

75°42.776'W; Sahagun, Vereda Las Llanadas, 8°56.533'N, 75°20.909'W; and Lorica, Colegio Instituto Técnico Agrícola, 9°24.067'N, 075°75.707'W). The landscape at the study sites is dominated by tropical savanna, small patches of forest, and cultivated land. Live-capture traps were set in a variety of habitats at each locality, and captured rodents were processed according to methods by Mills et al. (6). Rodents were anesthetized, and blood (by cardiac puncture) and tissue (liver, lung, spleen, heart, kidney) samples were collected into individual cryovials, placed in liquid nitrogen in the field, and transferred to freezers and stored at -80°C at the Instituto de Investigaciones Biológicas del Trópico, Universidad de Córdoba, Montería, Colombia. Rodent species were identified on the basis of morphologic analyses of formalin-fixed carcasses; chromosomal data and mitochondrial DNA sequencing of the cytochrome b gene were used to confirm identification of most antibody-positive animals. Alcohol-preserved voucher specimens are archived at the Museum of Texas Tech University (Lubbock, TX, USA).

Blood samples were tested for arenavirus immunoglobulin (Ig) G by indirect immunofluorescent antibody assays. Guanarito and Pichindé virus-infected Vero E6 cells were used as antigens on spot slides. The secondary antibody was a fluorescein-conjugated goat,

antimouse IgG. Serum samples were screened at a dilution of 1:10 and endpoint titers were measured by using serial 2-fold dilutions (1:10–1:320) (7). Attempts to amplify viral RNA in tissues by reverse transcription PCR were unsuccessful.

We collected 210 sigmodontine rodents of 3 species: 181 *Z. brevicauda*, 28 *Oligoryzomys fulvescens*, and 1 *Oecomys concolor*. Eleven serum samples, 10 from *Z. brevicauda* and 1 from *O. fulvescens* rodents, had detectable arenavirus antibody. Three *Z. brevicauda* rodent samples had antibody reactive to both Pichindé and Guanarito virus, and 7 more were positive for either Pichindé or Guanarito arenaviruses (Table).

We used only 2 viral antigens in our screening belonging to the 2 viruses that are either known to occur in Colombia (Pichindé virus) or known to be hosted by species that we captured (Guanarito virus). Among the 10 *Z. brevicauda* samples with detectable antibody, 5 reacted only to Pichindé virus antigen or their antibody titer to Pichindé virus was at least 4-fold higher than their titer to Guanarito virus (Table), suggesting those rodents were infected with Pichindé or a closely related virus. Additional studies, including isolation and sequencing are needed to definitively identify this virus.

Surprisingly, only 2 *Z. brevicauda* rodent (1.1%) had antibody only to Guanarito virus or had a 4-fold greater

titer to Guanarito virus, much lower than the 15% antibody prevalence in the same species in the Venezuelan hemorrhagic fever–endemic area, Portuguesa State, Venezuela (5). Our testing protocols differed from the earlier study, and we have not definitively identified Guanarito virus in those 3 rodents; nevertheless, this low prevalence might help explain the absence of Venezuelan hemorrhagic fever in Colombia, although inadequate surveillance is a second possible explanation.

The single antibody-positive *O. fulvescens* rodent had a low antibody titer only to Pichindé virus. This apparent 4% antibody prevalence is based on only 28 mice. The significance of this finding is not clear but may represent spillover or an undescribed arenavirus specific to the species *O. fulvescens*. Again, additional studies are needed.

Our results demonstrate the presence of ≥ 1 arenaviruses circulating among common rodent hosts in Caribbean Colombia. We emphasize that many New World arenaviruses are likely cross-reactive to the antigens we used; recovery and sequencing of viral RNA will be essential to fully characterize these viruses. Hemorrhagic fever of arenaviral origin should be included in the differential diagnosis of tropical fevers, at least in our study region. As the human population of the rural Department of Córdoba and adjacent areas of

Table. Comparison of PICHV and GTOV antibody titers in rodents after serologic screening of 210 sigmodontine rodents, Córdoba, Colombia, November 1, 2008–June 10, 2009*

Specimen no.	Species	PICHV titer	GTOV titer	Rural locality
49	<i>Zygodontomys brevicauda</i>	40	<10	Monteria
70	<i>Z. brevicauda</i>	80	10	Monteria
105	<i>Z. brevicauda</i>	40	<10	Monteria
107	<i>Oligoryzomys fulvescens</i>	20	<10	Monteria
272	<i>Z. brevicauda</i>	80	10	Monteria
289	<i>Z. brevicauda</i>	20	<10	Monteria
317	<i>Z. brevicauda</i>	40	<10	Monteria
344	<i>Z. brevicauda</i>	20	<10	Monteria
345	<i>Z. brevicauda</i>	<10	20	Monteria
209	<i>Z. brevicauda</i>	10	80	Lorica
211	<i>Z. brevicauda</i>	<10	40	Lorica

*PICHV, Pichindé arenavirus; GTOV, Guanarito arenavirus. Titers in **boldface** are ≥ 4 -fold higher than titers for the other antigen.

the Caribbean coast of Colombia continues to increase, the potential for arenavirus-related disease could become a public health concern.

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High Incidence of Guillain-Barré Syndrome in Children, Bangladesh

To the Editor: Bangladesh has achieved remarkable success in its drive to eliminate poliomyelitis; no case has been reported from that country since 2000. Still, the nonpolio incidence rate of acute flaccid paralysis (AFP) in Bangladesh is 3.25 cases per 100,000 children <15 years of age (1). Guillain-Barré syndrome (GBS), an acute polyradiculoneuropathy, is the most frequent cause of AFP (2). GBS in Bangladesh is frequently preceded by an enteric infection caused by *Campylobacter jejuni* (3). Frequent exposure to enteric pathogens at an early age may increase the incidence of GBS. We hypothesized that most AFP cases in Bangladesh can be diagnosed as GBS. Our objective was to estimate the crude incidence rate of GBS among children <15 years of age in Bangladesh.

In collaboration with the World Health Organization (WHO), the Government of Bangladesh conducts active AFP surveillance. AFP is defined as acute onset of focal or general flaccid (hypotonic) weakness without other obvious cause (e.g., trauma) in children <15 years of age.

Data on the number of reported AFP cases in Bangladesh during 2006 and 2007 were obtained. On the basis of clinical and other information routinely collected through the surveillance system, we defined a GBS case as presence of an acute flaccid (hypotonic) paralysis and symmetrical weakness (4) in the absence of injury or birth trauma.

Bangladesh is divided into 6 divisions (major administrative regions) comprising 64 districts. Crude incidence data for GBS were calculated per division and per district on the basis of the population <15 years of age reported by WHO and the Government of Bangladesh.

In 2006 and 2007, a total of 1,619 and 1,844 AFP cases, respectively, were reported in children <15 years of age, of which 608 (37%) and 855 (46%) cases, respectively, fulfilled the GBS case definition. The crude incidence rate for GBS in children <15 years of age varied from 1.5 to 1.7 cases per 100,000 population in the 3 northern divisions (Dhaka, Rajshahi, and Sylhet) and from 2.1 to 2.5 per 100,000 in the 3 southern divisions (Khulna, Barisal, and Chittagong) (online Appendix Figure, www.cdc.gov/EID/content/17/7/1317-appF.htm). Overall, the crude incidence rate of GBS in children <15 years of age varied from 1.5 to 2.5 cases per 100,000 population per year in the 6 divisions of Bangladesh. Incidence rates were high (>5.0/100,000) in the Meherpur and Barisal districts in southern Bangladesh. We found a seasonal fluctuation in the frequency of patients with GBS; the most cases occurred in May (n = 159) and the lowest in February (n = 84). GBS occurred predominantly among boys (59%).

Most incidence studies reported in the literature originate from Europe and North America (5). A recent review reported that the best estimate of the global incidence of GBS in children <15 years of age is