To define the prevalence of pilus islet 2 (PI-2)–encoded pili in *Streptococcus pneumoniae* in a geographically defined area, we examined 590 *S. pneumoniae* isolates from population-based surveillance of invasive pneumococcal disease in Atlanta, Georgia, USA, 1994–2006. In 2006, PI-2 was present in 21% of all invasive isolates, including serotypes 1 (100%), 7F (89%), 11A (21%), 19A (40%), and 19F (75%). Only serotype 19F is included in the 7-valent pneumococcal conjugate vaccine that is in use worldwide. In 1999, PI-2-containing isolates were of the same serotypes but accounted for only 3.6% of all invasive isolates. The increase of PI-2 in 2006 resulted predominantly from the emergence of serotype 19A isolates of sequence type 320 and the expansion of serotype 7F isolates. The increase in PI-2-containing isolates and the finding that isolates of all identified serotypes expressed highly conserved PI-2 pili supports their potential as a vaccine candidate.

*Streptococcus pneumoniae* is a major human pathogen, which causes pneumonia, sinusitis, otitis, bacterial meningitis, and septicemia. Introduction of the 7-valent pneumococcal conjugate vaccine (PCV7, Prevnar; Wyeth, Madison, NJ, USA) in 2000 has dramatically decreased invasive pneumococcal disease in the United States in children and in adults (1) and is now in use worldwide. PCV7 includes capsule polysaccharide serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F, serotypes that were responsible for >80% of cases of invasive pneumococcal disease in the United States before the introduction of the conjugate vaccine (1). In recent years other non-PCV7 serotypes, including 7F and 19A, have been emerging in the United States, (2–4).

Pili have recently been identified on several gram-positive bacteria (5–7). To date, 2 different pilus islets (PIs) have been described in *S. pneumoniae* that encode the structural and biosynthetic genes for 2 antigenically different types of pili, PI-1 (8) and recently described PI-2 (9). The so-called *rlrA* pathogenicity islet (10) or PI-1 is a 14-kb genetic region present in 21%–27% of clinical isolates, depending on the geographic region analyzed. It is most prevalent in, but not restricted to, the PCV7-vaccine serotypes 4, 6B, 9V, 14, and 19F (11–13).

PI-2 is a 7-kb region located between the genes that encode peptidase T (PepT) and ferrochelatase (HemH) (9). The region is composed of 5 genes, which encode 2 surface proteins, PitA and PitB, with *pitA* a pseudogene due to a stop-codon in the N terminus, a signal peptidase-like protein (SipA), and 2 sortases (SrtG1 and SrtG2); the latter is nonfunctional in most of the strains (9). This PI has been reported to be present in ≈16% of the analyzed isolates belonging to serotypes 1, 2, 7F, 19A, and 19F (9). The presence of PI-2 pili appears to be a clonal property, rather than serotype associated, an observation that has been described for PI-1 as well (11–13). For isolates of serotype 19A, PI-2–containing isolates have been shown to be associated with sequence type (ST) ST320, a clone related to the worldwide distributed multidrug-resistant serotype 19F clone Taiwan19F-14 (4,14). ST320 belongs to clonal complex (CC) CC271 (with ST271 as the predicted founder of this CC); isolates of this CC have been shown to encode both PIs and express both pili concomitantly on their surface (9).

The PI-2 pilus is composed of polymers of the major pilus protein PitB (9). In contrast to other pili in gram-
positive bacteria that contain accessory pilus proteins, that serve as adhesins (15–17) or adaptors for pilus attachment to the cell wall (18,19), PI-2 pili appear to consist solely of PitB polymers, and these polymers themselves have been shown to mediate adhesion of *S. pneumoniae* to eukaryotic cells (9).

Bagnoli et al. identified PI-2–containing isolates in a random, worldwide collection of pneumococcal isolates (9). We describe the distribution of PI-2 pili in a well-documented, comprehensive collection from a defined geographic region, i.e., in invasive pneumococcal isolates collected as part of a population-based surveillance program conducted in the metropolitan area of Atlanta, Georgia, USA.

**Materials and Methods**

**Strain Collection**

A set of 590 strains of *S. pneumoniae* from population-based surveillance of invasive pneumococcal disease isolated between 1994 and 2006 in the 8-county metropolitan Atlanta area, Georgia Health District 3 formed the basis for this study. The isolates were obtained as a part of the Centers for Disease Control and Prevention–sponsored Active Bacterial Core surveillance of the Georgia Emerging Infections Program (1,20). Information about serotype and antimicrobial drug susceptibility was determined for all isolates as described (21). Only viable strains with documented serotype and available antibiogram were included in the study. Laboratory strain *S. pneumoniae* R6 is a non-encapsulated derivative of the serotype 2 strain D39 (22).

**PCR-based Screening for PI-2**

To ascertain whether PI-2 was absent from the described insertion site (9), we designed primers pepT_F and hemH_R (Table 1) against the flanking genes pepT and hemH. To determine whether PI-2 was present, we used primers sipA_up and sipA_dn (Table 1) designed against the PI-2–specific gene sipA. We tested for PI-1 using primers Rlr_up_F and Rlr_do_R for its absence and Rlr_SrtC_F and Rlr_srtD_R for its presence, respectively (Table 1).

**Multilocus Sequence Typing and Clonal Complexes**

Multilocus sequence typing (MLST) was performed as described (23). CCs were assigned by using the eBurst program (24) to partition the MLST database (http://spneumoniae.mlst.net). The predicted founder of a CC was assigned by eBurst to the ST with the highest number of single-locus variants.

**Construction of a pitB-Deletion Mutant**

The 3′- and 5′-flanking regions of pitB were amplified by using primer pairs sipA_up F/PilA_del_3 and PilA_del_2/SrtC_do_R (Table 1). Both PCR products were used in a PCR-based overlap-extension reaction, which introduced an MfeI restriction site between the 2 fused fragments. The PCR product was cloned into pCR-Blunt II (Invitrogen, Carlsbad, CA, USA), resulting in pCR-pitB. The erm-cassette of pSK-erm (25) was amplified by using primers erm_F and erm_R (Table 1) and ligated into MfeI-digested pCR-pitB. The resulting plasmid, pCR-pitB:erm, was used to transform GA41070 according to established protocols (26), which resulted in mutant GA41070ΔpitB.

**Production of Recombinant PitB and Antiserum**

The region of *pitB* that encoded the mature protein (bp 126–bp 1128) was amplified from chromosomal DNA of serotype 1 strain GA19686 and cloned into the *Escherichia coli* expression vector pET-32a+ (Stratagene, La Jolla, CA, USA). The thioredoxin/6xHis-PitB fusion protein was purified by using the B-PER 6xHis Fusion Purification Kit (Pierce, Rockford, IL, USA). The N-terminal thioredoxin/6xhis-tag was removed by cleavage with recombinant enterokinase (Novagen, Madison, WI, USA).

<p>| Table 1. Primers used in this study of invasive <em>Streptococcus pneumoniae</em> isolates, Atlanta, Georgia, USA, 1994–2006†‡ |</p>
<table>
<thead>
<tr>
<th>Region</th>
<th>Primer</th>
<th>Sequence (5′ → 3′)†‡</th>
</tr>
</thead>
<tbody>
<tr>
<td>Erm cassette</td>
<td>erm_F</td>
<td>gcctagacGTTAGATTAATCCCTACGAGTCGAC</td>
</tr>
<tr>
<td></td>
<td>erm_R</td>
<td>gcctagacCTCCATTCCTTATAGAAAGTGTAAAAC</td>
</tr>
<tr>
<td>PI-2</td>
<td>pepT_F</td>
<td>TAAAGAGCGGTCAAGAGATTTGG</td>
</tr>
<tr>
<td></td>
<td>hemH_R</td>
<td>AATAATGGGGCTCCAAAATCAAGC</td>
</tr>
<tr>
<td></td>
<td>sipA_up_F</td>
<td>CTCTAGGAGGGATCTTCTTTATCATC</td>
</tr>
<tr>
<td></td>
<td>sipA_do_R</td>
<td>CTACAGCGGTGGTCTGATGTTCC</td>
</tr>
<tr>
<td></td>
<td>PilA_del_3</td>
<td>gcaattgccgggccgtcagtgCTTATAGGGGATGTTCCAAAAG</td>
</tr>
<tr>
<td></td>
<td>PilA_del_2</td>
<td>cttagccgccccgttcagtGCTGGGGGCGATGATG</td>
</tr>
<tr>
<td></td>
<td>Rlr_up_F</td>
<td>CTCCCAGGAATGTTCTCAAGTGG</td>
</tr>
<tr>
<td>PI-1</td>
<td>Rlr_do_R</td>
<td>GTCTTAGGAATATCAGTTTAGGTGTCG</td>
</tr>
<tr>
<td></td>
<td>Rlr_srtC_F</td>
<td>GGGGAAAGATTGCTGGGAGACCTT</td>
</tr>
<tr>
<td></td>
<td>Rlr_srtD_R</td>
<td>GCTTGCGCTCGACGGTGCC</td>
</tr>
</tbody>
</table>

†PI, pilus islet.

‡Not included are primers used for sequencing and multilocus sequence typing.

§Lowercase letters represent bases added for mutagenesis; uppercase letters represent bases complementary to *S. pneumoniae*. 

956 Emerging Infectious Diseases • www.cdc.gov/eid • Vol. 16, No. 6, June 2010
PI-2 Pili of *Streptococcus pneumoniae*

Preparation of Cell Wall Extracts from *S. pneumoniae*

Cultures were grown to mid-exponential phase (optical density$_{600}$ 0.5–0.6) in Todd-Hewitt medium containing 0.5% yeast extract. After centrifugation, the cell pellets were resuspended in 1/20 of the culture volume in extraction buffer (50 mmol/L Tris-HCl; pH 8.0) containing 200 U/mL mutanolysin (Sigma-Aldrich, St. Louis, MO, USA) and 30% raffinose. After 2 h of incubation at 30°C, cell wall extracts were collected by centrifugation at 12,000 × g for 15 min; 20 μL cell wall extract were mixed with sodium dodecyl sulfate–polyacrylamide gel electrophoresis sample buffer and boiled for 10 min immediately before separation on a 4%–12% Tris-HCl polyacrylamide gel (NuPAGE; Invitrogen). For immunostaining, proteins were transferred to a nitrocellulose membrane and detected with polyclonal anti-PitB antiserum at a dilution of 1:20,000.

Statistical Analysis

Proportions were compared by using the Fisher exact test. p values <0.005 were considered significant.

Results

PI-2 in Strains of Serotypes 1, 7F, 11A, 19A, and 19F

To determine the post PCV-7 prevalence of PI-2 in *S. pneumoniae*, we analyzed all 381 viable and documented invasive isolates collected in 2006 in metropolitan Atlanta (381/409) in a PCR-based screen. Two PCRs were performed with each strain; 1 resulted in a PCR product in strains that lacked PI-2, and a second resulted in a product only in strains that contained PI-2. Strains that produced a PCR product for PI-2 were further characterized by amplification of *pitB*, which encoded the pilus backbone protein. For all strains, 1 of the 2 PCRs resulted in a PCR product. No strain gave a PCR product for both PCRs, which could have indicated the presence of the PI in an alternative integration site. These results strongly suggest that the *pepT-hemH* intergenic region is the only integration site of PI-2 in *S. pneumoniae*. Overall, 21% of all viable isolates collected in 2006 contained PI-2. The PI-2 of *S. pneumoniae* strain GA47784 was assigned the GenBank accession no. GU256423.

PI-2 was identified only in isolates of serotypes 1, 7F, 11A, 19A, and 19F (Figure 1). Serotypes 19A and 7F were the most frequent PI-2–containing serotypes isolated in 2006, with a PI-2 prevalence of 40% and 89%, respectively. In serotype 11A, PI-2 was identified sporadically, whereas for serotype 19F, 75% of the isolates contained PI-2. Serotype 1 was represented by a single isolate in 2006. To determine the prevalence of PI-2 in serotype 1 isolates, we analyzed an additional 20 invasive isolates, collected in Atlanta from 1994 through 2005; all contained PI-2. Serotype 1 had the highest prevalence of PI-2 of all analyzed serotypes. Except for serotype 19F, the PI-2–containing serotypes are not included in PCV7.

To further characterize PI-2 of a serotype 11A strain, we sequenced the 7-kb region between the flanking genes *pepT* and *hemH* of strain GA47784. The sequence was 99.9% identical to the published PI-2 sequences, with the highest sequence identity to serotype 1 strain PN1110, with only 1-bp difference over the entire PI-2 region. The mutation was located downstream of the stop-codon in *pitA*, which results in premature termination of PitA, as has been reported (9).

Effects of PCV7 Introduction on PI-2 Distribution

To assess whether PI-2 was associated with the emergence of serotypes in the aftermath of PCV7 introduction (2000), the PI-2 distribution was determined in invasive isolates from 1999 that belonged to the PI-2–containing serotypes (Figure 2). The numbers of isolates for the different serotypes in the 2 periods reflected the expansion of isolates of the nonvaccine serotypes 7F and 19A and the decline in 19F isolates (Figure 2) that has been described in....

Figure 1. Frequency of pilus islet 2 (PI-2)–containing *Streptococcus pneumoniae* invasive isolates in metropolitan Atlanta, Georgia, USA, 2006. Black, PI-2–containing isolates; white, PI-2–lacking isolates. *Serotypes included in the 7-valent pneumococcal conjugate vaccine.*
RESEARCH

The United States since the introduction of PCV7 (4). PI-2 was present in isolates of all 5 serotypes before the introduction of PCV7; hence, PI-2 was not recently acquired. For serotype 7F, the ratio between PI-2–containing and PI-2–lacking isolates in 2006 compared to the ratio in 1999 remained essentially the same (Figure 2), which suggests that PI-2 pili may not provide a major selective advantage to invasive 7F isolates. Serotype 19A showed a significant increase in PI-2–containing isolates. To assess the presence of PI-2 in other serotypes in 1999, we analyzed isolates of all represented serotypes of PI-2: all isolates of infrequent serotypes (1, 5, 7C, 7F, 8, 9A, 9N, 10A, 13, 15B, 15C, 16, 16F, 18F, 20, 23A, 24B, 31, 33F, 34, 35B, 35F, 37) and at least 10% of the isolates of serotypes with >10 isolates (3, 4, 6A, 6B, 9V, 11A, 12F, 14, 18C, 19A, 19F, 22F, 23F). No additional PI-2–containing isolates were detected, which suggests that serotypes 1, 7F, 11A, 19A, and 19F were the only serotypes that contained substantial numbers of PI-2–containing isolates. These PI-2–containing isolates accounted for ≈3.6% of all viable isolates collected in 1999. The estimation of total viable isolates in 1999 was based on the known proportion of viable isolates to total collected isolates of the PI-2–containing serotypes of 1999 (93%).

Genetic Relationships Between PI-2–containing Strains

To analyze the clonality of the PI-2–containing isolates, we performed MLST for all PI-2–containing isolates from 1999 and 2006 (Table 2). For serotype 1, the isolates from 1999 and 2006 belonged to ST304 and ST227, respectively (Table 2). Both STs have been described as the most prevalent serotype 1 STs in the United States (27,28). PI-2–containing serotype 7F isolates of 2006 belonged to either ST191 or ST1176, and ST191 isolates were responsible for the strong expansion of this serotype in 2006. 7F isolates lacking PI-2 were tested and found to be of ST191 (data not shown), which indicated that PI-2 is present in only a subpopulation of this ST. ST1176 is unrelated to ST191, and 7F isolates of this ST were not present in 1999. All PI-2–containing isolates of serotype 11A belonged to ST62. Analysis of additional serotype 11A isolates that lacked PI-2 showed that they all belonged to ST62 (data not shown). This finding suggests that the structure of this serotype is clonally homogeneous and that PI-2 is present in a subset of this ST.

For serotype 19A, 27 of the 39 PI-2–containing 19A isolates in 2006 were ST320 (Table 2). This ST was absent in 1999 (29); thus these isolates were the major contributor to the increase of PI-2–containing isolates of serotype 19A in 2006. Additionally, PI-2 was identified in 19A isolates of ST1339 and ST2268, and both sequence types belonged to CC2090. One ST1339 isolate was present in 1999, which indicated that PI-2 was present in this ST before PCV7 was introduced. 19A isolates of 2006 that lacked PI-2 were tested and found to belong to other STs, e.g., ST199 and ST695, STs consistent with the described population structure of 19A isolates in the United States (4).

ST analysis of the remaining isolates of 19F isolates in 2006 showed that PI-2 was present in isolates that belonged to CC271 (ST236, ST651, ST3039) and also in the unrelated CC251 (ST251, ST654, ST1258) (Table 2). However, analysis of several 19F isolates that lacked PI-2 revealed isolates of ST3039 (data not shown); this finding indicated either a loss of PI-2 or a recent acquisition of the islet in a subset of isolates of this ST. Isolates of CC271 have been described to encode and express PI-1 and PI-2 concomitantly (9). Analysis for the presence of PI-1 in our

![Table 2. Clonal distribution of PI-2-containing *Streptococcus pneumoniae* isolates, Atlanta, Georgia, USA, 1994–2006*](https://www.cdc.gov/eid)
population confirmed PI-1 in isolates of CC271 (ST320, ST271, ST3039, ST236, ST651). Additionally, PCR results showed that PI-1 was in ST1339 isolates but not in isolates of the single locus variant ST2268. Isolates of all other PI-2–containing serotypes or STs did not contain PI-1.

Antimicrobial Drug Resistance and PI-2–containing Isolates

19F and 19A isolates of CC271 are closely related to the globally distributed multidrug-resistant clone Taiwan19F, and all were resistant to penicillin, erythromycin, cotrimoxazole, tetracycline, and chloramphenicol. In addition, serotype 19A isolates of CC2090 (ST1339, ST2268) were resistant to penicillin, erythromycin, and cotrimoxazole. All PI-2–containing 19A isolates in 2006 were multidrug resistant. In contrast, PI-2–containing isolates of serotypes 1, 7F, 11A, and 19F (ST654, ST251) were susceptible to all tested antimicrobial drugs. Hence, antimicrobial drug resistance was not a consistent feature of PI-2–containing isolates.

Expression of High Molecular Weight PitB on the Surface of PI-2–containing Isolates

The first report of PI-2 pili described mutations in several genes encoded by PI-2 (namely, pitA and srtG2), which resulted in their inactivation (9). To confirm that PI-2–containing isolates indeed express PitB polymers on their surface, we produced an anti-PitB antiserum to detect PitB polymers. The specificity of the PitB antiserum was tested against cell wall extracts of strain GA41070 and its isogenic pitB deletion mutant, GA41070ΔpitB (Figure 3). In the wild type, the antibody reacted with several bands that produced the high molecular weight (HMW) banding pattern characteristic of gram-positive pili (30,31) (Figure 3). As expected, the PitB monomer and the HMW banding pattern were absent in the pitB-deletion mutant (Figure 3).

To analyze whether the PI-2–containing isolates of the different STs produced PitB polymers, we used Western blots to examine their cell wall extracts (Figure 4). All PI-2–containing strains showed the same HMW banding pattern (Figure 4, lanes 2–15). In addition, all 81 PI-2–containing isolates from 2006 were analyzed by whole cell dot blots that confirmed that all isolates expressed PitB on their surface (data not shown).

High Conservation of PitB among Different STs

To determine the degree of variation in PitB, we sequenced pitB in isolates representing the different STs (Table 2). The pitB sequences were 100% identical for all strains, except the 7F isolates GA47077 (ST191) and GA47340 (ST1176), which have a single point mutation that has been observed in other serotype 7F strains (9). The high conservation of PitB was consistent with the reported PitB sequences by Bagnoli et al. (9) and is an essential feature of PI-2 as a potential vaccine candidate. Additionally, we analyzed the gene for the second surface protein encoded in PI-2, pitA, and confirmed the presence of the described stop-codon in all strains representing the different STs (Table 2).

Discussion

PI-1 pili on S. pneumoniae have been reported as promising vaccine candidates for this important human pathogen (13,32). One limitation to this approach is that PI-1 has been found in only ≈25% of strains; it is most frequently found in pneumococcal isolates of serotypes covered by PCV7, which are decreasing in countries that have introduced the vaccine. The recent identification of PI-2 in serotypes not covered by PCV7 has raised the possibility that broad coverage of strains can be achieved by a bivalent pilus-based vaccine.

The first report on PI-2 pili described the serotypes and sequence types associated with PI-2 (9). In the current study, we defined the prevalence of PI-2–expressing isolates in a defined geographic region and population and assessed whether the introduction of PCV7 had an effect on the distribution of PI-2–containing isolates. Twenty-one
lates. Therefore, our higher results are consistent with the reported strain collection, including pre- and post-PCV7 isolates, whereas the earlier report was based on a random population, indicating rate of PI-2 in our study refers to a post-PCV7-era prevalence of PI-2 in serotype 11A (35). Isolates that cause invasive disease have been identified as belonging to ST62, the ST most common among serotype 11A (35). The low prevalence of PI-2 in serotype 11A suggests that PI-2 pili are not essential for isolates of this serotype to cause invasive disease.

The genetic profile of the serotype 19A isolates in Atlanta is consistent with the described clonal distribution of this serotype in the United States in the post-PCV7 period (4). Several capsular switching events have contributed to the emergence of serotype 19A, including (but not limited to) ST320, originating from a multidrug-resistant derivative of clone Taiwan\textsuperscript{16}-14(ST236), and CC2090 (ST1339, ST2289) from the highly related clone North Carolina\textsuperscript{26}-23 (39). Unlike serotype 7F, 19A is characterized by a heterogeneous clonal structure (4), and the presence of PI-2 is a feature of some of the clones.

Most of the PI-2–containing isolates that we found do belong to the serotypes reported for PI-2 by Bagnoli et al. (9). Serotype 2 was absent from our population during the surveillance period of 1994 to 2006, and in addition we identified PI-2 in serotype 11A. Overall, we confirmed the presence of PI-2 in emerging non-PCV7 serotypes. However, other non-PCV7 serotypes that do not have PI-2 are emerging as well, e.g., 15BCF, 22F, 33F, and 38 (2,40), which indicates that other mechanisms may influence the emergence of new serotypes.

Of note, 3 of the 4 non-PCV7 serotypes with PI-2–containing isolates (serotypes 1, 7F, 19A) are included in the 13-valent pneumococcal conjugate vaccine now being introduced in Europe, Canada, and the United States. Use of this vaccine is expected to greatly reduce the frequency of all included serotypes and may thus reduce the proportion of S. pneumoniae isolates that contain PI-2. In summary, the prevalence of PI-2 increased from \textasciitilde3.6% of the invasive pneumococcal isolates in 1999 to 21% in 2006 in metropolitan Atlanta, Georgia, especially in the emerging serotypes 7F and 19A. PI-2–containing isolates of all identified sequence types expressed polymers of the highly conserved pilus protein PitB on their surface. These findings support the potential of PI-2 pili as a vaccine candidate.

**Acknowledgments**

We are grateful to Monica M. Farley, Wendy Baughman, Sarah W. Satola, and the Georgia Emerging Infections Program...
for providing the clinical isolates and the corresponding documenta-
tion. We thank Leslie McGee and Scott T. Chaney for se-
quently type data of serotype 19A isolates.

This work was supported by funds from National Institutes of
Health grant #R01AI 070829 (D.S.S.) and a Veterans Admin-
istration Merit Award from the Department of Veterans Affairs
(D.S.S.).

Dr Zähner is a senior research associate at Emory University
School of Medicine, Atlanta. Her main research interests are sur-
face structures and antimicrobial resistance in S. pneumoniae.

References

1. Whitney CG, Farley MM, Hadler J, Harrison LH, Bennett NM, Lyn-
field R, et al. Decline in invasive pneumococcal disease after the
introduction of protein-poly saccharide conjugate vaccine. N Engl J
2. Hicks LA, Harrison LH, Flannery B, Hadler JL, Schaffner W, Craig
AS, et al. Incidence of pneumococcal disease due to non-pneumo-
coccal conjugate vaccine (PCV7) serotypes in the United States dur-
Dis. 2007;196:1346–54. DOI: 10.1086/521626
3. Singleton RJ, Hennessy TW, Bulkow LR, Hammitt LL, Zulz T, Hurl-
burd DA, et al. Invasive pneumococcal disease caused by nonvaccine
serotypes among Alaska Native children with high levels of 7-valent
pneumococcal conjugate vaccine coverage. JAMA. 2007;297:1784–
92. DOI: 10.1001/jama.297.16.1784
Schaffner W, Lexau C, et al. Population snapshot of emerging Strepto-
Dis. 2008;197:1016–27. DOI: 10.1086/528996
2005;309:105. DOI: 10.1126/science.1111563
al. Group A streptococcus produce pilus-like structures containing
protective antigens and Lancefield t antigens. Proc Natl Acad Sci U
SA. 2005;102:15641–6. DOI: 10.1073/pnas.0507808102
7. Ton-That H, Schnewind O. Assembly of pili on the surface of
Corynebacterium diphtheriae. Proc Natl Acad Sci U S A. 2005;102:
15641–6. DOI: 10.1073/pnas.0507808102
al. A pneumococcal pilus contains virulence and host inflamma-
tory responses. Proc Natl Acad Sci U S A. 2006;103:2857–62. DOI:
10.1073/pnas.0511017103
9. Bagnoli F, Moschioni M, Donati C, Dimitrovská V, Ferlenghii I, Fac-
ciotti C, et al. A second pilus type in Streptococcus pneumoniae is
prevalent in emerging serotypes and mediates adhesion to host cells.
10. Hava DL, Camilli A. Large-scale identification of serotype 4
Streptococcus pneumoniae virulence factors. Mol Microbiol. 2002;
M, et al. Association of the pneumococcal pilus with certain cap-
sular serotypes but not with increased virulence. J Clin Microbiol.
12. Aguilar SI, Serrano I, Pinto FR, Melo-Cristino J, Ramirez M. The
presence of the pilus locus is a clonal property among pneumococcal
invasive isolates. BMC Microbiol. 2008;8:41. DOI: 10.1186/1471-
2180-8-41
13. Moschioni M, Donati C, Muzzi A, Masignani V, Censini S, Han-
age WP, et al. Streptococcus pneumoniae contains 3 rIIA pilus vari-
ants that are clonally related. J Infect Dis. 2008;197:888–96. DOI:
10.1086/528375
R, et al. Nomenclature of major antimicrobial-resistant clones of
Streptococcus pneumoniae defined by the pneumococcal molecular
15. Abbot EL, Smith WD, Siou GP, Chiriboga C, Smith RJ, Wilson JA,
et al. Pili mediate specific adhesion of Streptococcus pyogenes to hu-
j.1462-5822.2007.00918.x
diphtheriae employs specific minor pilins to target human pharyn-
j.1462-5822.2007.05630.x
17. Konto-Ghiorghi Y, Mairey E, Mallet A, Dumelien G, Caliet E, Trieu-
Cuot P, et al. Dual role for pilus in adherence to epithelial cells and
5:e1000422. DOI: 10.1371/journal.ppat.1000422
JL. Sortase A utilizes an ancillary protein anchor for efficient cell
2008;76:3550–60. DOI: 10.1128/IAI.01613-07
19. Mandlik A, Das A, Ton-That H. The molecular switch that activates
the cell wall anchoring step of pili assembly in gram-positive bac-
pnas.0806350105
20. Whitney CG, Farley MM, Hadler J, Harrison LH, Lexau C, Reingold
A, et al. Increasing prevalence of multidrug-resistant Streptococcus
DOI: 10.1056/NEJM200012283432603
A, et al. The emergence of Streptococcus pneumoniae resistant to
macrolide antimicrobial agents: a 6-year population-based assess-
22. Ottolenghi E, Hoteckiss RD, Release of genetic transforming agent
from pneumococcal cultures during growth and disintegration. J Exp
Med. 1962;116:491–519. DOI: 10.1084/jem.116.4.491
23. Enright MC, Spratt B. A multilocus sequence typing scheme for
Streptococcus pneumoniae: identification of clones associated with
serious invasive disease. Microbiology. 1998;144:3049–60. DOI:
10.1099/00221287-144-11-3049
24. Feil EJ, Li BC, Aanensen DM, Hanage WP, Spratt BG. eBURST:
inferring patterns of evolutionary descent among clusters of related
bacterial genotypes from multilocus sequence typing data. J Bacte-
25. Biswas I, Scott JR. Identification of rocA, a positive regula-
26. Allolio G, Martin B, Granadel C, Claverys JP. Development of com-
petence in Streptococcus pneumoniae: pheromone autoinduction and
control of quorum sensing by the oligopeptide permease. Mol Mi-
crobiol. 1998;29:75–83. DOI: 10.1046/j.1365-2958.1998.00904.x
27. Brueggenmann AB, Spratt BG. Geographic distribution and clonal
diversity of Streptococcus pneumoniae serotype 1 isolates. J Clin
Microbiol. 2003;41:4966–70. DOI: 10.1128/JCM.41.11.4966-
4970.2003
28. Gonzalez BE, Hulten KG, Kaplan SL, Mason EO Jr. Clonality of
Streptococcus pneumoniae serotype 1 isolates from pediatric pa-
10.1128/JCM.42.6.2810-2812.2004


Address for correspondence: Dorothea Zähner, Department of Medicine, Division of Infectious Diseases, Emory University School of Medicine, Atlanta, GA 30322, USA; email: dzahner@emory.edu