Influenza surge planning is premised on a high incidence of illness among elderly persons, but if the current pattern of illness continues, healthcare facilities also should prepare to treat younger persons who may constitute the bulk of cases. Additionally, studies of persons born during 1957–1968 should be conducted to quantify antibody levels to pandemic (H1N1) 2009 virus, focusing on the degree of preexisting immunity that may have existed and was boosted by prior encounters with subtype H1N1 viruses.

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Swine Influenza A Vaccines, Pandemic (H1N1) 2009 Virus, and Cross-Reactivity

To the Editor: Since its first emergence in the human population in spring 2009 (1–3) infections with pandemic (H1N1) 2009 virus have been reported in pigs, turkeys, and some carnivore species (4,5). The pandemic (H1N1) 2009 virus can be experimentally transmitted between pigs (6). The reported transmissibility of the virus raises the question as to whether authorized swine influenza vaccine strains may be cross-reactive to pandemic (H1N1) 2009 virus. Kyriakis et al. (7) investigated the cross-reactivity of 66 pig serum samples from different infection and vaccination trials and reported cross-reactions between the avian-like H1N1 viruses circulating in the European pig population (avH1N1) and the classical swine H1N1 viruses (cH1N1) with pandemic (H1N1) 2009 virus by hemagglutination inhibition assay.

To investigate this cross-reactivity in more detail, a neutralization test was applied in the study we report here. A serial dilution of serum samples was prepared (log4). All virus strains were adjusted to 100 fifty-percent tissue culture infectious doses. This working dilution of virus was mixed with serum dilutions and incubated 1 hour at 37°C. Madin-Darby bovine kidney monolayers were infected with the neutralization mixtures. After 48 hours of incubation, cells were fixed with acetone (4°C–8°C) and investigated by indirect immunofluorescent assay. Finally, the 50% neutralization titer was calculated.

Hypermune serum samples were established by using a 4-fold vaccination of pigs with antigens of H1N1 vaccine strains (A/New Jersey/8/1976, A/sw/Netherlands/25/1980, A/sw/IDT/Re230/1992, A/sw/Haslumìne/IDT2617/2003), and a strain of pandemic (H1N1) 2009 virus (A/Hamburg/7/2009) by using Freund adjuvant. Blood samples were taken 14 days after last immunization. A vaccine containing the pandemic (H1N1) 2009 virus was produced. Swine influenza vaccines available in central Europe and the newly produced vaccine containing pandemic (H1N1) 2009 virus (A/Hamburg/7/2009) were administered to pigs (2-fold vaccination with 1–2 mL of the vaccine 21–28 days apart intramuscularly). Blood was withdrawn 7 days after second administration.

In addition, an experimental aerosol infection was conducted by using the parental strain of the most recent avH1N1 strain contained in a European swine influenza vaccine (A/sw/Haslumìne/IDT2617/2003). Blood samples were taken 10 days after infection.

The investigation of the hyperimmune serum samples detected neutralizing activity between the pandemic (H1N1) 2009 virus and European avH1N1 vaccine strains (A/sw/Netherlands/25/1980, A/sw/IDT/Re230/1992, A/sw/Haslumìne/IDT2617/2003), as well as with the cH1N1 strain A/New Jersey/8/1976
(Fort Dix reассortant). The hyper-immune serum established against pandemic (H1N1) 2009 virus also showed cross-reactivity with European avH1N1 virus. The reactions against several strains of the pandemic virus were similar, reflecting high titers against pandemic (H1N1) 2009 virus but also cross-reactions with hyperimmune serum samples of all swine influenza A virus H1N1 vaccine strains (online Appendix Table, www.cdc.gov/EID/content/16/6/1029-appT.htm).

The bivalent vaccines induced high titers of neutralizing antibodies against avH1N1 virus and human-like H3N2 virus (huH3N2). Only a low number of pigs reacted with H1N2 virus whereas the trivalent vaccine induced high neutralizing activity in serum samples of all vaccinated pigs. The vaccines induced neutralizing antibodies against pandemic (H1N1) 2009 virus. The titers were lower in comparison to those obtained for avH1N1 and not all pigs responded. The reactions were best for the vaccines containing mineral oil. Pigs vaccinated with the trivalent vaccine with carbomer adjuvant showed almost no antibodies against pandemic (H1N1) 2009 virus, although the vaccine strain reacted well in hyperimmunization tests.

A vaccine batch of the trivalent vaccine was produced that contained mineral oil instead of carbomer. All pigs vaccinated with the trivalent vaccine with mineral oil adjuvant showed a comparable level of protection (limited period of viral shedding). Mineral oil adjuvants can induce severe distress in pig herds due to their limited safety. Despite cross-reactivity between avH1N1 and ch1N1 with pandemic (H1N1) 2009 virus, the highest degree of cross-neutralization was achieved by the vaccine containing pandemic (H1N1) virus strain.

Proof of cross-reactivity was also reflected in the infection trial. Pigs infected with avH1N1 responded to avH1N1 as well as to pandemic (H1N1) 2009 virus. All results were additionally confirmed by hemagglutination inhibition assay (data not shown).

Furthermore, 1,559 pig serum samples from 195 German pig herds collected from mid-June through mid-September 2009 were tested in routine diagnostics by hemagglutination inhibition assay. All reflected almost similar results for avH1N1 and the pandemic (H1N1) 2009 virus (sero-prevalences for individual pigs were pandemic [H1N1] 2009 virus 52%, avH1N1 53%, huH1N2 28%, and huH3N2 52%; for pig herds pandemic [H1N1] 2009 virus 46%, avH1N1 46%, huH1N2 24%, and huH3N2 45%). These results suggest cross-reactivity between porcine H1N1 virus and pandemic (H1N1) 2009 virus. Despite this cross-reactivity, a vaccine consisting of pandemic (H1N1) 2009 virus is superior in terms of efficacy in comparison with vaccines already authorized in Europe.

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References


4. Influenza pandemic (H1N1), animal. Latest information on influenza A (H1N1) [cited 2010 Jan 31]. http://www.promedmail.org


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The strain). To correspond to the peak, which is between days 7 and 14 after hyperimmunization (depending on the immunization scheme), between days 7–10 after second administration of inactivated vaccines, and between days 10 and 14 after infection (depending on the immunization scheme), between days 7–10 after second administration of inactivated vaccines, and between days 10 and 14 after infection (depending on the immunization scheme).

Appendix Table. Serologic cross-reactivity between vaccine strains used in European swine influenza A virus vaccines and pandemic (H1N1) 2009 virus and influenza A viruses currently circulating in the human population, measured by neutralization test*

<table>
<thead>
<tr>
<th>Group</th>
<th>European swine influenza A virus</th>
<th>Human influenza A virus</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. pigs</td>
<td>Serum samples</td>
<td>Neutralization test (data not shown).</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hyperimmunization†</td>
<td></td>
<td></td>
</tr>
<tr>
<td>New Jersey/8/76 (cH1N1)</td>
<td>1</td>
<td>1,288 (1)</td>
</tr>
<tr>
<td>Swine/Netherlands/25/80 (avH1N1)</td>
<td>1</td>
<td>1,288 (1)</td>
</tr>
<tr>
<td>Swine/IDT/Rm23/92 (avH1N1)</td>
<td>1</td>
<td>1,288 (1)</td>
</tr>
<tr>
<td>Swine/Haseleunne/IDT2617/2003 (avH1N1)</td>
<td>2</td>
<td>8,128 (1)</td>
</tr>
<tr>
<td>Hamburg/7/2006§ (pandemic [H1N1] 2009 virus)</td>
<td>2</td>
<td>513–813 (2)</td>
</tr>
<tr>
<td>Vaccination‡</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bivalent vaccine 1 (cH1N1+H3N2)</td>
<td>10</td>
<td>129–8,128 (10)</td>
</tr>
<tr>
<td>Bivalent vaccine 2 (avH1N1+H3N2)</td>
<td>12</td>
<td>81–8,128 (12)</td>
</tr>
<tr>
<td>Bivalent vaccine 3 (avH1N1+huH3N2)</td>
<td>20</td>
<td>32–2,042 (20)</td>
</tr>
<tr>
<td>Trivalent vaccine (avH1N1+huH1N2+huH3N2)</td>
<td>28</td>
<td>129–3,236 (28)</td>
</tr>
<tr>
<td>Monovalent pandemic (H1N1) 2009 vaccine</td>
<td>6</td>
<td>129 (1)</td>
</tr>
<tr>
<td>Infection§</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* The name of this strain was changed several times; the designations Hamburg/4/2009, Hamburg/5/2009, and Hamburg/7/2009 refer to the same strain.
† Inactivated vaccines: bivalent vaccine 1 (Gripovac) contains antigens of strains A/NewJersey/8/1976 (cH1N1) and A/PortChalmers/1/1973 (H3N2), with mineral oil used as adjuvant (batch SK-01700); bivalent vaccine 2 (RESPIPORC FLU), identical with Ingelvac FLU) consists of strains IDT/Re230/1992 and IDT/Re220/1992, which are reassortant strains based on Belgian swine influenza viruses, with Al(OH)3 + mineral oil used as adjuvants (batches 019–022 08 04); bivalent vaccine 3 (RESPIPORC FLU3, identical with Gripovac 3) contains strains A/swine/Haseleunne/IDT2617/2003 (avH1N1), A/swine/Bakum/1832/2000 (huH1N2), and A/swine/Bakum/IDT1769/2003 (huH3N2), with a carbonic adjuvant (batch 005 08 06); the monovalent pandemic vaccine consists of the pandemic (H1N1) 2009 virus strain A/Hamburg/7/2009 with a carbomer adjuvant (batch 001 07 09).