To determine whether HIV-1 infection and HIV-1–related immunosuppression were risk factors for severe malaria in adults with some immunity to malaria, we conducted a case–control study in Luanshya, Zambia, during December 2005–March 2007. For each case-patient with severe malaria, we selected 2 matched controls (an adult with uncomplicated malaria and an adult without signs of disease). HIV-1 infection was present in 93% of case-patients, in 52% of controls with uncomplicated malaria, and in 45% of asymptomatic controls. HIV-1 infection was a highly significant risk factor for adults with severe malaria compared with controls with uncomplicated malaria (odds ratio [OR] 12.6, 95% confidence interval [CI] 2.0–78.8, p = 0.0005) and asymptomatic controls (OR 16.6, 95% CI 2.5–111.5, p = 0.0005). Persons with severe malaria were more likely to have a CD4 count <350/μL than were asymptomatic controls (OR 23.0, 95% CI 3.35–158.00, p<0.0001).

The geographic overlap between HIV-1 infection and malaria, particularly in eastern and southern Africa, has caused concern since the 1980s. The degree of interaction between HIV-1 infection and malaria emerged during 1999–2009 and has been extensively reviewed for both nonpregnant and pregnant adult women (1,2). The effect of HIV-1 on malaria seems to be driven mainly by the incapacity of the immune system to control parasite load, leading to a higher prevalence of infection (3), a higher incidence of clinical malaria (4,5), and a risk for treatment failure (6) in immunosuppressed HIV-1 patients.

Reports of HIV-1 infection as a risk factor for hyperparasitemia or severe malaria are few and limited. In urban Burkina Faso, >30% of adults with severe malaria were also infected with HIV-1, whereas HIV-1 prevalence in the general adult population was ≈5%–14% (7). In South Africa, in an area of low malaria transmission (<1 case/1000/
year), HIV-1 infection was associated with severe falciparum malaria (8–10). Similarly, in Mumbai, India, where malaria transmission is low, HIV-1 prevalence was higher in persons with severe malaria than in the general population (11). Therefore, in areas of low malaria transmission, HIV-1 infection seems to be an important risk factor for severe malaria and death. In areas of high malaria transmission (>1/case/1000/year), the relationship between HIV-1 infection and severe malaria is less well established, with a small hospital-based study in Zimbabwe reporting higher risk for severe malaria and related death in HIV-1–infected adults than in HIV-1–uninfected adults (12). In all these studies, either the number of cases was small or HIV-1 testing was not performed, and CD4 cell count was not routinely done. The effect of both HIV-1 infection and malaria is important in several African countries with high malaria endemicity. We report the results of a matched case–control study exploring whether HIV-1 is an important risk factor for severe malaria in adults living Luanshya, Zambia, an area of high malaria transmission (>200/cases/year).

Methods

Study Design

We chose a case–control design for this study because severe malaria is relatively rare in areas of stable transmission (13). Each adult with severe malaria was matched with 2 controls: 1 adult who had uncomplicated malaria and 1 asymptomatic adult in the community. We matched for major confounding variables: age group (15–19, 20–29, 30–39, and 40–49 years), sex, area of residence, and seasonal variation. Controls were recruited within 4 weeks after identification of case-patients.

Study Site and Patients

The study was conducted at Thompson Hospital, a government district hospital serving Luanshya district in the Copperbelt province of Zambia. In this district, 99% of the population is at risk for malaria (Figure), and 30% of women attending voluntary counseling and testing at the antenatal care department are infected with HIV-1 (15). Luanshya is not a site of sentinel surveillance for HIV, but sentinel surveillance conducted during late 2004 and early 2005 in the adjacent district of Ndola showed an HIV prevalence rate of 30% (16). From October 2005 through April 2007, all patients 15–49 years of age who sought treatment for symptoms and signs of febrile illness were screened for falciparum malaria by thick and thin blood smear. Patients in whom severe malaria was diagnosed were enrolled after informed consent was given by the patient or by his or her legally authorized representative. We defined severe malaria (16) as fever (body temperature ≥37.5°C) or history of fever in the previous 48 hours, Plasmodium falciparum monoinfection with a density of at least 100 parasites per 200 leukocytes (assumed to be ≥4,000 parasites/μL) in the absence of any other evident causes of illness, and at least 1 of the following signs: impaired consciousness (Glasgow Coma Score <10), multiple grand mal convulsions, jaundice, hypoglycemia (glycemia ≤2.5 mmol/L), hyperparasitemia (parasite density >100,000/μL), renal impairment, and cardiorespiratory distress. We collected information about anemia but did not include anemia in the case definition because it can be HIV-1 related, resulting in possible bias, and because uncomplicated and severe malaria episodes can be misclassified in HIV-1–infected persons.

Patients with severe malaria were treated according to the national guidelines for severe malaria in Zambia (i.e., quinine 10 mg/kg/8 h for 7 d), with adequate supportive treatment (17). On the basis of information about areas of unstable malaria and of HIV surveillance data in Zambia, we predicted that HIV-1 increased the risk for severe malaria 5-fold and assumed a 30% HIV-1 prevalence (primary exposure) among controls. Using a 1:1 case:control ratio, we determined that we needed to recruit 30 case–control pairs to detect this risk with 80% power. Recruitment of cases continued until the required sample size was reached.

Controls with uncomplicated malaria were selected from the clinic closest to the homes of the case-patients. The first patient attending the clinic and exhibiting fever or having a history of fever in the previous 48 hours, without any other disease, with a positive rapid diagnostic test, residing in the catchment area of the health center, matching sex and...
age group of the case-patient, and willing to participate was recruited as a control. Pregnant women were excluded.

Field trips were organized to case-patients’ residences, where, following the random sampling method, a community control was identified and independently assessed. The first healthy asymptomatic adult fulfilling the matching criteria and willing to participate was recruited as a community control. Pregnant women were excluded. All community controls were assessed for possible confounding factors and screened for falciparum malaria by thick and thin blood smear. We sought to identify controls within 4 weeks after we recruited a case-patient.

Laboratory Tests

Laboratory technicians were blinded to patient identity and to patient-related parameters. We used microscopy to screen case-patients and community controls for malaria using Giemsa staining 10% for 5 min. A thin blood film was examined to identify type of parasite; a thick film was taken for quantification. Computation of parasite density took into consideration the actual leukocyte count. In asymptomatic controls, we screened for P. falciparum infection by using a rapid diagnostic test, Malaria Pf immunochromatographic test (ICT) (ICT Diagnostics, Cape Town, South Africa). For each participant, a blood sample (0.5 mL) was collected for HIV-1 testing and CD4 cell count, and for hemoglobin (1 mL), by venipuncture. Neither the study staff nor the patient had access to the HIV-1 test results. HIV-1 testing followed an unlinked anonymous procedure: blood samples were sent to the hospital laboratory, where they were processed anonymously. Determine HIV1/2/O (Abbott Laboratories, Abbott Park, IL, USA) was the first test. If test results were negative, the patient was considered not infected. If results were positive or indeterminate, the blood sample was tested with Unigold Recombinant HIV-1/2 (Trinity Biotech PLC, Bray, Ireland); if results were positive, the patient was classified as HIV-1 infected. If results were negative, the outcome was considered indeterminate, and the sample was further tested with ELISA and Western blot. Patients, after recovering from malaria, and controls were counseled about HIV-1 and offered the opportunity for voluntary counseling and testing.

CD4 counts were determined by flow cytometry on a CyFlow (Cyflow Counter, Partec, Germany) within 5 hours after collection, and multiset software was used to obtain the absolute counts and the CD4+ lymphocyte ratio (18). A FACSCount machine (Becton Dickinson, Sparks, MD, USA) was used as a quality control to validate the accuracy of the Cyflow data over time and served as backup.

Ethics and Consent

The trial was approved by the ethical committee of the Institute of Tropical Medicine, Antwerp, Belgium, and by the Research Ethics Committee of the Tropical Disease Research Centre, Ndola, Zambia. The written informed consent signed by study participants or their legally authorized representatives described the purpose of the study, procedures followed, and risks and benefits of participation. The consent form was in both English and the local language. Patients were counseled on HIV and offered the opportunity to undergo voluntary counseling and testing in conjunction with the study; this procedure was followed to ensure adherence to the standard of counseling and testing stipulated in the guidelines for HIV/AIDS counseling in Zambia (17).

Statistical Analysis

Crude associations between the primary outcome measures and potential risk factors, including HIV-1 status and HIV-1–related immunosuppression (HIV-1+ with CD4 ≤350/μL and HIV-1+ with CD4 >350/μL), were described by using summary statistics, means, and count and were tested by using Wilcoxon signed rank test for matched variables.

Results

From December 2005 through March 2007, we recruited 30 case-patients with severe malaria, 30 controls with uncomplicated malaria, and 30 asymptomatic community controls who fulfilled all inclusion criteria. Because of the low malaria incidence among adults, 5 controls with uncomplicated malaria who met the matching criteria could not be identified within 28 days after admission of the case-patient to the hospital. One case-patient and corresponding matched controls were omitted because of missing laboratory results. One case-patient was recruited during March–October 2006, the low malaria transmission season. Median age was 33 years, and men and women were equally represented (Table 1). Eighty-six percent of participants lived in concrete houses. Households contained an average of 4.7 inhabitants, few of whom slept under a bed net (1.5 persons per household, 32%). Only 24% of case-patients (7/29) had slept under a bed net before admission.

Impaired consciousness and hypoglycemia were the most common signs of severe malaria in HIV-1–infected case-patients (Table 2). Nineteen percent of patients, all HIV-1 infected, had anemia. All parasite densities were above the fever threshold (2,088/μL–635,500/μL), conservatively set at 2,000 parasites/μL. Seven of 13 patients with a parasite density below the geometric mean were treated with quinine only; others conservatively received some antimicrobial drugs. Six case-patients, all HIV-1–infected, had hyperparasitemia, 5 with >200,000 parasites/μL. None of the case-patients were receiving antiretroviral drugs or cotrimoxazole prophylaxis. Fifteen case-patients received concomitant antimicrobial drug treatment(s) determined
by clinical symptoms: penicillin (8 patients), amoxicillin (1), gentamicin (2), metronidazole (1), flucanazole (2), chloramphenicol (3), ciprofloxacins (1), and cotrimoxazole (4). Clinical history, symptoms, outcome, illness duration, and other relevant parameters were similar in case-patients who received concomitant antmicrobial drugs and in those who received quinine only. Five (19%) patients died, 4 within 2 days after admission. Median length of hospital stay for successfully treated patients was 5.5 days. Case-patients were more likely to have used antimalarial treatment (45%) during the week before admission than were controls with uncomplicated malaria (28%) and asymp- tomatic controls (17%) (Table 1). Three case-patients used antimicrobial agents the week before admission, compared with 2 controls with uncomplicated malaria and 1 asymptomatic control.

Table 1. Risk factors for severe malaria in case–control study, Luanshya, Zambia

<table>
<thead>
<tr>
<th>Risk factors</th>
<th>Case-patients, n = 29</th>
<th>Controls with uncomplicated malaria, n = 29</th>
<th>Asymptomatic controls, n = 29</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Demographic characteristics*</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GM† age, y (range)</td>
<td>33 (18–50)</td>
<td>33 (20–49)</td>
<td>33 (18–50)</td>
<td></td>
</tr>
<tr>
<td>No. (%) male</td>
<td>14 (48)</td>
<td>14 (48)</td>
<td>14 (48)</td>
<td>0.13</td>
</tr>
<tr>
<td>Living conditions†</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No. (%) living in mud/clay hut</td>
<td>4 (14)</td>
<td>4 (14)</td>
<td>5 (17)</td>
<td>0.91</td>
</tr>
<tr>
<td>No. (%) living in concrete house</td>
<td>25 (86)</td>
<td>25 (86)</td>
<td>24 (83)</td>
<td></td>
</tr>
<tr>
<td>GM no. persons living in house* (95% CI)</td>
<td>4.7 (3.7–6.0)</td>
<td>3.6 (2.9–4.5)</td>
<td>4.7 (3.8–5.8)</td>
<td>0.76</td>
</tr>
<tr>
<td>Mean no. sleeping under bed net (95% CI)</td>
<td>1.5 (0.7–2.3)</td>
<td>1.5 (0.7–2.3)</td>
<td>1.4 (0.5–2.3)</td>
<td>0.85</td>
</tr>
<tr>
<td>No. (%) sleeping under bed net</td>
<td>7 (24)</td>
<td>9 (31)</td>
<td>9 (31)</td>
<td>0.80</td>
</tr>
<tr>
<td>No. (%) using antimalarial drug during previous week</td>
<td>13 (45)</td>
<td>8 (28)</td>
<td>5 (17)</td>
<td>0.13</td>
</tr>
<tr>
<td>No. using quinine</td>
<td>2</td>
<td>2</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>No. using sulfadinoxine pyrimethamine</td>
<td>8</td>
<td>5</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>No. using artemether–lumefantrine</td>
<td>5</td>
<td>1</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>No. (%) with HIV-1</td>
<td>27 (93)</td>
<td>15 (52)</td>
<td>13 (45)</td>
<td>0.03</td>
</tr>
<tr>
<td>GM CD4 count (95% CI):‡</td>
<td>173 (125–240)</td>
<td>205 (112–377)</td>
<td>677 (427–1074)</td>
<td></td>
</tr>
<tr>
<td>No. (%) CD4 count &lt;200/μL‡</td>
<td>1123 (48)</td>
<td>514 (36)</td>
<td>1/12 (8)</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>No. (%) CD4 count &lt;350/μL‡</td>
<td>1923 (83)</td>
<td>1114 (79)</td>
<td>1/12 (8)</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

*Matching variables.  †GM, geometric mean; CI, confidence interval.
‡Not measured in 4 case-patients, 1 control with uncomplicated malaria, and 1 asymptomatic control because of technical constraints.

Table 2. Clinical features of case-patients with severe malaria in case–control study, by HIV-1 infection status, Luanshya, Zambia

<table>
<thead>
<tr>
<th>Clinical features</th>
<th>HIV-1 infected, n = 27</th>
<th>HIV-1 uninfected,* n = 2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Signs and symptoms, no. (%)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fever (&gt;37.5°C)</td>
<td>20 (74)</td>
<td>1 (50)</td>
</tr>
<tr>
<td>History of fever</td>
<td>25 (93)</td>
<td>2 (100)</td>
</tr>
<tr>
<td>Impaired consciousness (Glasgow Coma Score ≤10)</td>
<td>15 (56)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Severe anemia (&lt;7 g/dL)†</td>
<td>5 (19)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Convulsions</td>
<td>6 (22)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Jaundice</td>
<td>3 (11)</td>
<td>1 (50)</td>
</tr>
<tr>
<td>Hypoglycemia (&lt;2.5 mmol/L)</td>
<td>11 (41)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Hyperparasitemia (≥100,000 parasites/μL)</td>
<td>6 (22)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Renal impairment</td>
<td>0 (0.0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Leukocyte count, mean 1,000/μL (SD)‡</td>
<td>6.9 (3.9)</td>
<td>4.2 and 8.7</td>
</tr>
<tr>
<td>Lymphocytes, % (SD)‡</td>
<td>25 (11)</td>
<td>18</td>
</tr>
<tr>
<td>Monocytes, % (SD)‡</td>
<td>11 (6)</td>
<td>5.2</td>
</tr>
<tr>
<td>Granulocytes, % (SD)‡</td>
<td>62 (20)</td>
<td>76.8</td>
</tr>
<tr>
<td>Parasite density, geometric mean/μL (95% confidence interval)</td>
<td>43,314 (25,467–81,145)</td>
<td>11,745 and 38,942</td>
</tr>
<tr>
<td>Concomitant antimicrobial drugs</td>
<td>15 (56)</td>
<td>0</td>
</tr>
</tbody>
</table>

*Because only 2 results were HIV negative, no SD or confidence interval was specified.
†Anemia was not an inclusion criterion.
‡Not measured in 4 patients because of technical constraints.
Risk Analysis

Case-patients differed from controls in use of other drugs during the previous week, HIV-1 infection, and CD4 count. No parasitemia was detected in any asymptomatic controls. Because of the matching, controls with uncomplicated malaria and asymptomatic controls were similar to case-patients for all other assessed risk factors (Table 1).

HIV-1 infection was detected in 45% of asymptomatic controls, 52% of controls with uncomplicated malaria, and 93% of case-patients. HIV-1 infection was not a risk factor for uncomplicated malaria (odds ratio [OR] 1.3, 95% confidence interval [CI] 0.5–3.7, p = 0.59) (Table 3). Case-patients were more likely to be infected with HIV-1 than were controls with uncomplicated malaria and asymptomatic controls (OR 12.6, 95% CI 2.0–78.8, p = 0.0005, and OR 16.6, 95% CI 2.5–111.8, p = 0.0005, respectively).

Eighty-three percent of case-patients had a CD4 count <350/μL, compared with 79% of controls with uncomplicated malaria (p = 0.76) and 8% of asymptomatic controls (p <0.0001). Controls with uncomplicated malaria were more likely than asymptomatic controls to have a CD4 count <350/μL (OR 7.67, 95% CI 1.78–33.01, p = 0.001). Case-patients were more likely to have a CD4 count <350/μL than were asymptomatic controls (OR 23.00, 95% CI 3.35–158.00, p <0.0001) but not controls with uncomplicated malaria (OR 3.00, 95% CI 0.83–10.83, p = 0.32).

The extremely high proportion of low CD4 count in both case-patients and controls with uncomplicated malaria might be confounded by a temporary malaria-induced reallocation of specific T-cells (19). Therefore, 28 days after successful treatment, the absolute CD4 count was measured in 10 HIV-infected case-patients. During this period, the mean CD4 count increased >2-fold, from 142 (95% CI 76–269) to 320 CD4/μL (95% CI 169–607) (data not shown). However, the proportion of case-patients with CD4 count <350/μL remained substantial (70%), although slightly less than at admission (90%).

**Table 3. HIV-1 infection and HIV-1-related immunosuppression as risk factors for nonsevere and severe malaria, case–control study, Luanshya, Zambia**

<table>
<thead>
<tr>
<th>Participant characteristics</th>
<th>No. (%)</th>
<th>OR (95% CI)</th>
<th>p value</th>
<th>OR (95% CI)</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>HIV-1 infected†</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Asymptomatic controls</td>
<td>13/29 (45)</td>
<td>1</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Controls with uncomplicated malaria</td>
<td>15/29 (52)</td>
<td>1.3 (0.5–3.7)</td>
<td>0.59</td>
<td>1</td>
<td>–</td>
</tr>
<tr>
<td>Case-patients (severe malaria)</td>
<td>27/29 (93)</td>
<td>16.6 (2.5–111.8)</td>
<td>0.0005</td>
<td>12.6 (2.0–78.8)</td>
<td>0.0005</td>
</tr>
<tr>
<td>CD4 cell count &lt;350/μL‡</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Asymptomatic controls</td>
<td>1/12 (8)</td>
<td>1</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Controls with uncomplicated malaria</td>
<td>11/14 (79)</td>
<td>7.67 (1.78–33.01)</td>
<td>0.001</td>
<td>1</td>
<td>–</td>
</tr>
<tr>
<td>Case-patients (severe malaria)</td>
<td>19/23 (83)</td>
<td>23.00 (3.35–158.00)</td>
<td>&lt;0.0001</td>
<td>3.00 (0.83–10.83)</td>
<td>0.08</td>
</tr>
</tbody>
</table>

†p value obtained by using matched-pairs signed-ranks test.
‡CD4 count not measured in 4 HIV-1–infected case-patients, 1 control with uncomplicated malaria, and 1 asymptomatic control because of technical constraints. p value obtained by using Wilcoxon signed-ranks test and Wilcoxon rank-sum test.

Discussion

In Luanshya, Zambia, an area where malaria is mesoendemic, HIV-1 infection is an important risk factor for severe malaria in adults, primarily in those with a low CD4 count. The increased risk for severe malaria in HIV-1–infected persons already has been reported from areas of low and unstable transmission (8–11) but never has been firmly established in areas of stable malaria transmission. The paucity of information results from the difficulty of obtaining it. Studies collecting relevant information retrospectively are vulnerable to considerable bias, and prospective studies are difficult to carry out where adults have acquired immunity and severe malaria is rare (20). In our study, recruitment of 30 persons with severe malaria from a population of 10,000 persons took 2 years. However, even prospectively, diagnosing severe malaria with certainty might be difficult, particularly in the presence of HIV-1 co-infection, because several opportunistic infections of AIDS patients could have clinical presentations similar to those of severe malaria. Persons with severe malaria could have had a concomitant *Streptococcus pneumoniae* or *Salmonella enterica* serovar Typhimurium infection (21) or HIV-1 immunosuppression-related meningitis, such as cryptococcal meningitis. Although, none of the case-patients we recruited were identified by the treating physicians as having clinical AIDS, in the absence of a blood culture, physicians conservatively decided to concomitantly prescribe antimicrobial drugs in 15 cases. More detailed review of patients’ files showed that most of these persons had hypoglycemia or hyperparasitemia, and none were in septic shock. Moreover, these patients recovered rapidly after treatment, and all but 1 were recruited during the high malaria transmission season, which support the argument that these persons had true cases of severe malaria.

The increased risk for uncomplicated malaria in HIV-1–infected patients with a low CD4 count is consistent with information in several cohort studies (7). Although the extremely high proportion of a low CD4 count in patients...
in our study might have been confounded by a temporary malaria-induced reallocation of specific T cells; 70% had a CD4 count <350/μL 1 month after successful treatment. Therefore, immunosuppression is likely an additional risk, but absolute CD4 count cannot be interpreted during a severe malaria episode.

Almost half of malaria cases throughout the world occur in areas where the disease is holoendemic (22). HIV-1 program managers working in areas where both diseases are prevalent should be aware that HIV-1 infection—and certainly HIV-1–related immunosuppression—are important risk factors for severe malaria. Early detection of HIV-1 infection is extremely important because comprehensive measures to prevent malaria and chemoprophylaxis with cotrimoxazole could be promptly implemented to protect against uncomplicated and severe malaria, a disease with a high fatality rate (23). Clinicians in such settings also should be aware of the strong association between severe malaria and HIV-1 so they can assess patients for other underlying diseases and offer the opportunity for voluntary counseling and testing for HIV-1 when patients have recovered from malaria.

Acknowledgments

We thank the case-patients, controls, and families who contributed to this study. We also thank the staff at Thomson Hospital (Mulenga Peter, Pierre Tshibangu, Margret Makulu, Judy Nalawwe, Jacob Phiri, Tina Kosamu, Maybin Silawwe, and Sara Musukwa) and the staff at the health centers where the study was carried out.

The study was funded by the Belgian Development Cooperation in the framework of an institutional collaboration between the Tropical Diseases Research Centre in Ndola, Zambia and the Institute of Tropical Medicine in Antwerp, Belgium (budget number: 96603).

Dr Chalwe is a medical scientist with the Tropical Diseases Research Centre, Ndola, Zambia. His research interests are poverty-related diseases, particularly malaria and HIV/AIDS.

References


Address for correspondence: Jean-Pierre Van geertruyden, Department of Parasitology, Unit of Epidemiology, Institute of Tropical Medicine Antwerpen, Nationalestraat 155 B2000, Antwerp, Belgium; email: jpvangeertruyden@itg.be
Earning CME Credit

To obtain credit, you should first read the journal article. After reading the article, you should be able to answer the following, related, multiple-choice questions. To complete the questions and earn continuing medical education (CME) credit, please go to [http://www.medscape.com/cme/eid](http://www.medscape.com/cme/eid). Credit cannot be obtained for tests completed on paper, although you may use the worksheet below to keep a record of your answers. You must be a registered user on Medscape.com. If you are not registered on Medscape.com, please click on the New Users: Free Registration link on the left hand side of the website to register. Only one answer is correct for each question. Once you successfully answer all post-test questions you will be able to view and/or print your certificate. For questions regarding the content of this activity, contact the accredited provider, CME@medscape.net. For technical assistance, contact CME@webmd.net. American Medical Association’s Physician’s Recognition Award (AMA PRA) credits are accepted in the US as evidence of participation in CME activities. For further information on this award, please refer to [http://www.ama-assn.org/ama/pub/category/2922.html](http://www.ama-assn.org/ama/pub/category/2922.html). The AMA has determined that physicians not licensed in the US who participate in this CME activity are eligible for [AMA PRA Category 1 Credits™](http://www.ama-assn.org/ama/pub/category/2922.html). Through agreements that the AMA has made with agencies in some countries, AMA PRA credit is acceptable as evidence of participation in CME activities. If you are not licensed in the US and want to obtain an AMA PRA CME credit, please complete the questions online, print the certificate and present it to your national medical association.

**Article Title:**

**Increased Risk for Severe Malaria in HIV-1–infected Adults, Zambia**

**CME Questions**

1. Which of the following best explains the large scale of the HIV-1–malaria interaction that has emerged in the last decade?
   A. Longer survival of patients infected with HIV
   B. Effects of increased antiretroviral drug use
   C. Poor immune function and higher susceptibility
   D. Poor control of malaria worldwide

2. A 28-year-old Zambian patient presents with a fever of 38.5°C, *Plasmodium falciparum* on thick smear with 120 parasites per 200 white blood cells, and jaundice. Which of the following best describes the likely diagnosis?
   A. Uncomplicated malaria
   B. Moderately severe malaria
   C. Severe malaria
   D. HIV and malaria

3. The study noted the importance of fever as an indicator of severe malaria in patients infected with HIV-1. Which of the following features were most commonly encountered in addition to fever?
   A. Impaired consciousness and jaundice
   B. Impaired consciousness and hypoglycemia
   C. Multiple convulsions and jaundice
   D. Hypoglycemia and jaundice

4. Which of the following best describes the association between HIV-1 infection and risk for severe malaria in the population studied?
   A. HIV-1 infection is a risk factor for uncomplicated and severe malaria
   B. Risk for severe malaria is only increased in patients with HIV-1 with a CD4 count <250 cells/μL
   C. HIV-1 infection increases the risk for severe malaria
   D. Risk for severe malaria is increased only in patients with AIDS

**Activity Evaluation**

<table>
<thead>
<tr>
<th>1. The activity supported the learning objectives.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Strongly Disagree</td>
</tr>
<tr>
<td>Strongly Agree</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>2. The material was organized clearly for learning to occur.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Strongly Disagree</td>
</tr>
<tr>
<td>Strongly Agree</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>3. The content learned from this activity will impact my practice.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Strongly Disagree</td>
</tr>
<tr>
<td>Strongly Agree</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>4. The activity was presented objectively and free of commercial bias.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Strongly Disagree</td>
</tr>
<tr>
<td>Strongly Agree</td>
</tr>
</tbody>
</table>