Acute Encephalopathy Associated with Influenza A Infection in Adults


We report acute encephalopathy associated with influenza A infection in 3 adults. We detected high cerebrospinal fluid (CSF) and plasma concentrations of CXCL8/IL-8 and CCL2/MCP-1 (CSF/plasma ratios >3), and interleukin-6, CXCL10/IP-10, but no evidence of viral neuroinvasion. Patients recovered without sequelae. Hyperactivated cytokine response may play a role in pathogenesis.

Influenza-associated acute encephalopathy has been described in children, and results in a high frequency of neurologic sequelae and death. Altered consciousness, disorientation, and seizures occur within a few days after the onset of fever and respiratory symptoms (1–3). In some patients, symptoms are transient but in others rapid progression to necrotizing encephalitis, deep coma, and death may occur (1–3). Cases in adults are infrequently reported and remain poorly characterized, although the more complex clinical scenarios in adults may have hindered case recognition (1–4–6). The pathogenesis is unclear, but a hyperactivated cytokine response, rather than viral invasion, is believed responsible in most childhood cases (1–5). We describe 3 cases of acute encephalopathy associated with influenza A infection in adults. The clinical, virologic, immunologic findings (cytokines in plasma and cerebrospinal fluid [CSF]), and CSF penetration of oseltamivir for these cases are reported.

The Study

At Prince of Wales Hospital, Hong Kong (7), from January 2007 through August 2008, influenza infection was diagnosed for >460 hospitalized adult patients for whom acute febrile respiratory illnesses had been diagnosed. Nasopharyngeal aspiration and immunofluorescence assays (IFA) were used for rapid diagnosis of influenza A and B infection, confirmed by virus isolation. Thirteen (2.8%) patients had signs of confusion or altered consciousness, together with fever and respiratory symptoms (mean ± SD age 77.7 ± 8.8 years). We studied 3 patients from whom CSF was obtained for analysis, and who fulfilled the definition of influenza-associated acute encephalopathy (altered mental status >24 hours within 5 days of influenza onset and without alternative explanation) (1,2,4–6).

Nasopharyngeal aspirates were subjected to IFA, virus isolation, and subsequent subtyping (7). CSF specimens were subjected to virus isolation using MDCK cells, and reverse transcription–PCR to detect influenza virus RNA by using H1/H3 subtype-specific primers. Herpes simplex virus, herpes zoster virus, and enterovirus DNA/RNA was detected using PCRs (online Technical Appendix, available from www.cdc.gov/EID/content/16/1/139-Techapp.pdf).

CSF and plasma samples collected on the same day were analyzed simultaneously for the concentrations of 11 cytokines/chemokines by bead-based multiplex flow cytometry. Their assay methods and plasma reference ranges (established from >100 healthy persons) have been described (online Technical Appendix) (7). In CSF, in patients without central nervous system (CNS) disease/infection, cytokines/chemokines are either undetectable (e.g., interleukin-6 [IL-6], CXCL8/IL-8, CXCL10/IP-10, CXCL9/MIG) or present at low levels (e.g., CCL2/MCP-1) (8–10).

Concentrations of oseltamivir phosphate (OP) and its biologically active metabolite oseltamivir carboxylate (OC) were measured in CSF and plasma taken simultaneously from 1 patient who received concurrent treatment, using tandem mass spectrometry (11). The assay methods have been described (online Technical Appendix).

The clinical and virologic findings are summarized in Table 1. All case-patients were elderly (72–86 years of age), but none were known to have neuropsychiatric illness, dementia, or to be taking psychotropic medication. None had received updated influenza vaccination (6). Confusion and altered consciousness developed in patients 1 and 2 one to 2 days after the onset of fever and cough. These patients had no meningismus, focal neurologic deficit, hypotension, respiratory distress, or metabolic disturbances. Brain computed tomography (CT) scans showed no acute cerebral lesion. CSF analyses showed no bacterial or viral pathogen or pleocytosis. Oseltamivir was given to patient 2 only when influenza A was later confirmed.
by nasopharyngeal aspirate/IFA; patient 1 did not receive antiviral treatment. Both patients recovered in the next 2 days. Patient 3 had fever, severe chronic obstructive pulmonary disease exacerbation requiring noninvasive ventilatory support, complicated by acute coronary syndrome. He was given oseltamivir, 75 mg 2×/day, after influenza A infection was confirmed. Agitation and confusion developed in the patient on day 3–4 of illness (onset after the third dose of oseltamivir), despite resolution of the patient’s respiratory failure. These symptoms were followed by involuntary, tremulous movements involving all 4 limbs, while at rest and during movement. Brain CT scan was normal. Electroencephalogram showed generalized slowing of background consistent with moderate encephalopathic change (similar to that observed in septic encephalopathy) (1,6). Findings are consistent with previous reports on adult cases of influenza-associated encephalopathy: patients are all unvaccinated, pleocytosis and cerebral imaging abnormalities (even with magnetic resonance imaging) are usually absent, and symptoms are generally self-limiting (1,6). Most reports have mentioned influenza A as a cause of encephalopathy, and more commonly subtype H3N2 (1–6).

### Table 1. Clinical and laboratory findings in 3 patients with acute encephalopathy associated with influenza infection, Prince of Wales Hospital, Hong Kong*

<table>
<thead>
<tr>
<th>Clinical and laboratory findings</th>
<th>Patient 1</th>
<th>Patient 2</th>
<th>Patient 3</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, y/sex</td>
<td>76/M</td>
<td>86/F</td>
<td>72/M</td>
</tr>
<tr>
<td>Concurrent illnesses</td>
<td>Ischemic heart disease</td>
<td>Diabetes mellitus, hypertension</td>
<td>COPD</td>
</tr>
<tr>
<td>Influenza vaccination within 6 mo</td>
<td>None</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>Symptoms on examination</td>
<td>Fever &gt;38°C, cough, disorientation, incoherent speech, mental dullness</td>
<td>Fever &gt;38°C, cough, delirious, impaired consciousness, did not follow verbal command</td>
<td>Fever &gt;38°C, cough, disorientation, agitation, incoherent speech, involuntary 4-limb tremor</td>
</tr>
<tr>
<td>Focal neurologic sign or meningism</td>
<td>Absent</td>
<td>Absent</td>
<td>Absent</td>
</tr>
<tr>
<td>Chest radiograph, consolidation</td>
<td>Absent</td>
<td>Absent</td>
<td>Absent</td>
</tr>
<tr>
<td>Antiviral (oseltamivir)</td>
<td>None</td>
<td>Given</td>
<td>Given</td>
</tr>
<tr>
<td>Outcome (duration of encephalopathy)</td>
<td>Recovered (2–3 d)</td>
<td>Recovered (3–4 d)</td>
<td>Recovered (6–7 d)</td>
</tr>
<tr>
<td>Brain CT scan (noncontrast)</td>
<td>Normal</td>
<td>Old ischemic changes; known small, calcified meningioma</td>
<td>Normal</td>
</tr>
<tr>
<td>Virus isolated from NPA</td>
<td>Seasonal (H1N1) 2008</td>
<td>Subtype H3N2</td>
<td>Subtype H3N2</td>
</tr>
<tr>
<td>CSF testing results</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Opening pressure, cm H2O</td>
<td>11</td>
<td>9</td>
<td>14</td>
</tr>
<tr>
<td>Cell count (x 10^5/L)</td>
<td>1</td>
<td>–</td>
<td>0</td>
</tr>
<tr>
<td>Glucose, mmol/L</td>
<td>4.2</td>
<td>7.4</td>
<td>3.7</td>
</tr>
<tr>
<td>Protein, g/L</td>
<td>0.46</td>
<td>0.47</td>
<td>0.16</td>
</tr>
<tr>
<td>Virus isolated</td>
<td>None</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td>RT-PCR for H3 and H1 influenza virus</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Bacterial culture</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Others</td>
<td>HSV, HZV, and enterovirus PCR negative</td>
<td>HSV, HZV, and enterovirus PCR negative</td>
<td>HSV PCR negative</td>
</tr>
</tbody>
</table>

*COPD, chronic obstructive pulmonary disease; CT, computed tomographic scan; NPA, nasopharyngeal aspirate; CSF, cerebrospinal fluid; RT-PCR, reverse transcription–PCR; HSV, herpes simplex virus; HZV, herpes zoster virus. In all cases, there was no hypoglycemia, and liver and renal function test results were normal. C-reactive protein level was elevated in all cases. For patient 3, an electroencephalogram was performed and showed generalized slowing of background consistent with moderate encephalopathic change (similar to that observed in septic encephalopathy) (1,6). Findings are consistent with previous reports on adult cases of influenza-associated encephalopathy: patients are all unvaccinated, pleocytosis and cerebral imaging abnormalities (even with magnetic resonance imaging) are usually absent, and symptoms are generally self-limiting (1,6). Most reports have mentioned influenza A as a cause of encephalopathy, and more commonly subtype H3N2 (1–6).
We measured oseltamivir concentrations because of the concerns over its neuropsychiatric side-effects in children and adolescents. However, only the active metabolite (OC) was detected in the CSF of patient 3; the CSF/plasma concentration ratio was 12%–13% (18.3/143.8 ng/mL) at 18-hours postdose. This degree of CSF penetration is similar to that observed among healthy patients, with a Cmax CSF/plasma concentration ratio of 3.5% (at ≈8 hours), and a ratio of ≈10% at 18 hours (concentration-time profiles for plasma/CSF differ). Assuming a similar ratio, the CSF OP concentration would have fallen below the assay’s detection limit (0.25 ng/mL) by 18 hours (11,15). The low CSF drug-penetration, together with high cytokines in CSF and symptom progression despite drug withdrawal suggest that the manifestations of patient 3 may have been disease-related. Symptoms developed in patients 1 and 2 without antiviral exposure. Further investigations on the CNS effects of oseltamivir in the clinical setting are needed.

Our study is limited by the small patient number and the lack of feasibility in obtaining CSF for study/comparison in influenza patients without neurologic symptoms. Further studies on the clinical spectrum of influenza encephalopathy and encephalitids in adults (1,6) and their pathogenesis are indicated. In conclusion, acute encephalopathy may occur in adults with influenza. Exuberant
defense against influenza virus is rarely detected in the CSF, and pleocytosis is often absent (1,2,4–6). High levels of cytokines (e.g., IL-6, soluble tumor necrosis factor receptor 1) can be consistently found in CSF/blood specimens, correlating with disease severity and outcomes (hyperactivated cytokine response is absent in febrile seizure associated with influenza) (2–4,8). We found a broader range of cytokines/chemokines being activated (7); for certain cytokines (CXCL8/IL-8, CCL2/MCP-1), the CSF concentrations were 3× those in plasma. IL-6, CXCL8/IL-8, CCL2/MCP-1 and CXCL10/IP-10 have been shown to play pathogenic roles in CNS viral infections, cerebral injury, and acute brain syndrome in susceptible patients (7,9,10,12). The high CSF/plasma ratios suggest that for some cytokines, activation within the CNS might have occurred along with respiratory-tract and systemic productions (cytokines are not detected in CSF normally; (Table 2) (4,7–10,12). Resident macrophages/monocytes, astrocytes, microglial and endothelial cells in the CNS are shown to release cytokines/chemokines when stimulated by viral/ influenza infection; activation mechanisms without involving overt CNS invasion have been suggested (1,4,9,12–14). Cytokines may cause direct neurotoxic effects, cerebral metabolism changes, or breakdown of the blood-brain-barrier (endothelial injury) to produce symptoms (1–4,8,12–14). Whether early viral suppression by antivirals can lead to attenuation of these cytokine responses and better outcomes warrants further study (7).

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cytokine/chemokine response may play an important role in its pathogenesis.

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References


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Technical Appendix

Methods on Virus Detection and the Measurements of Cytokine and Oseltamivir Concentrations

Section I. Methods of Virus Detection

Influenza viral RNA was detected using H1/H3 subtype-specific primers (1,2) (H1: 5′-TGA GGG AGC AAT TGA GTT CA-3′ and 5′-TGC CTC AAA TAT TAT TGT GT-3′; H3: 5′-GCA ACT GTT ACC CTT ATG AT-3′ and 5′-TCA TTG TTT GGC ATA GTC AC-3′). After RNA extraction, a 10-µL aliquot of the extracted preparation was mixed with 15 µL of reverse transcription–PCR (RT-PCR) master mix, which containing 12.5 µL of 2× reaction mix, 0.5 µL of SuperScript III RT/ Platinum Taq Mix (Invitrogen, Carlsbad, CA, USA), 5 pmole of each forward and reverse primer, and adjusted to a volume of 15 µL with nuclease-free distilled water. For influenza A H1, the reaction condition was 30 min at 45°C for reverse transcription followed by 2 min at 94°C, and 30 cycles of 1 min at 94°C, 1 min at 52°C and 1 min at 68°C; and a final extension of 10 min at 68°C. The reaction condition for influenza A H3 was similar, except that reverse transcription was carried out at 50°C and the annealing temperature was 55°C. The PCR products were separated by gel electrophoresis and visualized by SYBR safe DNA stain (Invitrogen) under gel documentation system. The product size of influenza A H1 was 431 bp; the influenza A H3 product was 232 bp.
Enterovirus RNA was detected by one-step real-time RT-PCR (3,4). Amplification is specific for the 5’ untranslated region. Enterovirus RNA extracted from cell-culture fluid of clinical isolate confirmed by immunofluorescent was used as the source of positive control. RNA from the plasmid pAW109 was used as internal control for nucleic acid extraction as well as for the presence of inhibitors in the clinical samples. Primers and probes used were as follows: primer for enterovirus (EV), 5’-ACATGGGTGTGAAGAGTCTATTGAGCT-3’, 5’-CCAAAGTAGTCTCGGTTCCGC-3’; primer for pAW, 5’-GCC TGG GTT CCC TGT TCC-3’, 5’-CGA CGT ACC CCT GAC ATG G-3’; probe for EV, 5’-(FAM)TCCGGCCCCTGAATGCGGCTAAT-3’ (TAMARA); and probe for pAW, 5’-(TET)CAGGCCAATGTCTCACCACC-3’ (TAMARA).

Herpes Simplex Virus I (HSV1) was detected by real-time PCR (5). Amplification is specific for glycoprotein D. DNA extracted from cell-culture fluid confirmed positive for HSV-1 by immunofluorescent staining was used as the source of positive control. TaqMan Exogenous Internal Positive Control (EXO-IPC) DNA (Applied Biosystems, Foster City, CA, USA) was used as internal control for nucleic acid extraction as well as for the presence of inhibitors in the clinical samples. Primers used were 5’-CGGCCGTGTGACACTATCG-3’, 5’-CTCGTAAAATGGCCCCTCC-3’; TaqMan Probe: 5’FAM-CCATACCGACCACACCGAACC-TAMRA-3’.

Herpes simplex virus 2 (HSV2) was detected by real-time PCR (5). Amplification is specific for glycoprotein G. DNA extracted from cell-culture fluid confirmed positive for HSV2 by immunofluorescent staining was used as the source of positive control. TaqMan Exogenous Internal Positive Control (EXO-IPC) DNA (Applied Biosystems) was used as internal control for nucleic acid extraction as well as for the presence of inhibitors in the clinical samples. Primers
used were 5′-CGCTCTCGTAAATGCTTCCCT-3′ and 5′-TCTACCCACAACAGACCACG-3′ (4)

Varicella zoster virus (VZV) was detected by real-time PCR (5,6). Amplification is specific for the diploid gene open reading frame 62 encoding for the major immediate transactivator. DNA extracted from cell-culture fluid of the VZV vaccine strain Varilrix (GlaxoSmithKline, Rixelsart, Belgium) or from clinical specimens confirmed positive for VZV by immunofluorescent staining was used as the source of positive control. TaqMan Exogenous Internal Positive Control (EXO-IPC) DNA (Applied Biosystems) was used as internal control for nucleic acid extraction as well as for the presence of inhibitors in the clinical samples. Primers used were 5′-TCTTGTCGAGGAGGCTTCTG-3′, 5′-TGTGTGTCCACCCGGATGAT-3′; TaqMan Probe: 5′-TET-TCTCGACTGGCTGGGACTTGCG-TAMARA-3′.

Section 2. Measurement of Cytokine Concentration

Plasma concentrations of 10 cytokines and chemokines: interleukin(IL)-1β, IL-6, IL-10, IL-12p70, tumor-necrosis-factor (TNF)-α, CXCL8/IL-8, monokine induced by interferon-γ (IFN-γ) (CXCL9/MIG), IFN-γ-inducible protein-10 (CXCL10/IP-10), monocyte chemoattractant protein-1 (CCL2/MCP-1), and regulated upon activation normal T cell-expressed and secreted (CCL5/RANTES), were assayed using cytometric bead array (CBA) reagents (BD Pharmingen, San Diego, CA, USA) with 4-color FACSCalibur flow cytometer (BD Biosciences Corp, San Jose, CA, USA) (2). In CBA, different bead populations with distinct fluorescence intensities had been coated with capturing antibodies specific for different cytokines or chemokines. After incubation with 50 µL of plasma/cerebrospinal fluid (CSF), the cytokine/chemokine captured beads were mixed with phycoerythrin-conjugated detection antibodies to form sandwich complexes. Fluorescence flow cytometry of the beads provided simultaneous quantification of a panel of
cytokines and chemokines. IFN-γ was quantified by an ELISA (R & D Systems Inc., Minneapolis, MN, USA).

Section 3. Measurement of Oseltamivir Concentration

Oseltamivir concentration assays were performed by the Clinical Pharmacology Laboratory in the Faculty of Tropical Medicine, Bangkok, by using tandem mass spectrometry (7). Plasma and CSF samples were analyzed by using mixed mode (MPC-SD, 3M Empore Bracknell, UK) solid phase extraction and HILIC-LC-MS/MS. Stable isotope labeled (i.e., deuterated oseltamivir phosphate [OP] and oseltamivir carboxylate [OC]) internal standards were added to the samples before solid phase extraction. The drugs were quantified using an API 5000 triple quadrupole mass spectrometer (Applied Biosystems/MDS SCIEX, Foster City, CA, USA), with a TurboVTM ionisation source interface operated in the positive ion mode. Quantification was performed using selected reaction monitoring for the transitions m/z 313 – 225 and 316 – 228 for OP and deuterated OP, respectively, and 285 – 197 and 288 – 200 for OC and deuterated OC, respectively. The performance of the assay was demonstrated by analysis of 3 replicates of quality control samples at 3 levels. The coefficients of variation (CV%) during the analysis of OP were 4.1%, 1.9% and 2.1% at 3 ng/mL, 20 ng/mL, and 150 ng/mL, respectively (limit of detection of OP, 0.25 ng/mL). The coefficients of variation (CV%) during the analysis of OC were 2.3%, 0.5%, and 3.1% at 30 ng/mL, 400 ng/mL, and 4,000 ng/mL, respectively.

References


