isolate of serotype O123:H– is recorded in the database of the World Health Organization Collaborating Centre for Reference and Research on Escherichia and Klebsiella (F. Scheutz, pers. comm.).

Two strains of STEC O123:H– stx2d were isolated from asymptomatic persons in Germany during 1996–2000 (8). A study in Australia in 2003 reported using a strain of O123:H– stx1 stx2 ehxA from Switzerland that had been isolated from a person with diarrhea (9).

We report foodborne transmission of STEC O123:H– that resulted in a cluster of clinical cases of infection. Eating ground beef is a well-established mode of STEC transmission, particularly for serotype O157:H7. STEC serotype O123:H– has been isolated from feces of healthy lambs and sheep in Spain (10) and in southwestern Australia (9) and is considered to be among the predominant ovine STEC serotypes in these countries.

This family outbreak shows that STEC serotype O123:H–, albeit rarely described as causing human illness, can cause severe human infection. This serotype can also cause clusters of STEC infections and be transmitted by ingestion of undercooked ground beef.

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References

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LETTERS

Austrian Syndrome Associated with Pandemic (H1N1) 2009 in Child

To the Editor: In 1957, an American internist reported the preference of Streptococcus pneumoniae for the aortic valve and its frequent association with meningitis and pneumonia (1), an association now known as Austrian syndrome. This syndrome mainly occurs in middle-age men who have predisposing factors, such as chronic alcoholism, altered immune state, duodenal fistula, and ear or sinus infections.

One case of Austrian syndrome has been reported in the pediatric age group, in a 7-year-old girl in whom aortic valve endocarditis developed...
after pneumococcal meningitis infection (2). In pneumococcal endocarditis, the native aortic valve is the most frequent location of vegetation. Valve replacement must be considered to avoid development of cardiogenic shock, whereas medical treatment alone may be adequate in some cases of mitral endocarditis (3,4).

We report a previously healthy adolescent with Austrian syndrome associated with pandemic (H1N1) 2009 infection. Cardiac involvement resulted in extensive mitral valve destruction requiring surgical valve replacement.

A 13-year-old boy had cough, nasal congestion, and a fever ≥103°F for 2 weeks before being seen at a hospital. On the day he was admitted to hospital, he became unarousable, and weakness was noted on his left side. His medical history included mild asthma requiring no therapy over the past 3 years. His immunizations were up to date except for seasonal influenza and pandemic (H1N1) 2009 vaccination.

When admitted to hospital, his temperature was 38.7°C, blood pressure 97/47 mm Hg, respiratory rate 77 breaths/min, and heart rate 150 beats/min. Pupils were 3 mm in diameter and reactive to light and accommodation. Weakness and hypotonia of the left upper and lower extremities were noted. He had neck stiffness without Kernig sign or Brudzinski sign. Complete blood count showed a leukocyte count of 100 cells/mm³, 71% neutrophils, 8% bands, 15% lymphocytes, 5% monocytes, and 1% eosinophils; protein 195 mg/dL, and glucose 6 mg/dL, with a blood glucose level of 140 mg/dL. Gram stain of the cerebrospinal fluid showed gram-positive cocci in pairs. The patient was treated with intravenous ceftriaxone, vancomycin, and dexamethasone. He subsequently became unconscious and hypotensive and required intubation with mechanical ventilation and intravenous dopamine. His nasal wash sample was positive for pandemic (H1N1) 2009 RNA by real-time reverse transcription–PCR; oseltamivir (75 mg) through a nasogastric tube every 12 h for 5 days was administered. Because of heart murmur, a 2-dimensional echocardiography was conducted; it showed a large mitral valve vegetation 1.6 cm × 2.1 cm attached to the posterior mitral leaflet and mild to moderate mitral insufficiency. Because of this finding, he was transferred to Children’s Hospital of Michigan for surgical intervention.

Physical examination at Children’s Hospital showed a well-nourished adolescent who was intubated and sedated. A grade 3 systolic ejection murmur at the left lower sternal border was noted. Neurologic examination showed sluggish pupils, decreased tone of the left extremities, and bilateral Babinski sign. No meningeal signs were observed. Magnetic resonance imaging of the head showed multiple areas of infarction, with the largest being in the right middle cerebral artery distribution and a smaller one in the left frontal region (Figure). A few small scattered areas of infarction bilaterally were suggestive of cerebral embolism.

Two days after transfer, the patient underwent mitral valve replacement with a St. Jude prosthetic valve (St. Jude Medical, St. Paul, MN, USA). A 2.5-cm vegetation was found on the atrial surface of the inferior aspect of the posterior mitral leaflet involving the inferior commissure. The posterior

Lumbar puncture showed a leukocyte count of 100 cells/mm³, 71% neutrophils, 8% bands, 15% lymphocytes, 5% monocytes, and 1% eosinophils; protein 195 mg/dL, and glucose 6 mg/dL, with a blood glucose level of 140 mg/dL. Gram stain of the cerebrospinal fluid showed gram-positive cocci in pairs. The patient was treated with intravenous ceftriaxone, vancomycin, and dexamethasone. He subsequently became unconscious and hypotensive and required intubation with mechanical ventilation and intravenous dopamine. His nasal wash sample was positive for pandemic (H1N1) 2009 RNA by real-time reverse transcription–PCR; oseltamivir (75 mg) through a nasogastric tube every 12 h for 5 days was administered. Because of heart murmur, a 2-dimensional echocardiography was conducted; it showed a large mitral valve vegetation 1.6 cm × 2.1 cm attached to the posterior mitral leaflet and mild to moderate mitral insufficiency. Because of this finding, he was transferred to Children’s Hospital of Michigan for surgical intervention.

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![Figure](image-url)
leaflet associated with the vegetation was destroyed. The vegetation culture, pericardial fluid culture, tissue culture from the resected mitral valve, and 3 blood cultures yielded no bacterial growth.

The patient required ventilator support for 7 days. Follow-up computed tomography on day 8 showed a stable appearance of cerebral infarcts. Coumadin was started to prevent thrombus development at the prosthetic valve. He went home after completing a 4-week treatment course of ceftriaxone. At that time, there was still noticeable left-sided weakness of the extremities. He could only communicate by using eye movements with no verbal response.

During the 20th century influenza pandemics, secondary bacterial pneumonia was a notable cause of death. The current pandemic (H1N1) 2009 outbreak is evolving rapidly, and it is unknown if pandemic (H1N1) 2009 may lead to an increase in rare complications of pneumococcal infection, such as endocarditis. Thus, Austrian syndrome should be considered in any patient with pandemic (H1N1) 2009 complicated by pneumococcal infection and a new heart murmur.

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References

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Rickettsia sibirica mongolitimonae in Traveler from Egypt

To the Editor: Tick-borne rickettsioses are zoonoses caused by spotted fever group (SFG) Rickettsia spp. (1), which have been reported as a frequent cause of fever in international travelers (2). In Egypt, Mediterranean spotted fever caused by Rickettsia conorii transmitted by the brown dog tick, Rhipicephalus sanguineus, is known to be present, although cases are rarely documented. Moreover, an emerging pathogen, R. aeschlimannii, has been detected in Hyalomma dromedarii ticks, collected from camels, and in H. impeltatum and H. marginatum rufipes, collected from cows (3). We report a case of Rickettsia sibirica mongolitimonae infection in a French traveler who returned from Egypt.

In September 2009, a previously healthy 52-year-old man living in France was admitted to the infectious diseases unit of a hospital in Nantes, France, with a 10-day history of fever, asthenia, headache, and arthromyalgia. Three days earlier, he had returned from a 2-week trip to Egypt. He had fever (38°C), painful axillary lymphadenopathies, and an inoculation eschar surrounded by an inflammatory halo on the left scapular area (online Appendix Figure, www.cdc.gov/EID/content/16/9/1495-appF.htm), but he did not have a rash. During his travel, he had been unsuccessfully treated for headache, arthromyalgia, and diarrhea by amoxicillin-clavulanate (3 g/d), nonsteroidal antiinflammatory drugs, and gentamicin cream on the eschar for 3 d. No tick bite was reported by the patient. We suspected an SFG rickettsiosis. The patient received 200 mg doxycycline in a single dose and rapidly improved.

The immunofluorescence assay for antibodies reactive against SFG antigens showed increased levels of immunoglobulin M (titer 16) and G (titer 128). Results of Western blot with cross-adsorption assays supported the hypothesis that the infection was caused by R. sibirica mongolitimonae (1). To identify the involved rickettsiae, PCR amplifications and sequencing gltA, ompA, and ompB fragment genes of Rickettsia spp. and multispacer typing (MST), based on the sequence of variable intergenic spacers, were performed by using DNA samples obtained from an eschar biopsy and a lesion swab (4,5). A negative control (sterile water and DNA from a sterile biopsy specimen) and a positive control (DNA from R. montanensis) were included in each test. Amplicon sequencing confirmed the presence of R. sibirica mongolitimonae DNA in patient samples. The sequence homology to R. sibirica mongolitimonae DNA was ompA, 99.4%; gltA, 99.7%; and ompB, 100% (GenBank accession nos. DQ097082, DQ097081, and AF123715, respectively). The MST sequences were 100% homologous to the genotype of R. sibirica mongolitimonae MST type U (ident HA-91). We injected shell vial cultures with eschar biopsy specimens (4). Fifteen days later, positive Gimenez staining and immunofluorescence confirmed