Rabbit Tularemia and Hepatic Coccidiosis in Wild Rabbit

To the Editor: Tularemia is a highly pathogenic zoonosis caused by the gram-negative intracellular bacterium Francisella tularensis. F. tularensis causes serious septicemia in animals, especially wild rodents and lagomorphs (rabbits and hares), and potentially fatal, multisystemic disease in humans. The human mortality rate can reach 30% in untreated persons (1). F. tularensis is listed as a category A bioterrorism agent by the Centers for Disease Control and Prevention alongside the causative agents of anthrax, plague, smallpox, botulism, and viral hemorrhagic fevers. Generally, lesions associated with septicemic tularemia include multifocal 1–2-mm, white foci of necrosis in the liver, spleen, lymph nodes, and lungs.

Eimeria stiedae is the causative agent of hepatic coccidiosis, a common disease of wild rabbits (2) that can result in severe hepatic injury and death in juveniles and neonates. The gross lesion associated with hepatic coccidiosis is unique and nearly pathognomonic. Because E. stiedae causes proliferation of bile duct epithelial cells, affected livers contain multifocal, well-demarcated, linear, occasionally branching, bosselated, yellow to pearl-gray lesions that reflect the course of the biliary tree.

We describe a unique case of tularemia in a rabbit co-infected with E. stiedae. This case was initially misdiagnosed as simple E. stiedae infection on the basis of the classical gross lesions of hepatic coccidiosis, which overshadowed the more subtle tularemia lesions.

A juvenile wild rabbit was brought to a local veterinary clinic for postmortem examination. The owner, located in southwestern Missouri near the Arkansas–Kentucky border, raises wild-captured rabbits in a 10-acre, fenced area reserved for the training of hunting dogs. Beginning in the summer of 2009, a gradual rabbit die-off occurred, progressing to almost complete depopulation by May 2010. The liver from the dead rabbit was submitted to the University of Missouri Veterinary Medical Diagnostic Laboratory (Columbia, MO, USA). Gross examination showed the liver contained multifocal to coalescing, linear, yellow to gray nodules consistent with the classical appearance of hepatic coccidiosis. Although no gross evidence of tularemia was observed, the specimen was treated as potentially infected with tularemia because the veterinarian requested F. tularensis testing. Samples were collected and processed for bacteriologic culture, PCR, and histologic evaluation within the confines of a certified biological hood.

The liver contained 2 distinct microscopic lesions. The first was severe biliary hyperplasia with numerous intraepithelial coccidia, consistent with hepatic coccidiosis, as was anticipated. The second, more surprising lesion was an acute, multifocal, necrotizing hepatitis (Figure). The differential diagnoses for acute, multifocal, necrotizing hepatitis in a rabbit include tularemia, Tyzzer disease, listeriosis, and salmonellosis. In this instance, F. tularensis was identified by bacterial culture (3) and PCR as previously described (4). No other pathogenic bacteria were isolated on culture. These results were reported to the veterinarian, the owner, and public health officials. All remaining biological specimens were immediately discarded following the University of Missouri’s select agent protocols, and further analysis was halted, preventing further typing of the isolated F. tularensis.

According to the Centers for Disease Control and Prevention, ≈126 cases of tularemia are reported annually in the United States (5). During 2000–2008, Missouri had the highest number of reported cases (228) followed by Arkansas (149) (5). Two subspecies of F. tularensis are endemic to the United States: the highly virulent F. tularensis subsp. tularensis (type A) and the moderately virulent F. tularensis subsp. holarctica (type B).

Figure. Liver from a juvenile wild rabbit with numerous oval Eimeria stiedae oocysts in the convoluted hyperplastic bile ducts (asterisks) and necrotizing hepatitis (arrow) by Francisella tularensis. Hematoxylin and eosin stain; scale bar = 200 μm.
Transmission of the bacterium occurs primarily through bites from arthropods, including the dog tick (Dermacentor variabilis), the wood tick (D. andersoni), the lone star tick (Amblyomma americanum), and the deer fly (Chrysops spp.). In addition, contact with infected animals, most commonly rabbits, wild rodents, and cats, is another common route of transmission to humans (1,6).

Tularemia occurs in various animal species. Lagomorphs, rodents, and sheep are most susceptible; infected animals are frequently found dead or moribund. Carnivores are less susceptible; however, feline tularemia occurs sporadically, and human infections associated with bites and scratches from infected cats have been recognized (7). In addition to arthropod bites, contact with infected dead rabbits or their tissues appears to be the most common source of human infection. A wide variety of case reports have been published describing unique incidences of rabbit–human transmission, including a lawn mower aerosolizing rabbit nests along with their occupants (8), consumption of undercooked rabbit meat (9), and contact with a “lucky” rabbit’s foot (10).

The purpose of this report is to alert veterinarians, veterinary laboratory personnel, and public health officials that rabbit tularemia can be easily overlooked on gross examination in animals displaying lesions of hepatic coccidiosis, a common disease of the wild rabbit. Therefore, all rabbits submitted for postmortem examinations should be regarded as potentially infected with tularemia, particularly during seasons when vectors are active.

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Imported Leishmaniasis in Dogs, US Military Bases, Japan

To the Editor: Leishmaniasis is found in canids in ≈50 of the 88 countries where leishmaniasis are found in humans (1). In Japan, 2 cases of imported canine leishmaniasis have been documented in dogs from Spain (2,3). We report 2 cases of leishmaniasis in dogs in which dermatitis developed mainly on the face. Leishmaniasis was diagnosed from results of a serologic rk39 test, followed by PCR of skin lesion specimens for the Leishmania spp.–specific small subunit (SSU) rRNA gene. Because the dogs had lived on a US military base in Sicily, Italy, for 3 years before their owners were transferred to Japan, the animals were likely infected with L. infantum in Italy.

Animal 1 was a 6-year-old female dog that had lived in Sicily for 3 years, since 2003, and had been brought to Japan in September 2006. While she lived in Italy, she had exhibited alopecic, pruritic, and crusty skin lesions, mainly around the face and on the forearms and hind legs.

In November 2006, the dog was brought to the US Army Veterinary Command’s Zama Veterinary Treatment Facility with dermatitis (online Appendix Figure, panel A, www.cdc.gov/EID/content/16/12/2017-appF.htm) and additional signs of kidney failure. A serum specimen was positive by the rk39 dipstick test for diagnosis of visceral leishmaniasis (Kalazar Detect; InBios, Seattle, WA, USA). A skin punch biopsy specimen was obtained for cultures and PCR for the parasites in December 2006. Cultures of 4 skin specimens were all negative, probably because of cool transportation of the samples for 1.5 days before the cultures were started. The dog’s condition was treated with ketoconazole and then allopurinol. The