Artesunate Misuse and *Plasmodium falciparum* Malaria in Traveler Returning from Africa

Dea Shahinas, Rachel Lau, Krishna Khairnar, David Hancock, and Dylan R. Pillai

*Plasmodium falciparum* malaria developed in an African-born traveler who returned to Canada after visiting Nigeria. While there, she took artesunate prophylactically. Isolates had an elevated 50% inhibitory concentration to artesinin, artemunane, and artemether, compared with that of other African isolates. Inappropriate use of artemisinin derivatives can reduce *P. falciparum* susceptibility.

Artemisinin derivatives were recently approved by the Food and Drug Administration for the treatment of *Plasmodium falciparum* malaria in North America and are available through the US Centers for Disease Control and Prevention and through Health Canada (1–3). Artemisinin-based combination therapy (ACT) remains the most effective therapy for *P. falciparum* malaria throughout the world, with the possible exception of the Thailand–Cambodia border (4). Because of the large numbers in the Toronto area of returning travelers and recent immigrants who have returned to countries of origin and visited friends and relatives, the Public Health Laboratory (Toronto) identifies ≥200 positive malaria smears annually; most *P. falciparum* isolates have come from sub-Saharan Africa. Evidence has indicated that such travelers tend not to seek medical advice before travel and are therefore at high risk of acquiring malaria (5).

The Patient

A 38-year-old Nigerian-born woman, who lived in the Toronto area (and has a good ability to recount her experiences), returned to Lagos, Nigeria, for a visit in January 2009. She did not seek pretravel advice. On arrival in Lagos, the woman purchased artemunate locally and began taking two 50-mg tablets weekly for the 4 weeks of her visit. Immediately on her return to Toronto, the patient experienced myalgia, nausea with vomiting, and chills, ≈7 days after she had taken her last dose of oral artemunate. She sought treatment at the emergency department of a community hospital. Physical examination showed that her temperature was 39.1°C and that she was dehydrated. Laboratory tests showed the following: leukocyte count 3,700 cells/μL, thrombocyte count 72 × 10^3 cells/μL, hemoglobin level 12.7 g/dL. Her chest radiograph showed that her lungs were clear. An examination of peripheral blood by thick and thin blood films showed a 0.7% parasitemia with *P. falciparum*. Her condition was treated with 1,250 mg of oral mefloquine as a single dose. She was treated as an outpatient, and she reported that symptoms promptly resolved over the next 48 hours without side effects.

A blood specimen was placed into culture in the Public Health Laboratory (Toronto), and the *P. falciparum* isolate was tested for drug susceptibility (6). The 50% inhibitory concentration (IC50) was the following for certain antimicrobial agents (tested in triplicate): chloroquine 170.5 ± 7.8 nmol/L, mefloquine 16.6 ± 0.7 nmol/L, artemisinin 20.1 ± 0.6 nmol/L, artesunate 6.2 ± 1.4 nmol/L, dihydroartemisinin 1.8 ± 0.9 nmol/L, and artemether 21.4 ± 5.3 nmol/L. For this *P. falciparum* isolate, IC50 was significantly higher for artemisinin, artemunate, and artemether than for other representative *P. falciparum* isolates imported from Africa (Figure). Because of the short half-life of artemunate, the weekly doses of the oral drug may have led to development of a resistant strain when the patient was in Nigeria. Artesunate-containing drugs therefore should not be used for prophylaxis or single drug therapy. The purchased artemunate may also have been counterfeit and may have contained lower levels of active drug. Although these data suggest that this isolate has reduced susceptibility to artemisinin derivatives, the correlation between in vitro susceptibility and treatment outcomes does not appear to be consistent (4).

Previous studies have reported that resistance to artemisinin is mediated by an increase in gene copy number, mutations within the efflux pump of the *P. falciparum* multidrug resistance 1 (*pfmdr1*) gene, or mutations in the calcium transporter *pfATPase6* (7,8). When we examined each gene, using a combination of real-time PCR and DNA sequencing, we found that *pfmdr1* copy number was elevated in this isolate relative to that of the susceptible control strain 3D7. We also observed nonsynonymous mutations in both *pfmdr1* (Y184F) and *pfATPase6* (A623E, S769N), whereas other implicated residues remained in the wild-type form (9) (Table). Similar molecular analysis of other representative imported African clinical isolates demonstrated variable mutations for *pfmdr1* and *pfATPase6* and copy number in relation to IC50 values for key drugs (Table). A trend,
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Albeit weak, was observed in which increased *pfmdr1* copy number was correlated with an elevated IC$_{50}$ to mefloquine ($r = 0.52$) and artemisinin ($r = 0.42$). The presence of an asparagine (N) at position 86 of Pfmdr1, when coupled to an elevated *pfmdr1* copy number, appeared to correlate well with reduced susceptibility to artemisinin (Table). Chavchich et al. recently demonstrated that increased *pfmdr1* copy number occurred in a laboratory strain placed under drug selection pressure with artemisinin derivatives (11). However, Imwong et al. have indicated that genetic polymorphisms and copy number in *pfmdr1* do not predict treatment outcome with ACT (10).

Findings in the published literature vary in terms of use of artemisinin derivatives for in vitro drug susceptibility testing. Jambou et al. reported treatment failures with ACT in Cambodia, French Guiana, and Senegal (8). These authors used artesether for testing and showed IC$_{50}$ values of $\approx 30$ nmol/L in their “resistant” isolates from Senegal. Noedl et al. described treatment failures with ACT in Cambodia, for which IC$_{50}$ values to dihydroartemisinin were $\approx 10$ nmol/L (12). Dondorp et al. showed IC$_{50}$ values of 4–6 nmol/L to dihydroartemisinin and 6–8 nmol/L to artesunate in a region of Cambodia and Thailand where ACT treatment failures have occurred (4). Systematic molecular surveillance and standardized drug-testing methods with clinical isolates are required to establish the molecular correlates of reduced susceptibility to antimalarial drugs. In this regard, efforts are ongoing under the auspices of the Worldwide Antimalarial Research Network (13).

**Conclusions**

The patient’s infection responded to mefloquine when she was back in Canada, possibly because of the high oral dose of mefloquine. Current guidelines from the US Centers for Disease Control and Prevention recommend quinine sulfate plus doxycycline, tetracycline, or clindamycin; or atovaquone-proguanil (Malarone; GlaxoSmithKline, Mississauga, Ontario, Canada) as first- and second-line treatment for uncomplicated *P. falciparum* malaria. Reduced susceptibility to artesunate is more likely to occur when it

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**Table. Results of sequencing single-nucleotide polymorphisms of *Plasmodium falciparum* isolate**

<table>
<thead>
<tr>
<th>Strain</th>
<th>Pfmdr1</th>
<th>PIATPase</th>
<th>pfmdr1 copy no.</th>
<th>CQ IC$_{50}$, mmol/L</th>
<th>MQ IC$_{50}$, mmol/L</th>
<th>ART IC$_{50}$, mmol/L</th>
</tr>
</thead>
<tbody>
<tr>
<td>3D7</td>
<td>N</td>
<td>Y</td>
<td>S</td>
<td>N</td>
<td>D</td>
<td>A</td>
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<tr>
<td>W2</td>
<td>Y</td>
<td>Y</td>
<td>S</td>
<td>N</td>
<td>D</td>
<td>A</td>
</tr>
<tr>
<td>Cameroon</td>
<td>Y</td>
<td>F</td>
<td>S</td>
<td>N</td>
<td>D</td>
<td>E</td>
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<tr>
<td>Congo</td>
<td>Y</td>
<td>F</td>
<td>S</td>
<td>N</td>
<td>D</td>
<td>E</td>
</tr>
<tr>
<td>Kenya</td>
<td>Y</td>
<td>F</td>
<td>S</td>
<td>N</td>
<td>D</td>
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<tr>
<td>Liberia</td>
<td>N</td>
<td>F</td>
<td>S</td>
<td>N</td>
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<tr>
<td>Nigeria C</td>
<td>N</td>
<td>F</td>
<td>S</td>
<td>N</td>
<td>D</td>
<td>E</td>
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<tr>
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<td>F</td>
<td>S</td>
<td>N</td>
<td>D</td>
<td>E</td>
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<tr>
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<td>N</td>
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<td>S</td>
<td>N</td>
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<tr>
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</tr>
<tr>
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<td>F</td>
<td>S</td>
<td>N</td>
<td>D</td>
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</tr>
</tbody>
</table>

*At Pfmdr1 and PIATPase residues previously implicated in artemisinin resistance and gene copy number of pfmdr1 by quantitative real-time PCR in relation to mean IC$_{50}$ (n = 3) data for key drugs (7,8,10). pfmdr1, *P. falciparum* multidrug resistance 1; CQ, chloroquine; MQ, mefloquine; ART, artemisinin; IC$_{50}$, 50% minimum inhibitory concentration; N, asparagine; Y, tyrosine; S, serine; D, aspartic acid; A, alanine; E, glutamic acid; 3D7, chloroquine-sensitive laboratory strain; W2, chloroquine-resistant laboratory strain; Nigeria A, clinical isolate described in this report.†*
is associated with inappropriate use of artemisinin derivatives than because of circulating artemisinin-resistant \textit{P. falciparum} in sub-Saharan Africa.

In an effort to achieve consensus that artesunate oral monotherapies should not be marketed, the World Health Organization convened the international pharmaceutical sector in April 2006. At that time, 15 companies agreed to cease manufacturing artesunate monotherapies. However, oral artesunate monotherapies may still be purchased over the counter in malaria-endemic countries, as this report shows. Thus, strains of \textit{P. falciparum} malaria are currently at risk of developing reduced susceptibility to artesunate derivatives.

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Dr Pillai is a medical microbiologist at the Public Health Laboratory (Toronto), clinical associate at the University Health Network, and assistant professor of medicine at the University of Toronto. His research interests focus on reversing mechanisms of antimicrobial resistance and laboratory surveillance of infectious diseases, including malaria and \textit{Streptococcus pneumoniae} and \textit{Clostridium difficile} infections.

**References**


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