Fatal Outbreak of Mycoplasma capricolum Pneumonia in Endangered Markhors

Stéphane Ostrowski, François Thiaucourt, Mulojon Amirbekov, Abdurahmon Mahmashoev, Lucia Manso-Silván, Virginie Dupuy, Dustmurod Vahobov, Orom Ziyoev, and Stefan Michel

A pneumonia outbreak reduced the numbers of a wild population of endangered markhors (Capra falconeri) in Tajikistan in 2010. The infection was diagnosed by histologic examination and bacteriologic testing. Mycoplasma capricolum subsp. capricolum was the sole infectious agent detected. Cross-species transmission from domestic goats may have occurred.

Mycoplasma capricolum subsp. capricolum and M. capricolum subsp. capripneumoniae are closely related subspecies of the M. mycoides cluster (1). Whereas M. capricolum subsp. capripneumoniae is the etiologic agent of contagious caprine pleuropneumonia (CCPP), a severe and typically lethal respiratory disease, M. capricolum subsp. capricolum infection is usually not fatal and instead results in chronic inflammation in a variety of organs, including joints, udder, eyes, and lungs (2). M. capricolum subsp. capricolum infection occurs worldwide and appears widespread but has rarely been found in species of small ruminants other than domestic goats and, more occasionally, sheep (2,3). This lack of evidence may be partially because few studies have applied sensitive molecular techniques for its detection in nondomestic ruminants (2,3). Domestic goats can carry M. capricolum subsp. capricolum asymptptomatically, notably in the ear canal (4), and pose an insidious risk for cross-species transmission with sympatric wild caprines (2,3).

The Study

The markhor (Capra falconeri) is an endangered wild goat in a continuous decline; the global population is <2,500 mature animals (5). In Tajikistan, <350 animals may survive in fragmented subpopulations in the remote Hazratisloh and Darvaz mountain ranges along the Afghanistan border (6). They live sedentarily over relatively small home ranges, moving <5 km per day (7). Throughout its range, the markhor has to forage in close proximity to domestic goats (8) and is therefore prone to infections of contagious agents transmitted by these animals.

During September 17–October 18, 2010, eleven markhors that displayed labored breathing and 64 markhors that had recently died were found in 5 localities (Table) in the district of Shuroabad, Khatlon Province, usually in close proximity to water sources (Figure 1, panel A). All but 4 carcasses were too scavenged for thorough examination in the field, and 1 dying adult female was sent on September 20 to the Republican Veterinary Laboratory in Dushanbe for necropsy. The most relevant necropsy findings noted in the field were the following: an abundant serous to mucopurulent nasal discharge; and internally, severe pneumonia associated with a variable level of pleural fluid. The female markhor examined in Dushanbe showed gray areas of consolidations in the apical and cardiac lobes of the right lung and yellow pleural fluid (Figure 1, panel B). The cut surface of the affected lobes revealed a fine granular texture, and mucopurulent exudate could be expressed from the bronchi. The joints, eyes, and udder were not affected. Regarding indications of CCPP, although gross lesions were limited to the thoracic cavity, fibrinous pleurisy (9) was not observed. The histopathologic findings were consistent with proliferative interstitial pneumonia associated with multifocal suppurations (Figure 1, panel C). Findings included diffuse thickening of the interlobular septa, alveolar epithelialization, and polymorphonuclear leukocytes in alveolar and bronchiolar spaces (Figure 1, panel D). Also, disseminated and abundant neutrophil infiltration of alveolar spaces was found that, when...
Fatal Pneumonia in Markhors

ruptured, created multifocal nodules. The left lung was congested.

No bacteria were isolated from pleural fluid that was obtained aseptically and inoculated onto 5% sheep blood agar. However, when a healthy 5-month-old domestic goat (C. hircus) was inoculated intratracheally with 5 mL of this pleural fluid, pneumonia developed within 8 days. To evaluate the possibility that the outbreak might have been caused by *M. capricolum* subsp. *capripneumoniae*, which was first identified in Tajikistan in 2009 (10), samples were sent to Centre de Coopération International en Recherche Agronomique pour le Développement, Montpellier, France, a reference laboratory for CCPP for the World Organisation for Animal Health. Pleural fluid from the markhor was centrifuged at low speed (500 g for 10 min) to eliminate inflammatory cells, and DNA was extracted from the supernatant with the DNeasy blood and tissue kit (QIAGEN, Courtaboeuf, France), according to the manufacturer’s instructions. Real-time PCR (11), specific for *M. capricolum* subsp. *capripneumoniae*, provided negative results, whereas a partial 16S rRNA gene sequence could be amplified and sequenced (12). BLAST analysis (http://blast.ncbi.nlm.nih.gov/Blast.cgi) showed that the sequence corresponded to that of a mycoplasma from the *M. mycoides* cluster. Markhor pleural fluid and lung tissue from the domestic goat were then injected into modified Hayflick broth (1) and onto agar plates and incubated at 37°C in 5% CO₂. Typical *Mycoplasma* colonies appeared on solid medium after 1–2 days. A clone from the markhor culture, named 10074 (1.4), was used for PCR amplification and sequencing of *fusA*, *glpQ*, *gyrB*, *lepA*, and *rpoB* partial gene sequences as described (1). The *fusA* sequence was identical to a previously submitted sequence (GenBank accession no. EF071735). All other sequences were novel.

Figure 1. Pneumonia caused by *Mycoplasma capricolum* subsp. *capricolum* in markhors (*Capra falconeri*), Tajikistan, 2010. A) Adult male markhor found dead with signs of pneumonia and no indications of emaciation. B) Disseminated gray areas of consolidation in the cardiac lobe of the right lung with mucopurulent exudate in bronchi. C) Diffuse proliferative interstitial pneumonia associated with a lesion of suppuration (hematoxylin and eosin stain; original magnification ×40). D) Interstitial pneumonia showing fibrotic thickening of alveolar walls and epithelialization of pneumocytes (hematoxylin and eosin stain; original magnification ×250).
and have been assigned new GenBank accession nos.: HQ882179 (glpQ), HQ882180 (gyrB), HQ882181 (lepA), HQ882178 (rpoB). The 5 protein-coding sequences were then concatenated and incorporated in the \( M. \) mycoides cluster phylogenetic tree (Figure 2). The strain clustered with \( M. \) capricolum subsp. capricolum and was therefore identified as belonging to this subspecies. The isolate from the inoculated domestic goat was undistinguishable from 10074 (1.4) by sequencing of the 5 housekeeping genes.

The pattern of inflammation in the right lung in the necropsied markhor was consistent with \( M. \) capricolum subsp. capricolum respiratory infection in domestic goats (2), and the presence of \( M. \) capricolum subsp. capricolum in pleural fluid, the lack of findings indicative of alternative etiologic agents, and the reproduction of the disease in the domestic goat with isolation of \( M. \) capricolum subsp. capricolum from the lungs support a causal relationship between \( M. \) capricolum subsp. capricolum and disease in markhors.

Conclusions

Although because of difficult field conditions, only 1 markhor could be thoroughly investigated, these findings support the hypothesis that \( M. \) capricolum subsp. capricolum, a newly recorded pathogen in free-ranging wild ruminants, may be responsible for the pneumonia epizootic observed in endangered markhors. This outbreak claimed \( \approx20\% \) of the population remaining in Tajikistan, and more markhor deaths might have remained undetected.

The source of \( M. \) mycoides infection in markhors is unknown, but domestic goats, which have contact with markhors, particularly in the summer, might have been responsible for the emergence of \( M. \) capricolum subsp. capricolum in this wild species. In November 2008, a disease resembling CCPP affected domestic goats and, to a lesser extent, sheep in Shuroabad District (<40 km from the area where the dead markhors were found) with a case-fatality rate of 20%–30% (13). However, the origin of this outbreak could not be investigated. Although the clinicopathologic features of the disease in the markhors resembled CCPP, \( M. \) capricolum subsp. capricolum was identified as the most probable causative agent. In fact, several mycoplasmas have been associated with respiratory diseases in ruminants, including \( M. \) capricolum subsp. capricolum infection in young domestic goats (2,14). Further testing should evaluate the presence of sick animals or asymptomatic carriers of \( M. \) capricolum subsp. capricolum in livestock and wildlife in Shuroabad District.

Environmental and nutritional stressors may exacerbate the susceptibility of ruminants to \( M. \) mycoides infections (2,15). The disease appeared at the end of summer, when markhors are forced by livestock and guard dogs to retreat to suboptimal habitats with poor forage (8). It was also the end of the dry season, when markhors may have contact with livestock at the few remaining water sources.

Findings from this study support the conclusion that the markhor is vulnerable to \( M. \) capricolum subsp. capricolum. In the generalized context of increasing encroachment of livestock into wild habitats, markhors and other wild caprines might be at risk for future mycoplasmosis outbreaks. This case emphasizes the need for continuous disease surveillance in domestic animals that have contact with valuable wildlife resources.

Acknowledgments

We thank L. Boulouha for performing the histopathologic examination; I. Ikromov and his staff who reported the outbreak, centralized reports of dead markhors, and captured the sick female markhor; and P. Zahler and 2 anonymous reviewers for their helpful comments.

The study was supported by the Deutsche Gesellschaft für Internationale Zusammenarbeit and the Wildlife Conservation Society.

Dr Ostrowski is a wildlife veterinarian and ecologist working as ecosystem health manager in central and western Asia for the Global Health Program, Wildlife Conservation Society.
His research interests concern the effects of diseases on wild host abundance and diversity in conjunction with human modifications of the environment.

References


Address for correspondence: Stéphane Ostrowski, Wildlife Conservation Society, 2300 Southern Blvd, Bronx, NY 10460, USA; email: sostrowski@wcs.org