Co-detection of Pandemic (H1N1) 2009 Virus and Other Respiratory Pathogens

Kassi Koon, Catherine M. Sanders, Jessica Green, Leslie Malone, Holly White, Delineliz Zayas, Rebecca Miller, Stanley Lu, and Jian Han

From May through October 2009, a total of 10,624 clinical samples from 23 US states were screened for multiple respiratory pathogen gene targets. Of 3,110 (29.3%) samples positive for pandemic (H1N1) 2009 virus, 28% contained ≥1 other pathogen, most commonly *Staphylococcus aureus* (14.7%), *Streptococcus pneumoniae* (10.2%), and *Haemophilus influenzae* (3.5%).

For previous and current influenza A pandemics, postmortem studies have established a strong link between secondary bacterial infections and increased deaths (1,2). Numerous respiratory pathogens can be detected from a single sample by using a multiplex molecular method called target-enriched multiplex PCR (3–6). During the 2006 influenza season, this method was used at Vancouver Children and Women’s Hospital to study 1,742 patients with acute respiratory infections; >2 pathogens were detected for ≈27% of patients studied (7). We used this method to learn more about infections occurring concurrently with pandemic (H1N1) 2009.

The Study

From May through October 2009, a total of 10,624 clinical samples from 23 states throughout the United States were submitted to Diatherix Laboratories (www.diatherix.com; Huntsville, AL, USA) and screened for multiple respiratory pathogen gene targets. Diatherix, a reference laboratory certified by the Clinical Laboratory Improvement Amendments, provides molecular differential detection services based on target-enriched multiplex PCR technology. The respiratory infections panel detects bacterial and viral pathogens associated with respiratory infections and includes targets for the following: adenovirus (types 3, 4, 7, 21), coxsackievirus, echovirus, human metapneumovirus (types A and B), influenza virus (types A and B), parainfluenza virus (types 1–4), respiratory syncytial virus (types A and B), rhinovirus, *Acinetobacter baumannii*, *Chlamydophila pneumoniae*, *Haemophilus influenzae*, *Klebsiella pneumoniae*, *Legionella pneumophila*, *Mycoplasma pneumoniae*, *Neisseria meningitidis*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Streptococcus pneumoniae*, and *Streptococcus pyogenes* (group A). Additionally, targets specific for pandemic (H1N1) 2009 virus were developed, validated, and approved by the US Food and Drug Administration under Emergency Use Authorization provisions for patient testing.

Of the respiratory specimens shipped by overnight mail from the 23 states, >95% were nasopharyngeal swabs in transfer buffer. High-throughput nucleic acid extraction was performed automatically by using KingFisher 96 instrumentation (Thermo Scientific, Hudson, NH, USA) and MagnetX chemistry (Scigenix, Marietta, GA, USA) according to manufacturers’ specifications. Multiplex PCR amplification and Luminex (Austin, TX, USA) liquid suspension detection methods were based on internally validated protocols. Reactions were amplified by using ABI 9700 thermocyclers (Applied BioSystems, Singapore), and the resulting PCR products were detected by using the LiquiChip 200 Workstation (Luminex) according to previously described protocols (3,6).

Of the 10,624 samples studied, 4,690 (44.1%) were negative for all pathogens detectable with the assay. Among the 7,514 (70.73%) samples negative for pandemic (H1N1) 2009 virus, 3 bacterial pathogens predominated: *S. aureus* (875; 11.65%), *S. pneumoniae* (573; 7.63%), and *H. influenzae* (411; 5.47%) (Table 1). The most common viral pathogens in the pandemic (H1N1) 2009-negative samples were from the family *Picornaviridae*: coxsackie/echovirus (650; 8.65%), and rhinovirus (449; 5.98%) (Table 2).

Of the 10,624 samples studied, 3,110 (29.3%) were positive for pandemic (H1N1) 2009 virus, representing 52.4% of samples positive for any pathogen. Among pandemic (H1N1) 2009 virus–positive samples, ≥1 other pathogen was co-detected for 28% (Figure). The most commonly co-detected pathogens were *S. aureus* (458; 14.73%), *S. pneumoniae* (316; 10.16%), and *H. influenzae* (110; 3.54%) (Table 1).

A significant difference (*t* = 25.6, *p* = 0.01) was found for the age distribution between patients with positive and negative pandemic (H1N1) 2009 virus results. The mean ± SD age was 19.64 (±14.45) years for those who were pandemic (H1N1) 2009 positive and 29.67 (±19.74) years for those who were negative. The median age of the 5 patients for whom 3 other pathogens were co-detected with pandemic (H1N1) 2009 virus (Figure) was 15.5 years. *S. pneumoniae* was detected in all 5 of these samples. For most,
Emerging Infectious Diseases • www.cdc.gov/eid • Vol. 16, No. 12, December 2010

the other 2 pathogens were bacteria; for only 1, a virus (parainfluenza) was detected. Of the 96 samples in which pandemic (H1N1) 2009 virus and 2 other pathogens were co-detected (Figure), 30 (31.25%) contained pandemic (H1N1) 2009 virus and 2 other pathogens were bacteria; for only 1, a virus was 4.25 years, whereas the median age of all 96 patients was 8.2 years. The median age for the 28% of patients for whom ≥1 other target was detected was 11.8 years.

Conclusions

The main finding of this large-scale clinical study was the co-detection of multiple pathogens with the pandemic influenza virus strain. In 44% of samples, no pathogens were detected, which may represent infection with common pathogens not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay.

Table 1. Results of screening of clinical samples from 23 US states for pandemic (H1N1) 2009 virus and bacterial respiratory targets, May–October 2009*

<table>
<thead>
<tr>
<th>No. (%) samples</th>
<th>S. aureus</th>
<th>P. aeruginosa</th>
<th>S. pyogenes</th>
<th>N. meningitidis</th>
<th>H. influenzae</th>
<th>C. pneum</th>
<th>K. pneum</th>
<th>M. pneum</th>
<th>S. pneum</th>
<th>A. baumannii</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pandemic (H1N1)</td>
<td>458 (0.096)</td>
<td>3 (0.129)</td>
<td>4 (0.225)</td>
<td>110 (0.32)</td>
<td>3 (0.032)</td>
<td>1 (0.032)</td>
<td>1 (0.032)</td>
<td>316 (1.093)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2009 positive, 3,110 (29.270)</td>
<td>(14.727)</td>
<td>(0.129)</td>
<td>(0.225)</td>
<td>(0.32)</td>
<td>(0.032)</td>
<td>(0.032)</td>
<td>(1.093)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pandemic (H1N1)</td>
<td>875 (0.373)</td>
<td>28 (0.452)</td>
<td>34 (0.213)</td>
<td>16 (0.106)</td>
<td>411 (0.156)</td>
<td>8 (0.586)</td>
<td>8 (0.106)</td>
<td>573 (2.023)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2009 negative, 7,514 (70.730)</td>
<td>(11.645)</td>
<td>(0.452)</td>
<td>(0.213)</td>
<td>(0.106)</td>
<td>(2.023)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total, 10,624 (100.00)</td>
<td>1,333 (0.292)</td>
<td>31 (0.358)</td>
<td>38 (0.216)</td>
<td>23 (0.085)</td>
<td>521 (0.085)</td>
<td>9 (0.621)</td>
<td>186 (1.751)</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Screening for Legionella pneumophila detected no bacteria in any samples. S. aureus, Staphylococcus aureus; P. aeruginosa, Pseudomonas aeruginosa; S. pyogenes, Streptococcus pyogenes; N. meningitidis, Neisseria meningitidis; H. influenzae, Haemophilus influenzae; C. pneum, Chlamydophila pneumoniae; K. pneum, Klebsiella pneumoniae; M. pneum, Mycoplasma pneumoniae; S. pneum, Streptococcus pneumoniae; A. baumannii, Acinetobacter baumannii. Boldface indicates predominant pathogens.

The main finding of this large-scale clinical study was the co-detection of multiple pathogens with the pandemic influenza virus strain. In 44% of samples, no pathogens were detected, which may represent infection with common pathogens not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay. For example, bocavirus and all coronavirus groups not detected by the assay.
The true value of a multiplex molecular method of screening for infectious respiratory agents depends on the clinical relevance. Among the samples with \( \geq 1 \) positive results, 53% had positive results for viral pathogens without co-detection of bacterial pathogens. For these patients, prescription of antimicrobial drugs on the basis of clinical findings alone could serve to spread drug resistance through selective pressure on normal flora. Furthermore, limited secondary treatment resources, such as oseltamivir administration during a pandemic, could be prioritized on the basis of screening results. Of the 10,624 samples studied, 70.7% were negative for the pandemic (H1N1) 2009 virus strain.

Our findings suggest that multiplex screening for respiratory pathogens is useful for providing rapid surveillance information to inform physicians who would otherwise base decisions on clinical signs and symptoms alone. Electronic reporting of empirical laboratory respiratory pathogen detection provided by a Clinical Laboratory Improvement Amendments–approved laboratory can greatly enhance surveillance data collection. Because most states have the authority to collect data of public health relevance, the screening service provided by the Diatherix Laboratories could facilitate reporting of notifiable diseases.

Mrs Koon is pursuing a doctorate degree in public health at Walden University, Minneapolis, MN, USA. Her research interests include developing multiplex amplification assays for respiratory pathogens and infectious disease surveillance.

References


Address for correspondence: Catherine M. Sanders, HudsonAlpha Institute of Biotechnology–Han Lab, 601 Genome Way, Huntsville, AL 35806, USA; email: csanders@hudsonalpha.com