
European Perspective of 2-Person Rule for Biosafety Level 4 Laboratories

To the Editor: Recently, the directors of Biosafety Level 4 (BSL-4) laboratories in the United States published their views of the requirement of having ≥2 persons present at all times while biological work is undertaken in a BSL-4 laboratory (1). They concluded that safety and security would be better assured in some situations by video monitoring systems rather than by the presence of a fellow scientist. As members of the European Network of Biosafety Level-4 laboratories (Euronet-P4) who have developed guidelines in this area (2–4), we discussed the article during a recent network meeting. Biosafety and biosecurity are the major concerns for all involved in BSL-4 activities, and we support the authors’ initiative and broadly agree with their position. The consensus among European BSL-4 experts is that, in the interest of safety, standard practice should be for all laboratories to perform a risk assessment before any activity is undertaken. This preliminary assessment is the best way to determine procedures to be used, including whether 2 persons should work together as part of the laboratory procedure. A 2-person rule is inappropriate simply because the best approach is not to have inflexible rules that are not objectively assessed according to laboratory-specific circumstances.

Surveillance video monitoring and data storing have their place in protecting laboratory facilities from unauthorized access and theft of materials, but their effectiveness for ensuring proper handling of pathogens is quite limited. Finally, we agree with the authors that both biosafety and biosecurity must be founded on careful selection and monitoring of staff, without which even the most sophisticated control systems would fail.


Author affiliations: National Institute for Infectious Diseases, Rome, Italy (G. Ippolito, C. Nisii, A. Di Caro, M.R. Capobianchi); Health Protection Agency, London, UK (D. Brown, R. Gopal); Health Protection Agency, Salisbury, UK (R. Hewson, G. Lloyd); Bernhard Nocht Institute for Tropical Medicine, Hamburg, Germany (S. Gunther); Institute of Virology, Marburg, Germany (M. Eickmann); Swedish Institute for Infectious Disease Control, Solna, Sweden (A. Mirazimi, T. Koivula); and French National Institute for Health and Medical Research, Lyon, France (M.-C. Georges Courbot, H. Raoul)

DOI: 10.3201/ai1511.091134

References


Multidrug-Resistant Mycobacterium tuberculosis Strain from Equatorial Guinea Detected in Spain

To the Editor: Eleven years of molecular epidemiologic data allowed the Spanish Multidrug-resistant Tuberculosis (MDR TB) Surveillance Network to identify a specific MDR Mycobacterium tuberculosis strain that had been imported into Spain from Equatorial Guinea (1). Our study brings to light the potential dissemination of this strain (named MDR-TBEG) in Equatorial Guinea, a country where little is known about the extent and features of TB or MDR TB. It also highlights that MDR strains can spread across continents, and thus MDR TB’s emergence in any country becomes a global problem.

Ten MDR M. tuberculosis isolates obtained from 10 patients from Equa-
itorial Guinea were detected in Spain during 2000 through 2008. Evidence of clonality was found within the 10 isolates because all exhibited identical genetic profiles defined by different molecular epidemiology methods (2,3) and mutations involved in drug resistance (Figure). Notably, none of the remaining 504 MDR isolates in the Spanish database matched SIT177, a spoligotype belonging to the Latin American–Mediterranean (LAM9) subfamily (4).

The data routinely collected for all cases of MDR TB have been previously described (1). All 10 patients in the study were from Equatorial Guinea, a small African country on the Gulf of Guinea with a population of ≈500,000, an MDR TB rate >2.0% (3) of all combined (new and previously treated) TB cases, and an estimated adult HIV prevalence rate of 3.2% (www.who.int/globalatlas/predefinedReports/EFS2008/full/EFS2008_GQ.pdf). The MDR TB isolates were collected within a 9-year period (online Technical Appendix, available from www.cdc.gov/EID/content/15/11/1858-techapp.pdf): 1 in 2000, 2 in 2001, 3 in 2003, 1 in 2004, 2 in 2007, and 1 in 2008. According to their hospitals of origin, the patients were geographically dispersed in 6 different Spanish cities. We found that the interval between the patients’ arrival in Spain to the initiation of anti-TB treatment was <3 months in 6 patients, 3 of whom were clinically ill at the time of arrival. Seven patients were adult men, 2 were adult women, and 1 was an 8-year-old girl. The patients’ mean age was 30 years (range 8–54 years). Three patients were seropositive and 4 were seronegative for HIV infection (the HIV status of 3 patients was unknown). Data on prior anti-TB treatment was available for 7 case-patients, of whom only 1 had a history of antecedent TB chemotherapy. Altogether, 3 patients died before completing treatment, including 2 patients affected by miliary TB, 1 of whom was HIV-infected. The third patient who died was a student without a known history of immunosuppression or previous TB chemotherapy. Altogether, 3 patients died during their stay in Spain. We could not establish any epidemiologic links between these patients during their stay in Spain.

Analysis of drug resistance genes showed that all isolates harbored the inhA promoter mutation −15C→T (6). Alterations in the inhA gene were previously reported in 80% of the isoniazid-resistant isolates from Equatorial Guinea (5). Notably, a double mutation in the rpoB gene affecting codons 531 (Ser531Leu) and 561 (Ile561Val) was detected in the 10 MDR isolates. The presence of this uncommon mutation, Ile561Val, outside the rifampin resistance–determining region supports the hypothesis that the MDR isolates are clonal in origin. Furthermore, we demonstrated the absence of Ile561Val mutation in 3 drug-susceptible M. tuberculosis strains with an SIT177-LAM9 spoligotype pattern, which ruled out a relationship between this spoligotype and the Ile561Val mutation.

Further analysis with phylogenetic markers assigned MDR-TBEG to the principal genetic group 2, the Euro-American lineage of M. tuberculosis and its West African sublineage, on the basis of polymorphisms in codons katG463 and gyrA95, the 7-bp pk515/1 deletion, and RD174 (7,8), respectively. The analysis of the RD86 deletion confirmed that the strain belongs to the major RD86 sublineage of the LAM M. tuberculosis spoligotype family (9). This sublineage is a major cause of TB in Rio de Janeiro (Brazil) but has disseminated globally. Additional information on the geographic distribution of SIT177-LAM9 was obtained from the updated International Spoligotyping Database (SITVIT2) of the Institut Pasteur de Guadeloupe. SITVIT2 (consulted on 23 July 2008) contained 57 isolates belonging to SIT177. Almost 50% (n = 28) came from Brazil, and 14% from Africa (Morocco, n = 6; Senegal, n = 2). The remaining isolates with known countries of origin (n = 9) were distributed in other unrelated countries. These data indicate that this particular spoligotype pattern is widely distributed.

We identified 1 MDR strain of M. tuberculosis RD86 sublineage isolated in Spain from Equatorial Guinean patients. Although the transmission of MDR-TBEG in Spain could not be conclusively ruled out, the fact that MDR TB developed in most patients
within 3 months after their arrival, as well as the spatiotemporal distribution of the MDR TB cases and its clonal origin, strongly suggest that MDR-TBEG was imported into Spain and that active transmission of this particular clone could be occurring in Equatorial Guinea. However, additional molecular and epidemiologic studies should be conducted in this sub-Saharan country to ascertain its role in recent transmission of MDR TB. Greater international efforts should be made to provide appropriate tools to resource-limited areas for fighting against MDR TB and preventing development of extensively drug-resistant TB.

Acknowledgments

We thank Dessi Vaneva Marinova for assistance in writing the manuscript, and Alberto Cebollada, Carmen Lafoz, Ana Picó, and Daniel Ibarz for their excellent technical assistance. We are grateful to Thierry Zozio for helping with the geographic distribution of SIT177 in the International Spoligotyping Database (SITVIT2). Inquiries regarding the SITVIT2 should be addressed to nrastogi@pasteur-guadeloupe.fr.

This work was supported by the Spanish Fondo de Investigación Sanitaria (FIS nos. 06/1624, 03/0743 and 01/3088), CIBERES, and the Instituto de Salud Carlos III-Instituto Aragonés de Ciencias de la Salud (CM06/00100).

Patricia Gavín, María J. Iglesias, María S. Jiménez, Laura Herrera-León, Elina Rodríguez-Valín, Nalin Rastogi, Josefa March, Rosa González-Palacios, Elia Palenque, Rafael Ayarza, Elena Hurra, Isolina Campos-Herrero, María A. Vitoria, María A. Lezcano, María J. Revillo, Carlos Martín, and Sofía Samper

Author affiliations: Instituto Aragonés de Ciencias de la Salud, Zaragoza, Spain (P. Gavín, S. Samper); Hospital Universitario Miguel Servet, Zaragoza (P. Gavín, S. Samper, M.A. Lezcano, M.J. Revillo); Centro de Investigación Biomédica en Red Enfermedades Respiratorias, Madrid, Spain (P. Gavín, S. Samper, M.J. Iglesias, C. Martín, M.A. Lezcano, M.A. Vitoria, M.J. Revillo); Universidad de Zaragoza, Zaragoza (M.J. Iglesias, C. Martín); Instituto de Salud Carlos III, Madrid (M.S. Jiménez, J. March, L. Herrera-León, E. Rodríguez-Valín); Centro de Investigación Biomédica en Red de Epidemiología y Salud Pública, Madrid (E. Rodríguez-Valín); Institut Pasteur, Guadeloupe, France (N. Rastogi); Hospital Universitario Príncipe de Asturias, Alcalá de Henares, Madrid (R. González-Palacios); Hospital 12 de Octubre, Madrid (E. Palenque); Hospital Galdakao-Usansolo, Galdacano, Vizcaya, Spain (R. Ayarza); Hospital de Cruces, Baracaldo, Vizcaya, (E. Hurra); Hospital Universitario de Gran Canaria Doctor Negrín, Las Palmas de Gran Canaria, Spain (I. Campos-Herrero) and Hospital Clínico Universitario, Zaragoza (M.A. Vitoria)

DOI: 10.3201/eid1511.090449

References


Address for correspondence: Patricia Gavín, Laboratorio de Investigación Molecular, Consultas Externas Planta 4ª, Hospital Universitario Miguel Servet, Calle Cardenal Gómá sn, Zaragoza 50009, Spain; email: pgavinb.iacs@aragon.es
Multidrug-Resistant *Mycobacterium tuberculosis* Strain from Equatorial Guinea Detected in Spain

**Technical Appendix**

Table. Demographic and clinical characteristics of 10 patients from Equatorial Guinea infected with multiple drug resistant tuberculosis Equatorial Guinea strain*

<table>
<thead>
<tr>
<th>Patient no. †</th>
<th>Age, y/sex‡</th>
<th>Date of arrival in Spain</th>
<th>Date of treatment initiation</th>
<th>City of isolation</th>
<th>Resistance pattern§</th>
<th>TB type</th>
<th>Sputum smear¶</th>
<th>History of prior TB treatment</th>
<th>HIV serology</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>23/F</td>
<td>2000 Sep</td>
<td>2000 Nov</td>
<td>Madrid</td>
<td>HR</td>
<td>Pulmonary</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>2</td>
<td>29/F</td>
<td>2000 May</td>
<td>2000 Jul 20</td>
<td>Madrid</td>
<td>HRES</td>
<td>Pulmonary</td>
<td>Positive</td>
<td>No</td>
<td>NA</td>
</tr>
<tr>
<td>3</td>
<td>8/F</td>
<td>EG</td>
<td>NA</td>
<td>Barakaldo</td>
<td>HRZ</td>
<td>Pott’s disease</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>4</td>
<td>54/M</td>
<td>2002 Nov</td>
<td>2003 Jan</td>
<td>Madrid</td>
<td>HRES</td>
<td>Miliary pattern</td>
<td>Positive</td>
<td>No</td>
<td>Negative</td>
</tr>
<tr>
<td>5</td>
<td>21/M</td>
<td>2001 Jun</td>
<td>2003 May</td>
<td>Las Palmas de Gran Canaria</td>
<td>HREta</td>
<td>Pulmonary</td>
<td>Positive</td>
<td>No</td>
<td>Negative</td>
</tr>
<tr>
<td>6</td>
<td>30/M</td>
<td>2002 Nov</td>
<td>2003 Oct</td>
<td>Alcalá de Henares</td>
<td>HRZEta</td>
<td>Miliary pattern</td>
<td>Positive</td>
<td>NA</td>
<td>Positive</td>
</tr>
<tr>
<td>7</td>
<td>27/M</td>
<td>1994</td>
<td>2004 Jun</td>
<td>Madrid</td>
<td>HR</td>
<td>Lymph node</td>
<td>No</td>
<td>Negative</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>49/M</td>
<td>EG</td>
<td>2008 Jan</td>
<td>Zaragoza</td>
<td>HR</td>
<td>Pulmonary</td>
<td>Positive</td>
<td>No</td>
<td>Positive</td>
</tr>
<tr>
<td>9</td>
<td>41/M</td>
<td>EG</td>
<td>2007 Sep</td>
<td>Alcalá de Henares</td>
<td>HREClaEtaRfb</td>
<td>Pulmonary</td>
<td>Negative</td>
<td>Yes</td>
<td>Positive</td>
</tr>
<tr>
<td>10</td>
<td>24/M</td>
<td>By the end of 2002</td>
<td>2008 Mar</td>
<td>Galdakao</td>
<td>HREta</td>
<td>Pulmonary</td>
<td>Positive</td>
<td>No</td>
<td>Negative</td>
</tr>
</tbody>
</table>

*NA, not available; EG, resident of Equatorial Guinea referred to a Spanish hospital for diagnosis and treatment.
†All patients were born in Equatorial Guinea.
‡Age in years.
§H, isoniazid; R, rifampin; E, ethambutol; S, streptomycin; Z, pyrazinamide; Eta, ethionamide; Cla, clarithromycin; Rfb, rifabutin.
¶Sputum smear not applicable in Pott’s disease or lymph node.