

# *Plasmodium knowlesi* in Human, Indonesian Borneo

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*Plasmodium knowlesi* is now established as the fifth *Plasmodium* species to cause malaria in humans. We describe a case of *P. knowlesi* infection acquired in Indonesian Borneo that was imported into Australia. Clinicians need to consider this diagnosis in a patient who has acquired malaria in forest areas of Southeast Asia.

*Plasmodium knowlesi* is now recognized as a cause of potentially fatal human malaria in forest areas of Southeast Asia. We describe a case of *P. knowlesi* malaria acquired in Indonesia and imported to Australia.

## The Patient

A 39-year-old man from Australia came to a suburban hospital in Sydney, New South Wales, Australia, with a 2-week history of morning fevers and mild headaches. His symptoms started 13 days after he left Indonesian Borneo (Kalimantan). The patient had spent an average of 10 days per month for the past 18 months working adjacent to a forest area in South Kalimantan Province, Indonesian Borneo. The most recent visit was toward the end of the rainy season. He did not use any personal vector avoidance measures (mosquito nets, long clothing, insect repellent) or receive malaria chemoprophylaxis. The patient did not travel to any other malaria-endemic areas during this 18-month period.

He did not have a remarkable medical history. On examination, he was febrile (38.9°C) and had a heart rate of 88 beats/min, blood pressure of 128/78 mm Hg, normal respiration rate, and oxygen saturation 99% on room air. Physical examination were unremarkable. Laboratory investigations showed mild thrombocytopenia ( $106 \times 10^9$  cells/L, reference range  $150\text{--}450 \times 10^9$  cells/L), mild leukopenia ( $3.7 \times 10^9$  cells/L, reference range  $4.3\text{--}10 \times 10^9$  cells/L), and unremarkable results for levels of hemoglobin (142 g/L, reference range 130–180 g/L), bilirubin (12  $\mu\text{mol/L}$ ,

reference value  $<20 \mu\text{mol/L}$ ), and creatinine (95  $\mu\text{mol/L}$ , reference range 40–120  $\mu\text{mol/L}$ ).

Malaria parasites were seen on Giemsa-stained thick and thin blood films (parasitemia level 185 parasites/ $\mu\text{L}$ ). Parasite morphologic features resembled those of *P. malariae* with typical trophozoite band forms and heavily pigmented schizonts found inside smaller erythrocytes (Figure). Some parasites had morphologic features similar to those of *P. falciparum*. These similarities included ring forms and mature trophozoites with stippling of erythrocytes (Figure, panel B). A rapid diagnostic test result for histidine-rich protein 2 of *P. falciparum* was negative.

Given increased reports of *P. knowlesi* in Malaysian Borneo, we conducted molecular studies to identify the species. Results of multiplex PCRs for *P. falciparum*, *P. vivax*, *P. malariae*, and *P. ovale* were negative (1). Results of a PCR using *P. knowlesi*-specific primers (2) were positive for undiluted and diluted (1:50) blood samples. Sequencing of a small subunit rRNA gene product showed 100% identity with *P. knowlesi* (National Center for Biotechnology Information accession no. GU049678).

The patient was treated with atovaquone/proguanil, 250 mg/100 mg, 4 $\times$ /day for 3 days. His fever resolved and the thrombocyte count returned to the reference level within 48 hours. He did not show any complications.

## Conclusions

Naturally acquired human infection with *P. knowlesi* was first described in Malaysian Borneo in 1965 after an unusual sequence of events (3). Extensive investigation at this time failed to demonstrate zoonotic transmission of simian malaria to humans. More recently, molecular techniques have identified human infections with *P. knowlesi*, establishing it as the fifth *Plasmodium* species that infects humans (2). *P. knowlesi* accounts for most (70%) human malaria infections requiring hospitalization in Sarawak, Malaysian Borneo (4), and is widespread throughout Malaysia (5). Reports have also described human infections in Thailand (6), along the border of the People's Republic of China and Myanmar (7), Singapore (8), and the Philippines (9). A recent epidemiologic study reported that 4/22 malaria cases diagnosed by microscopy as *P. falciparum*, *P. vivax*, or mixed *P. falciparum*/*P. vivax* infections were identified retrospectively by PCR to be mixed infections that included *P. knowlesi* (10). Presumably, *P. knowlesi* may account for a higher proportion of cases if those diagnosed morphologically as *P. malariae* were investigated.

Human malaria was considered to be caused by only 4 *Plasmodium* species in the premolecular biology era. Simian and human malaria parasites, including *P. knowlesi* and *P. malariae*, are often indistinguishable morphologically. Clues to identification of *P. knowlesi* by light microscopy

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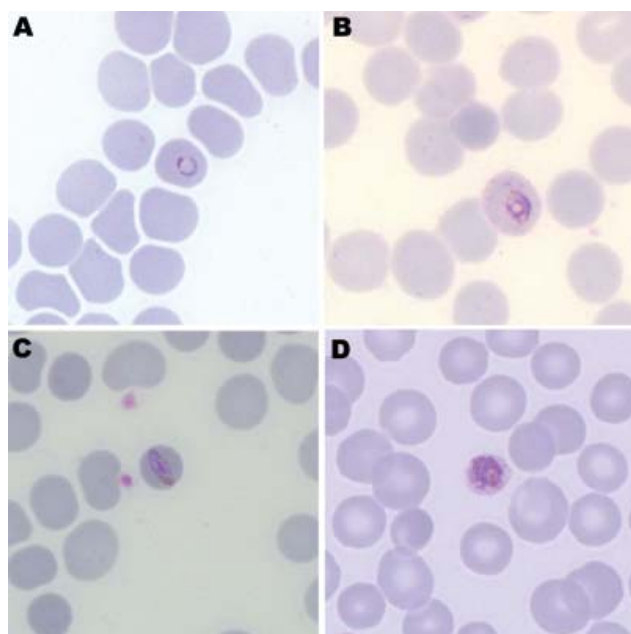


Figure. Giemsa-stained thin blood films of patient infected with *Plasmodium knowlesi*, showing a ring form (A), a trophozoite with Sinton and Mulligan stippling (B), a band form resembling *P. malariae* (C), and an early schizont (D). Original magnification  $\times 100$ .

that are useful, if present, include early trophozoites with fine ring forms, double chromatin dots, and 2–3 parasites per erythrocyte (resembling *P. falciparum*), trophozoites with a bird’s-eye appearance, mature trophozoites with a band appearance resembling *P. malariae* (Figure, panel C), and mature schizonts with a higher average merozoite count (16/erythrocyte) than in *P. malariae* (12/erythrocyte) (2,11).

Commercially available rapid diagnostic tests do not distinguish *P. knowlesi* from other forms of human malaria parasites. Lactate dehydrogenase produced by the 4 other *Plasmodium* spp. (pLDH) that cause human malaria is also present in *P. knowlesi*. Antibodies specific for pLDH of *P. falciparum* and *P. vivax* cross-react with pLDH of *P. knowlesi* (12) and therefore cannot be used to reliably distinguish *P. knowlesi* from mixed infections.

Distinction of *P. knowlesi* from *P. malariae* has useful management implications for patients and public health control measures. *P. knowlesi* potentially can cause severe disease and death, whereas *P. malariae* is generally benign. Daneshvar et al. recently published a prospective study of *P. knowlesi* infection in humans (4). They reported thrombocytopenia in 100% (107/107) of persons infected with *P. knowlesi* and lower mean  $\pm$  SD thrombocyte counts ( $71 \pm 35 \times 10^9$  cells/L) than in persons infected with *P. falciparum* ( $108 \pm 59 \times 10^9$  cells/L) or *P. vivax* ( $118 \pm 51 \times 10^9$  cells/L). Mean parasitemia level was 1,387 parasites/ $\mu$ L;

30.8% (33/107) of the case-patients had  $<500$  parasites/ $\mu$ L. Severe infection was found in 7 (6.5%) of 107 patients, and the case-fatality rate was 1.8% (2/107) among hospitalized patients (4).

Deaths and severe disease caused by *P. knowlesi* result from pulmonary and hepatorenal failure (5). Severity of *P. knowlesi* infection is related to potentially high parasitemia levels produced by its rapid and unique 24-hour erythrocytic cycle and its ability to infect all stages of erythrocytes (13). Sequestration is not thought to occur during *P. knowlesi* infection, and neurologic complications seen during *P. falciparum* infection have not been described. Although our patient was treated with atovaquone/proguanil, patients with similar uncomplicated cases have responded well to treatment with chloroquine (4).

Public health control is challenging in areas where zoonotic human malaria is endemic (14). Standard public health measures for malaria prevention (insecticide-treated nets, indoor residual spraying, and intermittent preventive treatment in the reservoir population) are likely to be less effective than for typical forms of human malaria. Nevertheless, travelers to malaria-endemic areas should be encouraged to practice mosquito bite protection measures and chemoprophylaxis.

*P. knowlesi* malaria is transmitted from long-tailed (*Macaca fascicularis*) and pig-tailed (*M. nemestrina*) macaques to humans by *Anopheles latens* mosquitoes (in the Kapit Division of Malaysian Borneo) when humans visit forest or forest fringe areas. However, transmission does not seem to occur readily in villages (2,15). Increased recognition of *P. knowlesi* indicates that human infection is possible by with other simian malaria parasites (*P. cynomolgi* and *P. inui*).

We report a patient with *P. knowlesi* infection that was acquired in Indonesia and imported to Australia. Fortunately, this patient had a low parasitemia level and mild disease. A high degree of clinical suspicion is likely to increase the number of *P. knowlesi* cases diagnosed in patients with malaria acquired in forest areas of Southeast Asia.

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Dr Figtree is microbiology registrar at Royal North Shore Hospital, Sydney. Her research interests include the molecular diversity of *P. vivax*.

#### References

1. Padley D, Moody AH, Chiodini PL, Saldanha J. Use of a rapid, single-round, multiplex PCR to detect malarial parasites and identify the species present. *Ann Trop Med Parasitol*. 2003;97:131–7. DOI: 10.1179/000349803125002977

2. Singh B, Kim Sung L, Matusop A, Radhakrishnan A, Shamsul SS, Cox-Singh J, et al. A large focus of naturally acquired *Plasmodium knowlesi* infections in human beings. *Lancet*. 2004;363:1017–24. DOI: 10.1016/S0140-6736(04)15836-4
3. Chin W, Contacos PG, Coatney GR, Kimball HR. A naturally acquired quotidian-type malaria in man transferable to monkeys. *Science*. 1965;149:865. DOI: 10.1126/science.149.3686.865
4. Daneshvar C, Davis TM, Cox-Singh J, Rafa'ee MZ, Zakaria SK, Divis PC, et al. Clinical and laboratory features of human *Plasmodium knowlesi* infection. *Clin Infect Dis*. 2009;49:852–60. DOI: 10.1086/605439
5. Cox-Singh J, David TM, Lee KS, Shamsul SS, Matusop A, Ratnam S, et al. *Plasmodium knowlesi* malaria in humans in widely distributed and potentially life threatening. *Clin Infect Dis*. 2008;46:165–71. DOI: 10.1086/524888
6. Jongwutiwes S, Putaporntip C, Takuya I, Tetsutaro S, Hiroji K. Naturally acquired *Plasmodium knowlesi* malaria in human, Thailand. *Emerg Infect Dis*. 2004;10:2211–3.
7. Zhu HM, Li J, Zheng H. Human natural infection of *Plasmodium knowlesi* [in Chinese]. *Zhongguo Ji Sheng Chong Xue Ji Sheng Chong Bing Za Zhi*. 2006;24:70–1.
8. Ng OT, Ooi EE, Lee CC, Lee PJ, Ng LC, Wong PS, et al. Naturally acquired human *Plasmodium knowlesi* infection, Singapore. *Emerg Infect Dis*. 2008;14:814–6. DOI: 10.3201/eid1405.070863
9. Luchavez J, Espino F, Curameng P, Espina R, Bell D, Chiodini P, et al. Human infections with *Plasmodium knowlesi*, the Philippines. *Emerg Infect Dis*. 2008;14:811–3. DOI: 10.3201/eid1405.071407
10. Berens-Rihas N. *Plasmodium knowlesi* found in several samples from Indonesia. *ProMed*. 2009 Jun 21 [cited 2010 Jan 12]. <http://www.promedmail.org>, archive no. 20090621.2278.
11. Lee KS, Cox-Singh J, Singh B. Morphological features and differential counts of *Plasmodium knowlesi* parasites in naturally acquired human infections. *Malar J*. 2009;8:73.
12. McCutchan TF, Piper RC, Makler MT. Use of malaria rapid diagnostic test to identify *Plasmodium knowlesi* infection. *Emerg Infect Dis*. 2008;14:1750–2.
13. Knowles RM, DasGupta BM. A study of monkey-malaria and its experimental transmission to man. *Indian Medical Gazette*. 1932;67:301–20.
14. Cox-Singh J, Singh B. Knowlesi malaria: newly emergent and of public health importance? *Trends Parasitol*. 2008;24:406–10. DOI: 10.1016/j.pt.2008.06.001
15. Vythilingam I, Tan CH, Asmad M, Chan ST, Lee KS, Singh B. Natural transmission of *Plasmodium knowlesi* to humans by *Anopheles latens* in Sarawak, Malaysia. *Trans R Soc Trop Med Hyg*. 2006;100:1087–8. DOI: 10.1016/j.trstmh.2006.02.006

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# etymologia

## *Clostridium difficile*

[klos-trid'e-əm di-fi-sil']

*Clostridium*, the genus name of these gram-positive, spore-forming, anaerobic bacteria, comes from Greek *klōstēr* (spindle) because, under the microscope, the colonies resemble spindles used in cloth weaving and long sticks with a bulge at the end. The species name *difficile* is a form of the Latin adjective *difficilis* because, when first identified (by Hall and O'Toole in 1935), the organism was difficult to isolate and grew slowly in pure culture. However, likely because of the familiarity of a French term with the same spelling and meaning, the French pronunciation has become widely used. These bacteria are part of the commensal intestinal flora in humans, and toxigenic strains of the organism can cause pseudomembranous colitis, a severe infection of the colon, after normal gut flora have been eradicated in patients who have received antimicrobial drugs.

**Source:** Kelly CP, Pothoulakis C, LaMont JT; *Clostridium difficile* colitis. *N Engl J Med*. 1994;330:257–62; Wells J. My phonetic blog. 2006. [www.phon.ucl.ac.uk/home/wells/blog0606.htm](http://www.phon.ucl.ac.uk/home/wells/blog0606.htm); [www.statemaster.com/encyclopedia/Clostridium-difficile](http://www.statemaster.com/encyclopedia/Clostridium-difficile); Dorland's illustrated medical dictionary, 31st ed. Philadelphia: Saunders Elsevier; 2007.