children (5–9 years), adolescents (10–19 years), young adults (20–29 years), middle-aged adults (30–59 years), and elderly (≥ 60 years).

A total of 881 case-patients and 883 controls were recruited for this study. A telephone-administered questionnaire was used to collect detailed information on exposures in the 7 days before onset of illness for case-patients and in the 7 days before interview for controls. The questionnaire comprised several sections, each representing a separate exposure group that listed questions pertaining to potential risk factors related to that group. The following sections were included: meat, poultry and seafood consumption; egg and dairy product consumption; produce consumption; water consumption; food-handling practices; animal and pet exposures; host factors; dining locations outside the home; overseas travel; and demographic information. To measure the effects between illness and consumption of cooked meat products or undercooked meat products, additional information was sought on whether the meat appeared undercooked (pink on the inside) when eaten. A detailed description of the study design, sample, and exposure measurements has been published elsewhere (7).

Data Analysis

A 2-stage model-building strategy was undertaken, first by determining a parsimonious multivariable model for each exposure group, and second by deriving an omnibus parsimonious model that combines significant exposure variables from all the exposure group multivariable

models. A more comprehensive description of the analytical model has been published elsewhere (7) and is included in the online Technical Appendix (available from www.cdc.gov/EID/content/14/6/895-Techapp.pdf).

We calculated PARs by using adjusted odds ratios (aORs) from the final multivariable logistic regression model for each variable that was significantly associated with an increased risk for infection, apart from host factors (10). Stata statistical software, release 7 (Stata Corp, College Station, TX, USA), was used for calculating 95% confidence intervals (CIs) around the PAR estimates. Using community incidence data derived from adjusted national surveillance data (3) coupled with PAR data from our case-control study, we used simulation techniques to estimate the total number of *Campylobacter* infections attributable to specific risk factors that occur in the community each year in Australia and to derive credible regions for these estimates by modeling the uncertainty in each variable component.

Simulation Methods

We assumed that 223,000 (95% CrI 94,000–363,000) cases of campylobacteriosis occur in Australia in a typical year (3). We then adjusted this figure by reviewing Australian notification data for the years 2001 through 2003 (11) and applying simulation techniques to estimate the proportion of cases that occur among persons ≥ 5 years of age. Similarly, we randomly generated simulated PAR values for each risk factor using aORs from the final model. The simulated campylobacteriosis case numbers and PAR-simulated values were multiplied together to produce distributions of the total number of *Campylobacter* infections attributable to each specific risk factor. Because some distributions are skewed, we present medians and 95% CrIs (defined to be the 2.5 and 97.5 percentiles) for the simulation results. Simulations were undertaken in SAS System for Windows, version 9.1 (SAS Institute Inc., Cary, NC, USA). A detailed description of the simulation technique used to derive these estimates is provided in the online Technical Appendix. The full description of the sample and the development of the final multivariable logistic regression model have been published elsewhere (7).

Results

Multivariable Analysis of Risk Factors

Table 1 reports results of univariable (crude) and multivariable logistic regression analyses for variables within each exposure group (adjusted for state, sex, and education), and the final multivariable model showing frequency and sample size, percentages, and crude odds ratios (ORs) and aORs, together with 95% CIs. The independent risk factors that were identified in the final model explained

only a limited proportion of illness (Nagelkerke $R^2 = 0.16$). Consumption of undercooked chicken (aOR 4.7, 95% CI 2.6–8.4), consumption of offal (aOR 2.0, 95% CI 1.0–4.0), ownership of domestic dogs <6 months of age (aOR 2.1, 95% CI 1.1–4.2), and ownership of domestic chickens <6 months of age (aOR 12.4, 95% CI 2.6–59.3) were the only independent risk factors for infection after adjusting for all other variables in the model. Consumption of cooked chicken was positively but not statistically associated with illness and warranted further consideration (aOR 1.4, 95%

CI 1.0–1.9, $p = 0.06$). Eating fresh fish, eating homemade foods containing raw eggs, eating organically grown fruit and/or vegetables, and eating homegrown fruit were independent factors associated with a statistically significant reduced risk for infection. Eating raw salads or vegetables, as measured by the vegetable index variable, was also associated with a reduced risk for infection. Drinking commercial bottled water, placing barbecued cooked meat back on the same plate used for raw meat, having liver disease, and having any immunosuppressive therapy in the 4-week

Table 1. Results of univariable (crude) and multivariable logistic regression analysis for variables within each exposure group and the final multivariable model, *Campylobacter* infection, Australia, 2001–2002*

| Exposure group/variables† | Case-patients, n/N (%) | Controls, n/N (%) | Univariable analysis | | Multivariable logistic regression analysis (exposure groups) | | Final multivariable model‡ | |
|--|---------------------------|----------------------|-------------------------|----------|--|----------|-------------------------------|----------|
| | | | OR | 95% CI | OR | 95% CI | aOR | 95% CI |
| Meat, poultry and seafood | | | | | | | | |
| Model 1 | | | | | | | | |
| No chicken | 110/711 (15.5) | 162/808 (20.0) | 1.0 | | 1.0 | | 1.0 | |
| Chicken, cooked | 528/711 (74.3) | 618/808 (76.5) | 1.3 | 1.0–1.7 | 1.3 | 1.0–1.8 | 1.4 | 1.0–1.9 |
| Chicken, undercooked | 73/711 (10.3) | 28/808 (3.5) | 3.8 | 2.3–6.3 | 4.4 | 2.6–7.5 | 4.7 | 2.6–8.4 |
| Offal | 36/852 (4.2) | 16/830 (1.9) | 2.2 | 1.2–4.4 | 2.1 | 1.1–3.9 | 2.0 | 1.0–4.0 |
| Fresh fish | 256/833 (30.7) | 332/827 (40.1) | 0.7 | 0.5–0.8 | 0.6 | 0.5–0.8 | 0.7 | 0.5–0.9 |
| Eggs and dairy products | | | | | | | | |
| Model 2 | | | | | | | | |
| Homemade foods containing raw eggs | 40/837 (4.8) | 70/822 (8.5) | 0.5 | 0.4–0.8 | 0.5 | 0.3–0.7 | 0.5 | 0.3–0.8 |
| Produce | | | | | | | | |
| Model 3 | | | | | | | | |
| Organic fruit and vegetables | 50/805 (6.2) | 100/804 (12.4) | 0.5 | 0.3–0.7 | 0.6 | 0.4–0.8 | 0.6 | 0.4–1.0 |
| Homegrown fruit | 84/845 (9.9) | 169/828 (20.4) | 0.4 | 0.3–0.6 | 0.5 | 0.4–0.7 | 0.4 | 0.3–0.6 |
| Vegetable index§ | | | | | | | | |
| 0 (no vegetables) | 141/853 (16.5) | 87/830 (10.5) | 1.0 | | 1.0 | | 1.0 | |
| 1 (1–2) | 339/853 (39.7) | 305/830 (36.7) | 0.7 | 0.5–0.9 | 0.7 | 0.5–1.0 | 0.7 | 0.5–1.0 |
| 2 (3–4) | 352/853 (41.3) | 382/830 (46.0) | 0.6 | 0.4–0.8 | 0.6 | 0.4–0.9 | 0.6 | 0.4–0.9 |
| 3 (5–6) | 21/853 (2.5) | 56/830 (6.7) | 0.2 | 90.1–0.4 | 0.3 | 0.1–0.5 | 0.2 | 0.1–0.5 |
| Water consumption | | | | | | | | |
| Model 4 | | | | | | | | |
| Commercial bottled water | 72/846 (8.5) | 47/820 (5.7) | 1.5 | 1.0–2.3 | 1.6 | 1.1–2.3 | NS | |
| Food-handling practices | | | | | | | | |
| Model 5 | | | | | | | | |
| Barbecued cooked meat placed back on plate used for raw meat | 21/511 (4.1) | 9/471 (1.9) | 2.2 | 1.0–5.5 | 2.3 | 1.0–5.4 | NS | |
| Animal and pet exposure | | | | | | | | |
| Model 6 | | | | | | | | |
| Domestic chickens | | | | | | | | |
| No domestic chicken | 783/846 (92.6) | 777/821 (94.6) | 1.0 | | 1.0 | | 1.0 | |
| Chicken <6 mo of age | 18/846 (2.1) | 5/821 (0.6) | 3.6 | 1.3–9.7 | 5.2 | 1.5–17.8 | 12.4 | 2.6–59.3 |
| Chicken ≥6 mo of age | 45/846 (5.3) | 39/821 (4.8) | 1.1 | 0.7–1.8 | 1.3 | 0.8–2.2 | 1.7 | 0.9–3.0 |
| Domestic dogs | | | | | | | | |
| No dog | 397/839 (47.3) | 452/819 (55.2) | 1.0 | | 1.0 | | 1.0 | |
| Dog <6 mo of age | 48/839 (5.7) | 17/819 (2.1) | 3.2 | 1.8–5.7 | 2.9 | 1.6–5.3 | 2.1 | 1.1–4.2 |
| Dog ≥6 mo of age | 394/839 (47.0) | 350/819 (42.7) | 1.3 | 1.1–1.6 | 1.2 | 1.0–1.5 | 1.2 | 0.9–1.5 |
| Host factors | | | | | | | | |
| Model 7 | | | | | | | | |
| Chronic gastrointestinal condition | 101/873 (11.6) | 50/831 (6.0) | 2.0 | 1.4–3.0 | 2.0 | 1.4–2.9 | 2.3 | 1.5–3.4 |
| Liver disease | 14/875 (1.6) | 2/830 (0.2) | 6.7 | 1.5–61.2 | 5.1 | 1.1–23.0 | NS | |
| Any immunosuppressive agent/therapy | 35/881 (4.0) | 12/833 (1.4) | 2.8 | 1.4–6.0 | 2.8 | 1.4–5.5 | NS | |

*Each model adjusted for state, sex, and education. aOR, adjusted odds ratio; CI, confidence interval; NS, not significant.

†The exposure period for foods is 7 d before onset of illness for case-patients and 7 days before interview for controls.

‡After removal of nonsignificant interaction terms.

§The vegetable index was created to indirectly measure the range of raw produce consumed in the 7-day exposure period for patients and controls. The values of this index variable represented a count of the number of different types of salad/vegetable foods eaten during the exposure period.

exposure period were all removed from the final model during the sequential backward elimination procedure. None of the investigated 2-factor interactions was statistically significant. There was no reason to suspect the adequacy of the final multivariable model (Hosmer-Lemeshow goodness-of-fit test, $p = 0.98$). Additional statistical information, including β -coefficients, standard errors, statistical significance tests, and goodness-of-fit statistics for all multivariable models, is provided in Table 1A in the online Technical Appendix.

PAR Proportions

Among the food exposures, the proportion of study patients who reported eating undercooked chicken was 10.3%. The proportion of *Campylobacter* illness in the study population that could be attributed to the consumption of undercooked chicken was estimated to be 8.1% (95% CI 5.2%–11.1%) (Table 2). A further 21.2% (95% CI 0.0%–36.9%) of *Campylobacter* infections in the population could be attributed to cooked chicken. The overall PAR associated with consumption of chicken was 29.3%.

The proportion of campylobacteriosis patients ≥ 5 years of age that typically occurs each year in Australia was estimated from the simulations to be 191,000 (95% CrI 79,000–310,000). Applying the simulated PAR estimates to the number of cases of campylobacteriosis in Australia among persons ≥ 5 years of age, we estimated 15,000 (95% CrI 6,000–26,500) cases of *Campylobacter* infection could be attributed to eating undercooked chicken in a typical year. Similarly, an additional 35,500 (95% CrI 0–83,500) cases of infection could be attributed to apparently well-cooked chicken. Overall, an estimated 50,500 (95% CrI 10,000–105,500) cases of campylobacteriosis could be attributed to consumption of chicken each year in Australia.

The proportion of case-patients who reported eating offal was 4.2%. The proportion of illness in the study population that could be attributed to the consumption of offal was estimated to be 2.1% (95% CI 0.0%–4.9%). This equates to $\approx 3,500$ (95% CrI 50–8,500) cases of campylobacteriosis each year in Australia.

Among the nonfood exposures, $\approx 5,000$ (95% CrI 500–11,500) cases of campylobacteriosis could be attributed to contact with dogs < 6 months old each year in Australia. Similarly, an estimated 3,500 (95% CrI 1,000–7,000) cases of campylobacteriosis could be attributed to contact with domestic chickens < 6 months old.

Discussion

The PAR proportions from this study indicate that chicken meat may be associated with $> 50,000$ cases of *Campylobacter* infection each year in Australia. These figures provide a strong argument for government and industry to focus efforts on reducing contamination of chicken carcasses with *Campylobacter* through either improved on-farm control or interventions during processing. In addition, the figures justify the continued need for government to continue educating consumers and foodhandlers about the risks associated with the handling of raw chicken and the potential for cross-contamination in the kitchen.

Several case-control studies of sporadic *Campylobacter* infection have calculated PARs of independent foodborne risk factors (4,5,9,12,13). In these studies, the PAR percentage associated with chicken meat was 4.9%–31%, compared with 29.3% in our study. However, none of these studies extrapolated their PAR proportions to provide estimates of the total magnitude of infection in their study populations. The use of surveillance data coupled with an understanding of underreporting of illness from the com-

Table 2. PAR proportions with 95% CIs and community estimates with 95% CrIs for exposures associated with an increased risk for *Campylobacter* infection in persons ≥ 5 y of age, Australia, 2001–2002*†

| Risk factor | No. case-patients | Proportion of case-patients (p_i) | aOR | PAR, % | 95% CI | Estimated no. community case-patients | 95% CrI |
|-----------------------------|-------------------|---------------------------------------|-----------|--------|----------|---------------------------------------|--------------|
| Food exposures | | | | | | | |
| Chicken consumption | | | | | | | |
| No chicken | 110 | 0.155 | Reference | | | | |
| Chicken, cooked | 528 | 0.743 | 1.4 | 21.2 | 0.0–36.9 | 35,500 | 0–83,500 |
| Chicken, undercooked | 73 | 0.103 | 4.7 | 8.1 | 5.2–11.1 | 15,000 | 6,000–26,500 |
| Offal consumption | | | | | | | |
| No | 816 | 0.958 | Reference | | | | |
| Yes | 36 | 0.042 | 2.0 | 2.1 | 0.0–4.9 | 3,500 | 50–8,500 |
| Nonfood exposures | | | | | | | |
| Dogs (domestic) | | | | | | | |
| No dog | 397 | 0.473 | Reference | | | | |
| Dog < 6 mo of age | 48 | 0.057 | 2.1 | 2.9 | 0.3–4.8 | 5,000 | 500–11,500 |
| Dog ≥ 6 mo of age | 394 | 0.47 | 1.2 | – | | | |
| Chickens (domestic) | | | | | | | |
| No domestic chickens | 783 | 0.926 | Reference | | | | |
| Chickens < 6 mo of age | 18 | 0.021 | 12.4 | 1.9 | 0.9–2.9 | 3,500 | 1,000–7,000 |
| Chickens ≥ 6 mo of age | 45 | 0.053 | 1.7 | – | | | |

*PAR, population-attributable risk; CI, confidence interval; CrI, credible interval.

†Calculated from adjusted odds ratios (aOR) derived from the final multivariable logistic regression model.

munity to surveillance systems allows for this extra important step to quantify the extent of illness caused by specific risk factors (3,14,15).

A recent Australian study indicates that 75% (95% CrI 67%–83%) of cases of *Campylobacter* infection may be due to foodborne transmission (1). In our study, the foodborne risk factor with the highest attributable risk was cooked chicken, with an estimated median of 21.2% (95% CrI 0.0%–36.9%); followed by undercooked chicken, with an estimated median of 8.1% (95% CrI 5.2%–11.1%); and offal, with an estimated median of 2.1% (95% CrI 0.0%–4.9%). Although the aOR for cooked chicken is considerably lower than that for undercooked chicken, the high proportion of exposed case-patients (74.3% reported eating cooked chicken) explains the higher PAR. The combined significant foodborne attributable risk estimate found in the study, 31.4% (95% CrI 10.4%–46.8%), is <75%, which suggests that transmission of infection from foodborne vehicles other than chicken is likely to occur.

We interpret the risk associated with cooked chicken as most likely due to the consumption of undercooked chicken that was reported by patients as apparently well cooked or from poor handling during the preparation and cooking of raw chicken. Cross-contamination of cooked or ready-to-eat foods from handling raw chicken and poor food hygiene are considered to be alternative routes of transmission of *Campylobacter* infection (12,16–21). Although no other foods were significantly associated with illness in our study, food-based risk factors implicated from case-control studies conducted outside Australia include eating barbecued red meat or sausages, raw seafood, nonpoultry meat prepared at a restaurant, or pork and drinking unpasteurized milk (4,6,8,9,22). The Nagelkerke R^2 value of 16% for the final most parsimonious multivariable model also suggests that a considerable proportion of our case-patients had unexplained risk factors. The difficulty associated with recalling exposures is a major limitation of case-control studies designed to identify multiple potential risk factors. Bias caused by misclassification of reported exposures invariably dampens estimated effect sizes and may partly explain the failure to identify significant associations between some potential risk factors and illness. It is also likely that a proportion of unexplained cases were in persons infected by a variety of foods that had been subject to cross-contamination from raw chicken in the kitchen during preparation (23). Because eating chicken meat is a relatively common exposure among both patients and controls, our estimates of effect for cooked and undercooked chicken meat may be underestimates, as will be the derived PAR for chicken meat. However, it is reasonable to assume that at least some of the infections that occur in persons in Australia may be acquired from foods other than chicken or offal.

Two Australian case-control studies, including a study of risk factors among young children, have now identified household puppies and domestic chickens as risk factors for *Campylobacter* infection (7,24). Among persons ≥ 5 years of age, an estimated 8,500 cases of infection could be attributed to these 2 exposures in a typical year; the numbers could be expected to be considerably higher if sporadic infections among children <5 years of age were taken into account. These estimates indicate that a substantial portion of disease is caused by transmission of infection through these routes and provide a timely reminder that public health interventions to reduce this infection in the community should not be directed only at foodborne sources. Although variables associated with a reduced risk for infection did not contribute information to this article, several foods were independently associated with a reduced risk for infection, in particular raw fruit and vegetables. A more detailed discussion on factors associated with a reduced risk for infection in our study is published elsewhere (7).

The method used in this study provides an innovative approach to calculate estimates of the total magnitude of infection associated with a specific risk factor in a population, including an estimate of uncertainty. The required components for these calculations include 1) the PAR obtained from a case-control study in which estimates of effect can be generalized to the population under study and 2) an estimate of total community incidence. The method used to derive the incidence used in this study was from reportable disease data from an existing surveillance system and an estimate of underreporting to the surveillance system. Underreporting factors were derived from data on the proportion of case-patients in the community who visit a doctor (P_D), the proportion of case-patients seen by a doctor who have a stool sample taken (P_S), the proportion of correctly identified pathogens in stool samples submitted to laboratories (P_L), and the proportion of positive results that are reported to the surveillance system (P_R). The product of these proportions ($P_D \times P_S \times P_L \times P_R$) is the reported fraction (3). The extent and nature of underreporting will vary with different surveillance systems and for different pathogens. In the future, as more refined methods for calculating the degree of underreporting are developed, these estimates will become more accurate.

PAR estimates are useful for providing a measure of the proportion of illness that can be attributed to individual or multiple causal factors; however, in case-control studies, errors in the estimates of the proportion of cases exposed to a risk factor and/or errors in the estimate of ORs may lead to biased PAR estimates. For example, 1 requirement for estimating PAR is that the study patients be randomly selected from the population of interest and that exposure information be reported without bias. One could argue that the use of culture-confirmed cases in our study is not repre-

sentative of all *Campylobacter* case-patients in the population because patients with more severe symptoms are more likely to have stools collected and tested (3). Therefore, if the exposure information collected from our study patients was different from all case-patients in the general population, the proportion of case-patients exposed to a particular risk factor may be a biased estimate.

Recall and reporting bias are other concerns with case-control studies that may lead to biased estimates of the OR and subsequently the PAR. This is a particular concern for subjective exposures such as undercooked chicken, which are very difficult to measure accurately within a case-control design, so significant associations need to be interpreted with caution. Similarly, it may be difficult for a study participant who reportedly consumed cooked chicken meat to know if the meat was thoroughly cooked. Whether there are differential information biases between case-patients and controls in the reporting of undercooked chicken meat is not clear. In fact, consumption of undercooked chicken may well be systematically underreported by patients. Given the very high prevalence of chicken consumption in the Australian community (81% during the 7-day period before interview among our study controls), finding consumption of undercooked chicken as a risk factor for infection, despite the low reported frequency of exposure, is not surprising. Our PAR estimate for undercooked chicken meat was 8.1%, similar to that reported elsewhere (3%–11%) (4,5,9). No other types of undercooked meat that were measured in our study (e.g., pork, lamb, and beef) were significantly associated with *Campylobacter* infection.

For diseases with multiple risk factors, the PAR estimate for any single factor should be adjusted for possible confounding and interaction by these other factors (25,26). Multivariable adjustment methods that use logistic regression allow estimates of PAR to a single factor while simultaneously adjusting for other factors in the model. However, if all relevant factors are not included in the model or the model does not have correct parametric form, the adjusted estimates of PAR may be biased (27).

The use of simulation techniques provides a simple but robust approach to accommodate asymmetric component distributions and account for uncertainty in our final estimates of the magnitude of foodborne *Campylobacter* infection in the community. Rather than calculate a single point estimate for the number of cases attributable to each foodborne risk factor, a simulated distribution of credible values was generated to model the uncertainty for each component in our calculations. Generating 95% CIs enabled us to confer a degree of confidence around our estimates.

Intercountry comparison of foodborne disease incidence is difficult without standardization of methods; however, the approach taken in this study may allow those countries that have the available data to conduct similar

studies. Furthermore, this model could be adopted or applied more widely to other foodborne and nonfoodborne pathogens under surveillance and enable calculation of population estimates of the magnitude of infection associated with specific risk factors.

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Technical Appendix

METHODS

Development of the multivariable logistic regression model

Initially, separate univariable analyses were performed on all study variables within each exposure group to generate crude odds ratios (OR) with 95% confidence intervals (CI). Exposure variables significantly associated with *Campylobacter* infection in these crude analyses were considered as candidate variables for development of the multivariable model. Using sequential backward elimination of non-significant variables (based on the model deviance statistic), multivariable logistic regression models were then constructed for each exposure group after controlling for state, sex and education. Confounders sex and education were identified from a separate multivariable logistic model of demographic variables and state was considered a design variable (Table 2A). Again using backward elimination, an omnibus multivariable (main-effects) model was then constructed using all significant exposure variables derived from each of the separate multivariable exposure group models as candidate variables and controlling for state, sex and education. Finally, once the most parsimonious multivariable model was identified, two-factor interactions were introduced into the model and backward elimination of non-significant terms were undertaken (based on the model deviance statistic) until the final model was ascertained. The two-factor interactions considered were based on biological plausibility and prior knowledge from the literature. The Hosmer-Lemeshow goodness-of-fit test was performed on all multivariable models to check model adequacy. SPSS (SPSS Version 11.0; SPSS Inc., Chicago) was used for all regression analyses and a significance level of $\alpha = 0.05$ was used to define statistical significance. To reduce the risk of a type I error, only significant variables were included in the multivariable logistic regression models. A detailed description of the analytical approach and pursuant results has been published elsewhere (7).

Simulation methods

To calculate the proportion of campylobacteriosis that occur among persons aged five years and older in Australia, Australian notification data for the years 2001 to 2003 was reviewed (12). The yearly proportions for cases aged 5 years and older among all notified cases reported by the National Notifiable Diseases Surveillance System (NNDSS) between 2001 and 2003 were 84.3%, 85.1% and 87.4% respectively.

Simulations of size 1,000,000 were undertaken in SAS (SAS Institute Inc. *The SAS System for Windows (9.1)*. Cary, N.C, USA) to estimate the total number of *Campylobacter* infections attributable to each specific risk factor identified in the final multivariable model using the following steps:

1. *Total Campylobacter case numbers (N_j)*. We assumed that 223,000 (95% CrI: 94,000, 363,000) cases of campylobacteriosis occur in Australia in a typical year (3). As this distribution is asymmetrical about its mean, a power transformation of 7/8 was applied (removing the asymmetry in the 95% CrI), 1,000,000 random variates generated, and then these variates were back-transformed to the original scale. We denote these back-transformed variates as N_j for $j=1, \dots, 1,000,000$.

2. *Eligible Campylobacter case numbers* (n_j). The total campylobacteriosis case numbers variates (N_j) represent expected cases for the whole population and need to be adjusted by the ≥ 5 years of age eligibility criterion. As the proportion of cases aged 5 years and older in the NNDSS between 2001 and 2003 was strongly time dependent, the simulated total campylobacteriosis case numbers (n_j) were partitioned into three separate one-year periods and eligible (i.e., ≥ 5 years of age) campylobacteriosis case numbers variates were created by sampling from binomial distributions given by:
- $n_j \sim \text{Binomial}(p=0.843, N_j)$ for $j=1, \dots, 333,333$ (i.e. 2001)
 - $n_j \sim \text{Binomial}(p=0.851, N_j)$ for $j=333,334, \dots, 666,666$ (i.e. 2002)
 - $n_j \sim \text{Binomial}(p=0.874, N_j)$ for $j=666,667, \dots, 1,000,000$ (i.e. 2003)
- based on the proportions reported in the NNDSS data (12).

3. *Population attributable risks (PAR) values*. Category-specific attributable risk proportion (PAR_i) was estimated by:

$$PAR_i = \frac{p_i(aOR_i - 1)}{aOR_i} \times 100\%, \quad (i)$$

where: aOR_i is the i^{th} category-specific adjusted odds ratio calculated from the logistic regression model and p_i is the proportion of all study cases falling into i^{th} exposure level for a categorical variable with k levels and reference category denoted by $i = 1$. The total population attributable risk proportion (PAR) is given by:

$$PAR = \sum_{i=2}^k PAR_i \quad (ii)$$

As the distribution of aOR_i is log-normal, PAR_i values for each category level i , $i > 1$, were derived in the following manner. Simulated $\log(aOR_i)$ values were randomly generated from a normal distribution with mean and standard deviation estimates derived from the i^{th} exposure category of the risk factor under investigation in the final multivariable logistic regression model, $i=2, \dots, k$. These generated $\log(aOR_i)$ values were exponentiated, producing aOR_i values. The proportion of people within each of the i^{th} exposure categories, denoted by p_i , was generated from a binomial distribution via: $x_i \sim \text{Binomial}(q_i, m)$ where q_i is the proportion of cases in the i^{th} exposure category, m is the number of cases with non-missing data, and $p_i = x_i / m$. Simulated PAR_i and PAR values were then derived by combining the generated aOR_i and p_i given by equations (i) and (ii) above. This process was repeated $j=1, \dots, 1,000,000$ times.

4. *Attributable Campylobacter case numbers*. Finally, eligible campylobacteriosis case numbers and PAR simulated values derived in Steps 2 and 3 above were multiplied together to produce distributions of the total number of *Campylobacter* infections attributable to each specific risk factor. Because some distributions are skewed, we present medians and 95% credible intervals (defined to be the 2.5 and 97.5 percentiles) for the simulation results.

RESULTS

Table 1A. Results of multivariable logistic regression analysis for variables within each exposure group (adjusted for state, sex and education), and the first model of the multivariable (main effects) model showing Beta-coefficients, Standard Errors (S.E.), Wald Statistics and Hosmer-Lemeshow goodness-of-fit statistics.*

| Exposure group / variables | Multivariable logistic regression analysis (Exposure groups) | | | | First multivariable (main effects) model | | | |
|---|--|------|----------------|---------|--|------|----------------|---------|
| | Beta-coefficient | S.E. | Wald statistic | P value | Beta-coefficient | S.E. | Wald statistic | P value |
| 1) Meat, poultry and seafood | | | | | | | | |
| No chicken | | | | | | | | |
| Chicken – cooked | 0.27 | 0.15 | 3.20 | 0.07 | -0.03 | 0.26 | 0.01 | 0.91 |
| Chicken – undercooked | 1.48 | 0.27 | 29.55 | <0.001 | 1.55 | 0.46 | 11.43 | 0.001 |
| Offal | 0.72 | 0.33 | 4.86 | 0.03 | 0.66 | 0.51 | 1.67 | 0.20 |
| Fresh fish | -0.44 | 0.12 | 13.73 | <0.001 | -0.54 | 0.20 | 7.66 | 0.006 |
| Hosmer-Lemeshow goodness-of-fit: $P = 0.70$ | | | | | | | | |
| 2) Eggs and dairy products | | | | | | | | |
| Homemade foods containing raw eggs | -0.72 | 0.21 | 11.23 | 0.001 | -0.35 | 0.37 | 0.90 | 0.34 |
| Hosmer-Lemeshow goodness-of-fit: $P = 0.56$ | | | | | | | | |
| 3) Produce | | | | | | | | |
| Organic fruit & vegetables | -0.60 | | 9.41 | 0.002 | -0.33 | 0.31 | 1.17 | 0.28 |
| Home grown fruit | -0.71 | 0.19 | | | | | | |
| Vegetable index: | | 0.16 | 20.88 | <0.001 | -0.89 | 0.29 | 9.50 | 0.002 |
| 0 (no vegetables) | | | | | | | | |
| 1 (1–2 vegetables) | -0.36 | 0.18 | 4.15 | 0.04 | 0.01 | 0.32 | 0.002 | 0.99 |
| 2 (3–4 vegetables) | -0.47 | 0.18 | 7.27 | 0.007 | -0.26 | 0.33 | 0.63 | 0.43 |
| 3 (5–6 vegetables) | -1.37 | 0.32 | 18.15 | <0.001 | -0.84 | 0.53 | 2.54 | 0.11 |
| Hosmer-Lemeshow goodness-of-fit: $P = 0.48$ | | | | | | | | |
| 4) Water consumption | | | | | | | | |
| Commercial bottled water | 0.46 | 0.20 | 5.15 | 0.02 | 0.16 | 0.30 | 0.29 | 0.59 |
| Hosmer-Lemeshow goodness-of-fit: $P = 0.52$ | | | | | | | | |
| 5) Food handling practices | | | | | | | | |
| Barbequed cooked meat placed back on original plate used for raw meat | 0.85 | 0.43 | 3.84 | 0.05 | 0.58 | 0.54 | 1.15 | 0.28 |
| Hosmer-Lemeshow goodness-of-fit: $P = 0.97$ | | | | | | | | |
| 6) Animal & pet exposure | | | | | | | | |
| Domestic chickens | | | | | | | | |
| No domestic chicken | | | | | | | | |
| Chicken aged <6 months | 1.65 | 0.63 | 6.82 | 0.009 | 2.11 | 1.17 | 3.26 | 0.07 |
| Chicken aged ≥6 months | 0.28 | 0.25 | 1.21 | 0.27 | 0.05 | 0.51 | 0.01 | 0.92 |
| Domestic dogs | | | | | | | | |
| No dog | | | | | | | | |
| Dog aged <6 months | 1.08 | 0.31 | 12.40 | <0.001 | 0.62 | 0.50 | 1.54 | 0.21 |
| Dog aged ≥6 months | 0.22 | 0.11 | 4.09 | 0.04 | 0.004 | 0.18 | 0.00 | 0.98 |
| Hosmer-Lemeshow goodness-of-fit: $P = 0.76$ | | | | | | | | |

| 7) Host factors | | | | | | | | |
|---|------|------|-------|--------|---|------|------|-------|
| Chronic gastrointestinal condition | 0.69 | 0.19 | 13.42 | <0.001 | 0.94 | 0.34 | 7.69 | 0.006 |
| Liver disease | 1.62 | 0.77 | 4.42 | 0.04 | -0.35 | 1.15 | 0.09 | 0.76 |
| Any immunosuppressive agent / therapy | 1.02 | 0.35 | 8.69 | 0.003 | 0.05 | 0.59 | 0.01 | 0.93 |
| Hosmer–Lemeshow goodness-of-fit: $P = 0.37$ | | | | | Hosmer–Lemeshow goodness-of-fit: $P = 0.84$ | | | |

*S.E., Standard Error of coefficient

Table 2A. Results of multivariable logistic regression analysis of all demographic variables together, showing frequency and sample size (n/N), Beta-coefficients, Standard Errors (S.E.), Wald Statistics, Odds Ratios (OR) together with 95% confidence intervals (CI), and the Hosmer-Lemeshow goodness-of-fit statistic.*

| Exposure group / variables | Cases n/N | Controls n/N | Beta-coefficient | S.E. | Wald statistic | P value | OR | 95%CI |
|--|--------------|-----------------|------------------|------|----------------|---------|-----|----------|
| Sex | | | | | | | | |
| Female | 412/881 | 451/833 | | | | | | |
| Male | 469/881 | 382/833 | 0.24 | 0.11 | 4.90 | 0.03 | 1.3 | 1.0, 1.6 |
| Age Group (years) | | | | | | | | |
| 5–9 | 66/881 | 65/833 | | | | | 1.0 | |
| 10–19 | 94/881 | 94/833 | -0.03 | 0.27 | 0.01 | 0.91 | 1.0 | 0.6, 1.6 |
| 20–29 | 181/881 | 144/833 | 0.07 | 0.24 | 0.09 | 0.77 | 1.1 | 0.7, 1.7 |
| 30–59 | 395/881 | 385/833 | -0.12 | 0.21 | 0.29 | 0.59 | 0.9 | 0.6, 1.4 |
| 60+ | 145/881 | 145/833 | 0.05 | 0.25 | 0.04 | 0.84 | 1.1 | 0.6, 1.7 |
| Urban / rural residence | | | | | | | | |
| Urban/town | 738/854 | 695/826 | | | | | | |
| Rural/remote | 116/854 | 131/826 | -0.19 | 0.16 | 1.39 | 0.24 | 0.8 | 0.6, 1.1 |
| Education level | | | | | | | | |
| Primary / Secondary to Year 10 | 89/801 | 115/775 | | | | | 1.0 | |
| Secondary Years 11&12 or Apprenticeship | 290/801 | 249/775 | 0.36 | 0.19 | 3.71 | 0.05 | 1.4 | 1.0, 2.1 |
| Certificate / Diploma | 183/801 | 176/775 | 0.25 | 0.20 | 1.56 | 0.21 | 1.3 | 0.9, 1.9 |
| University degree | 239/801 | 235/775 | 0.11 | 0.21 | 0.29 | 0.59 | 1.1 | 0.8, 1.7 |
| Household income | | | | | | | | |
| <\$25,000 | 141/708 | 174/713 | | | | | 1.0 | |
| \$25,000–\$50,000 | 206/708 | 215/713 | 0.12 | 0.18 | 0.46 | 0.50 | 1.1 | 0.8, 1.6 |
| \$50,001–\$100,000 | 266/708 | 236/713 | 0.29 | 0.20 | 2.13 | 0.14 | 1.3 | 0.9, 2.0 |
| >\$100,000 | 95/708 | 88/713 | 0.28 | 0.24 | 1.36 | 0.24 | 1.3 | 0.8, 2.1 |
| Health Care / Pensioner Concession card | | | | | | | | |
| Indigenous person | 279/848 | 307/824 | -0.04 | 0.15 | 0.07 | 0.79 | 1.0 | 0.7, 1.3 |
| Season | 19/852 | 22/824 | -0.27 | 0.36 | 0.57 | 0.45 | 0.8 | 0.4, 1.6 |
| Season | | | | | | | | |
| Spring | 95/881 | 105/833 | | | | | 1.0 | |
| Summer | 153/881 | 139/833 | 0.36 | 0.23 | 2.35 | 0.13 | 1.4 | 0.9, 2.3 |
| Autumn | 320/881 | 308/833 | 0.25 | 0.22 | 1.25 | 0.26 | 1.3 | 0.8, 2.0 |
| Winter | 313/881 | 281/833 | 0.37 | 0.23 | 2.46 | 0.12 | 1.4 | 0.9, 2.3 |

| | | | | | | | | | |
|-------|---------|---------|---|------|------|------|-----|----------|--|
| State | | | | | | | | | |
| Vic | 170/881 | 157/833 | | | | | | 1.0 | |
| Qld | 196/881 | 179/833 | -0.03 | 0.19 | 0.02 | 0.89 | 1.0 | 0.7, 1.4 | |
| SA | 215/881 | 182/833 | 0.14 | 0.19 | 0.53 | 0.47 | 1.2 | 0.8, 1.7 | |
| WA | 134/881 | 137/833 | -0.08 | 0.19 | 0.19 | 0.66 | 0.9 | 0.6, 1.3 | |
| Tas | 166/881 | 178/833 | -0.15 | 0.19 | 0.59 | 0.44 | 0.9 | 0.6, 1.3 | |
| | | | Hosmer-Lemeshow goodness-of-fit: $P = 0.96$ | | | | | | |

*S.E., Standard Error of coefficient; OR, odds ratio; CI, confidence interval.