Plasmodium knowlesi in Human, Indonesian Borneo

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Plasmodium knowlesi is now recognized as the cause of a small subunit rRNA gene product showed 100% identity with P. knowlesi (National Center for Biotechnology Information accession no. GU049678).

The Patient

A 39-year-old man from Australia came to a suburban hospital in Sydney, New South Wales, Australia, with a 2-week history of morning fevers and mild headaches. His symptoms started 13 days after he left Indonesian Borneo (Kalimantan). The patient had spent an average of 10 days per month for the past 18 months working adjacent to a forest area in South Kalimantan Province, Indonesian Borneo. The most recent visit was toward the end of the rainy season. He did not use any personal vector avoidance measures (mosquito nets, long clothing, insect repellent) or receive malaria chemophylaxis. The patient did not travel to any other malaria-endemic areas during this 18-month period.

He did not have a remarkable medical history. On examination, he was febrile (38.9°C) and had a heart rate of 88 beats/min, blood pressure of 128/78 mm Hg, normal respiratory rate, and oxygen saturation 99% on room air. Physical examination were unremarkable. Laboratory investigations showed mild thrombocytopenia (106 × 10⁶ cells/L, reference range 150–450 × 10⁶ cells/L), mild leukopenia (3.7 × 10⁹ cells/L, reference range 4.3–10 × 10⁹ cells/L), and unremarkable results for levels of hemoglobin (142 g/L, reference range 130–180 g/L), bilirubin (12 μmol/L, reference value <20 μmol/L), and creatinine (95 μmol/L, reference range 40–120 μmol/L).

Malaria parasites were seen on Giemsa-stained thick and thin blood films (parasitemia level 185 parasites/μL). Parasite morphologic features resembled those of P. malariae with typical trophozoite band forms and heavily pigmented schizonts found inside smaller erythrocytes (Figure). Some parasites had morphologic features similar to those of P. falciparum. These similarities included ring forms and mature trophozoites with stippling of erythrocytes (Figure, panel B). A rapid diagnostic test result for histidine-rich protein 2 of P. falciparum was negative.

Given increased reports of P. knowlesi in Malaysian Borneo, we conducted molecular studies to identify the species. Results of multiplex PCRs for P. falciparum, P. vivax, P. malariae, and P. ovale were negative (1). Results of a PCR using P. knowlesi-specific primers (2) were positive for undiluted and diluted (1:50) blood samples. Sequencing of a small subunit rRNA gene product showed 100% identity with P. knowlesi (National Center for Biotechnology Information accession no. GU049678).

The patient was treated with atovaquone/proguanil, 250 mg/100 mg, 4×/day for 3 days. His fever resolved and the thrombocyte count returned to the reference level within 48 hours. He did not show any complications.

Conclusions

Naturally acquired human infection with P. knowlesi was first described in Malaysian Borneo in 1965 after an unusual sequence of events (3). Extensive investigation at this time failed to demonstrate zoonotic transmission of simian malaria to humans. More recently, molecular techniques have identified human infections with P. knowlesi, establishing it as the fifth Plasmodium species that infects humans (2). P. knowlesi accounts for most (70%) human malaria infections requiring hospitalization in Sarawak, Malaysian Borneo (4), and is widespread throughout Malaysia (5). Reports have also described human infections in Thailand (6), along the border of the People’s Republic of China and Myanmar (7), Singapore (8), and the Philippines (9). A recent epidemiologic study reported that 4/22 malaria cases diagnosed by microscopy as P. falciparum, P. vivax, or mixed P. falciparum/P. vivax infections were identified retrospectively by PCR to be mixed infections that included P. knowlesi (10). Presumably, P. knowlesi may account for a higher proportion of cases if those diagnosed morphologically as P. malariae were investigated.

Human malaria was considered to be caused by only 4 Plasmodium species in the premolecular biology era. Simian and human malaria parasites, including P. knowlesi and P. malariae, are often indistinguishable morphologically. Clues to identification of P. knowlesi by light microscopy.
that are useful, if present, include early trophozoites with fine ring forms, double chromatin dots, and 2–3 parasites per erythrocyte (resembling \( P. falciparum \)), trophozoites with a bird’s-eye appearance, mature trophozoites with a band appearance resembling \( P. malariae \) (Figure, panel C), and mature schizonts with a higher average merozoite count (16/erythrocyte) than in \( P. malariae \) (12/erythrocyte) (2,11).

Commercially available rapid diagnostic tests do not distinguish \( P. knowlesi \) from other forms of human malaria parasites. Lactate dehydrogenase produced by the 4 other \( Plasmodium \) spp. (pLDH) that cause human malaria is also present in \( P. knowlesi \). Antibodies specific for pLDH of \( P. falciparum \) and \( P. vivax \) cross-react with pLDH of \( P. knowlesi \) (12) and therefore cannot be used to reliably distinguish \( P. knowlesi \) from mixed infections.

Distinction of \( P. knowlesi \) from \( P. malariae \) has useful management implications for patients and public health control measures. \( P. knowlesi \) potentially can cause severe disease and death, whereas \( P. malariae \) is generally benign. Daneshvar et al. recently published a prospective study of \( P. knowlesi \) infection in humans (4). They reported thrombocytopenia in 100% (107/107) of persons infected with \( P. knowlesi \) and lower mean ± SD thrombocyte counts (71 ± 35 × 10^9 cells/L) than in persons infected with \( P. falciparum \) (108 ± 59 × 10^9 cells/L) or \( P. vivax \) (118 ± 51 × 10^9 cells/L). Mean parasitemia level was 1,387 parasites/μL; 30.8% (33/107) of the case-patients had <500 parasites/μL. Severe infection was found in 7 (6.5%) of 107 patients, and the case-fatality rate was 1.8% (2/107) among hospitalized patients (4).

Deaths and severe disease caused by \( P. knowlesi \) result from pulmonary and hepatorenal failure (5). Severity of \( P. knowlesi \) infection is related to potentially high parasitemia levels produced by its rapid and unique 24-hour erythrocytic cycle and its ability to infect all stages of erythrocytes (13). Sequestration is not thought to occur during \( P. knowlesi \) infection, and neurologic complications seen during \( P. falciparum \) infection have not been described. Although our patient was treated with atovaquone/proguanil, patients with similar uncomplicated cases have responded well to treatment with chloroquine (4).

Public health control is challenging in areas where zoonotic human malaria is endemic (14). Standard public health measures for malaria prevention (insecticide-treated nets, indoor residual spraying, and intermittent preventive treatment in the reservoir population) are likely to be less effective than for typical forms of human malaria. Nevertheless, travelers to malaria-endemic areas should be encouraged to practice mosquito bite protection measures and chemoprophylaxis.

\( P. knowlesi \) malaria is transmitted from long-tailed (\( M. fascicularis \)) and pig-tailed (\( M. nemestrina \)) macaques to humans by \( Anopheles latens \) mosquitoes (in the Kapit Division of Malaysian Borneo) when humans visit forest or forest fringe areas. However, transmission does not seem to occur readily in villages (2,15). Increased recognition of \( P. knowlesi \) indicates that human infection is possible by with other simian malaria parasites (\( P. cynomolgi \) and \( P. inui \)).

We report a patient with \( P. knowlesi \) infection that was acquired in Indonesia and imported to Australia. Fortunately, this patient had a low parasitemia level and mild disease. A high degree of clinical suspicion is likely to increase the number of \( P. knowlesi \) cases diagnosed in patients with malaria acquired in forest areas of Southeast Asia.

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Dr Figtree is microbiology registrar at Royal North Shore Hospital, Sydney. Her research interests include the molecular diversity of \( P. vivax \).

References


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**etymologia**

**Clostridium difficile**

[klos-trid’e-əm di-fi-sil’]

_Clostridium_, the genus name of these gram-positive, spore-forming, anaerobic bacteria, comes from Greek _klōstēr_ (spindle) because, under the microscope, the colonies resemble spindles used in cloth weaving and long sticks with a bulge at the end. The species name _difficile_ is a form of the Latin adjective _difficilis_ because, when first identified (by Hall and O’Toole in 1935), the organism was difficult to isolate and grew slowly in pure culture. However, likely because of the familiarity of a French term with the same spelling and meaning, the French pronunciation has become widely used. These bacteria are part of the commensal intestinal flora in humans, and toxigenic strains of the organism can cause pseudomembranous colitis, a severe infection of the colon, after normal gut flora have been eradicated in patients who have received antimicrobial drugs.