Human adenovirus (HAdV) serotype 14 is rarely identified. However, an emerging variant, termed HAdV-14p1, recently has been described in the United States in association with outbreaks of acute respiratory disease with high rates of illness and death. We retrospectively analyzed specimens confirmed positive for HAdV by immunofluorescence, virus culture, or real-time PCR during July 1, 2009–July 31, 2010, and describe 9 cases of HAdV-14p1 infection with characteristic mutations in the fiber and E1A genes that are phylogenetically indistinguishable from the viruses previously detected in the United States. Three patients died; 2 were immunocompromised, and 1 was an immunocompetent adult. We propose that surveillance should be increased for HAdV-14p1 and recommend that this virus be considered in the differential diagnosis of sudden-onset acute respiratory disease, particularly fatal infections, for which an etiology is not clear.
Human adenoviruses (HAdVs) were identified independently by 2 groups during the early 1950s (1,2). HAdVs are nonenveloped, linear double-stranded DNA viruses encapsidated within a protein shell and have been categorized into 6 species (A–F) that contain 51 immunologically distinct serotypes (3). HAdVs most commonly cause acute respiratory disease; however, depending on the infecting HAdV serotype and tropism resulting from differential host receptor use, the wide variety of symptoms can include pneumonia, febrile upper respiratory illness, conjunctivitis, cystitis, and gastroenteritis (4). The severity of disease appears dependent on the immunocompetence and cardiopulmonary health of the host, and the spectrum of disease can range from subclinical to severe respiratory distress and death (4). Immunocompromised patients (especially bone marrow transplant [BMT] recipients) are particularly susceptible to HAdV infection, resulting in severe illness and deaths, whereas illness in immunocompetent patients generally resolves without major complication.

HAdV species B comprises 2 subspecies: B1 (including HAdV-3, -7, -16, -21, and -50) and B2 (HAdV-11, -14, -34, and -35). The subspecies B2 HAdV-14 (agent de Wit) was originally identified as an etiologic agent of acute respiratory disease in military recruits in the Netherlands during 1955 (5). Despite reports of subsequent outbreaks in Europe and Asia during the 1950s and early 1960s (6), global surveillance in the subsequent decades had not identified circulation of this serotype until spring 2006, when HAdV-14 emerged as a cause of a major proportion of acute febrile respiratory illness in military bases across the United States (7–9). In 2007, community-associated outbreaks were identified in California and New York (10), and during March–June 2008, ≈140 cases of HAdV-14 were identified in Oregon, Washington, and Texas. Overall, 38% of patients were hospitalized, 17% were admitted to intensive care units, and 5% died. During September 2008, an outbreak was reported with 32 cases of pneumonia on Prince of Wales Island off the coast of Alaska (11). In the United States, the earliest documented case of HAdV-14 infection, by retrospective testing, occurred in California during December 2003, and the most recent evidence of HAdV-14 had been in Pennsylvania during June 2009 (7). The virus has circulated uninterrupted in some military recruit camps (A.E. Kajon, unpub. data).

Sequence analysis of the fiber gene of HAdV-14 associated with the recent outbreaks has shown a 6-bp deletion resulting in a 2-aa deletion (lys-glu) at positions 251 and 252 in the knob region compared with the de Wit HAdV-14p prototype strain (10). The 252glu site is conserved in all other species B HAdVs and is located in the F–G loop of the fiber protein knob near a putative host receptor binding site. Further sequencing of the hexon gene hypervariable regions 1–7 and the E1A genes demonstrated that this emerging HAdV-14 was clearly separable from the reference strain de Wit (genome restriction type HAdV-14p) (9,11). This new genomic variant has been designated HAdV-14p1 on the basis of novel restriction profiles. Viral receptor binding and internalization studies that used recombinant HAdV-14p1 fiber regions containing the 2-aa deletion did not demonstrate substantial phenotypic differences between the new agent and the prototype virus (12). This finding suggests that HAdV-14p1 could be an immune escape mutant that has lost a potential neutralizing epitope, has modified postinternalization steps, or has enhanced binding to host cells through a yet-unidentified viral receptor.

However, no evidence suggests that HAdV-14p1 has reemerged because of altered virulence. Despite the mutations in the fiber and EIA genes, no other genetic differences with the prototype HAdV-14 clearly explain differing disease severity in US outbreaks (13). This observation suggests that reemergence of this agent might be more likely to have resulted from spread of an infectious agent in immunologically naive populations and changes in the rate of specific immunity in host populations over time. The recent whole-genome analysis of HAdV-14p1 in mild and severe infections supports this idea (13). In addition, reports of recent HAdV-14 and HAdV-14–11 outbreaks in Asia and the United States have detailed similar dynamics (14,15). The observed characteristics (including periods of little activity punctuated by distinct outbreaks with occasional severe disease and death) may simply be the natural pattern of adenovirus species B2 respiratory pathogens.

Whether HAdV-14p1 had circulated elsewhere before, or at the same time as, the outbreaks in the United States and whether it is currently circulating in Europe is unclear. In this report, we show that recent infection and deaths associated with HAdV-14p1 infection have occurred in Ireland and that this virus is genetically indistinguishable from HAdV-14p1 described in the United States.

Materials and Methods

Study Period

The study period was July 1, 2009–July 31, 2010. The study comprised 29 cases confirmed adenovirus positive by indirect immunofluorescence assay (IFA), viral culture, or real-time quantitative PCR (qPCR) at the National Virus Reference Laboratory (NVRL, Dublin, Ireland).

Virus Culture and Restriction Enzyme Analysis

HAdVs were cultured at NVRL and Lovelace Respiratory Research Institute (Albuquerque, NM, USA). Isolates were passaged once in human embryonic lung
carcinoma cells in 25-cm² flasks for detection of cytopathic effects and then subsequently expanded in 75-cm² flasks for extraction of viral DNA for restriction enzyme analysis as described (16). Viral DNA was digested with the same panel of endonucleases used to characterize the North American strain of HAdV-14 (7).

**Indirect Immunofluorescence**

We performed IFA for influenza A virus, influenza B virus, parainfluenza viruses 1–3, respiratory syncytial virus, and HAdV. We used the respiratory virus panel kit (Biotrin, Dublin, Ireland) and followed the manufacturer’s instructions.

**Molecular Analysis**

We performed qPCR for HAdV as described (17). Adenoviruses were initially typed by using partial hexon gene sequencing as described (17,18). We extracted 200 μL of serum, plasma, or nasopharyngeal aspirate by using the QIAamp DNA Mini Kit (QIAGEN, Crawley, UK) according to the manufacturer’s instructions or by using the MagNA Pure automated extraction platform (Roche, Lewes, UK) with an external lysis step. The final elution volumes for both methods was 50 μL. Real-time PCR specific for HAdV-11 and HAdV-14 (19) provided by the Centers for Disease Control and Prevention (Atlanta, GA, USA) was performed by using the ABI7500 SDS platform (Applied Biosystems, Foster City, CA, USA) with 5 μL of template and the Platinum qPCR SuperMix-UDG kit (Invitrogen, Paisley, UK) with 0.5-μmol/L forward and reverse primers and 0.1-μmol/L probe labeled at the 5’ end with 6-carboxyfluorescein and a 3’ quencher dye in 25-μL single-plex assays performed with the following cycling conditions: 50°C for 2 min, then 95°C for 2 min, followed by 45 cycles of 95°C for 15 sec and 60°C for 1 min, with data acquisition in the anneal/extension phase. Complete HAdV-14 fiber, E1A, and hexon genes were amplified as described (7) and sequenced bidirectionally. Nucleotide sequence data for HAdV-14p1 fiber, E1A, and hexon genes were submitted to GenBank (accession nos. HQ163915, HQ163916, and HQ265808).

**Phylogenetic Analysis**

The fiber, E1A, and hexon genes of the Ireland HAdV-14p1 strains were compared with recently described US HAdV-14p1 strains (7) and reference sequences from HAdV B2 subgenera obtained from GenBank. The accession numbers for all sequences are included on the tree (Figure 1). Lasergene version 8 (DNASTAR, Madison, WI, USA) was used for contiguous assembly (20), and the sequences were aligned by using ClustalW (21) implemented in Bioedit version 7.05 (22). Phylogenetic trees were constructed by using the maximum-likelihood method described in previous reports (7,23) and summarized in Table 1. The trees were built in PAUP* (23) on the basis of the HKY85 model of evolution and for the fiber tree also with a β distribution and used midpoint rooting. Bootstrap resampling (n = 1,000) was performed by using the neighbor-joining algorithm. Scale bars indicate nucleotide substitutions per site.
method under the HKY85 substitution model that used PAUP* version 4.0 β 10 (23), and bootstrapping with 1,000 replicates was used to analyze the stability of the tree topology.

**Results**

We retrospectively tested clinical specimens received by NVRL from 29 patients with positive results for HAdV by IF, virus culture, or qPCR during the study period by using specific HAdV-11 and -14 real-time assays. All 29 samples were negative for HAdV-11; however, 9 (31%) samples tested positive for HAdV-14 (Table). All patients sought care or were previously hospitalized during November 2009–July 2010. Of the 9 patients, 7 (78%) were male, 6 (67%) patients were ≤4 years of age, and the remaining 3 patients were ≥34 years of age. No epidemiologic links were known between any of the patients. No geographic clustering was observed; the cases were distributed throughout Ireland and involved several hospitals. All respiratory samples were IFA negative for the following respiratory viruses: influenza A, influenza B, parainfluenza viruses 1–3, and respiratory syncytial virus. Our earliest identified HAdV-14–positive specimen (November 10, 2009; case-patient 1 in Table) was positive for HAdV-14. A second HAdV-14–positive specimen from this patient on November 8 and a nasopharyngeal aspirate collected on October 29. No in

Table. Clinical characteristics of patients with confirmed HAdV-14p1, Ireland, July 1, 2009–July 31, 2010*

<table>
<thead>
<tr>
<th>Case no.</th>
<th>Age/sex</th>
<th>Location</th>
<th>Date</th>
<th>Sample type</th>
<th>Previous condition</th>
<th>Clinical characteristics</th>
<th>Outcome</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>4 y/M</td>
<td>Dublin</td>
<td>2009 Nov</td>
<td>NPA</td>
<td>Pierre Robin syndrome (craniofacial abnormality)</td>
<td>Fever, tachypnea, cough</td>
<td>Survived</td>
</tr>
<tr>
<td>2</td>
<td>7 mo/M</td>
<td>Dublin</td>
<td>2010 Mar</td>
<td>NPA</td>
<td>None known</td>
<td>Bronchiolitis</td>
<td>Survived</td>
</tr>
<tr>
<td>3</td>
<td>1 mo/M</td>
<td>Dublin</td>
<td>2010 May</td>
<td>NPA</td>
<td>None known</td>
<td>Bronchiolitis, diarrhea, vomiting</td>
<td>Survived</td>
</tr>
<tr>
<td>4</td>
<td>8 d/F</td>
<td>Dublin</td>
<td>2010 May</td>
<td>Serum (+2), urine</td>
<td>Preterm birth at 35 weeks' gestation</td>
<td>Hypotonic, abnormal LFT results</td>
<td>Died</td>
</tr>
<tr>
<td>5</td>
<td>46 y/F</td>
<td>Kilkenny</td>
<td>2010 May</td>
<td>Other/lung biopsy</td>
<td>Smoking, high BMI</td>
<td>Community-acquired pneumonia, sepsis</td>
<td>Died</td>
</tr>
<tr>
<td>6</td>
<td>34 y/M</td>
<td>Dublin</td>
<td>2010 May</td>
<td>BAL</td>
<td>Post-BMT, neutropenic</td>
<td>Unilateral pulmonary infiltrate, fever</td>
<td>Died</td>
</tr>
<tr>
<td>7</td>
<td>4 mo/M</td>
<td>Dublin</td>
<td>2010 Jul</td>
<td>NPA</td>
<td>None known</td>
<td>Bronchiolitis, diarrhea, vomiting</td>
<td>Survived</td>
</tr>
<tr>
<td>8</td>
<td>14 d/M</td>
<td>Cork</td>
<td>2010 Jul</td>
<td>Plasma (+2)</td>
<td>None known</td>
<td>ARDS</td>
<td>Survived</td>
</tr>
<tr>
<td>9</td>
<td>48 y/M</td>
<td>Dublin</td>
<td>2010 Jul</td>
<td>BAL</td>
<td>HIV+ (RNA &lt;50) HCV+ (RNA not detected); CD4 436 (15%)</td>
<td>Pneumonia</td>
<td>Survived</td>
</tr>
</tbody>
</table>

*HAdV-14p1, human adenovirus serotype 14p1; NPA, nasopharyngeal aspirate; LFT, liver function test; BMI, body mass index; BAL, bronchoalveolar lavage; BMT, bone marrow transplant; ARDS, acute respiratory distress syndrome; +, positive; HCV, hepatitis C virus; CD4, CD4 cells/mL.
in the hexon region the Ireland sequences exhibited much lower sequence identity to the HAdV-11a and -11p strains analyzed in the study; the HAdV-11a and -11p hexon genes were much more similar to each other and branched separately on the tree (Figure 1, panel C). Viral DNA extracted from each of the 8 isolates of HAdV-14 yielded profiles identical to each other and identical to those reported for the North American strain of HAdV-14. The Ireland isolates were therefore identified as corresponding to genome type 14p1 on the basis of the distinct Bci, BstEII, and PstI restriction patterns (Figure 2).

Of the 9 HAdV-14p1–infected patients, 8 (90%) had respiratory symptoms: pneumonia was diagnosed for 4, bronchiolitis for 3, and acute respiratory distress syndrome (ARDS) for 1. The other patient (case-patient 4; a premature infant born at 35 weeks) was hypotonic at birth with abnormal liver function (Table). Major underlying medical conditions were reported in 4 patients: a BMT recipient; a premature neonate; a patient co-infected with HIV and hepatitis C virus; and a patient with the craniofacial abnormality Pierre Robin syndrome, who had a prior tracheostomy and percutaneous enteral gastrostomy.

ARDS developed in the BMT recipient (case-patient 6) 56 days posttransplant; lymphocyte count had not recovered and was at undetectable levels (<0.3/mm³). The patient co-infected with HIV and hepatitis C virus was receiving antiretroviral therapy and had undetectable HIV RNA (<50 copies/mL), a CD4 count of 436 cells/mL (15%), and a resolved hepatitis C virus infection. No underlying conditions were reported for the 5 remaining patients. Three patients died in May 2010: the premature neonate, the BMT recipient, and a 46-year-old woman. The woman had no known notable medical history but had a high body mass index and smoked cigarettes (Table). Two patients (case-patients 4 and 8) received extracorporeal membrane oxygenation. Case-patient 4 had a natural killer (NK) cell deficiency and died. The nature of the NK cell defect in the premature infant was not defined, and bone marrow was analyzed with no pathologic findings noted. Leukocyte screen, however, showed a lack of NK cells; additional screen for immunodeficiencies (such as severe combined immunodeficiency) proved inconclusive, and a human leukocyte antigen–B57–negative haplotype was recorded. Case-patient 8, however, for whom ARDS had been diagnosed, received prolonged (>6 weeks) extracorporeal membrane oxygenation and recovered. Four patients had an unremarkable full recovery.

Of the 9 patients positive for Ireland HAdV-14p1, serum or plasma samples of 2 were submitted for HAdV DNA quantification. The preterm infant (case-patient 4) had a HAdV load in serum of 8.18 log₁₀ viral genomes/mL, and the infant with ARDS (case-patient 8) had a plasma adenoviral load of 8.83 log₁₀ viral genomes/mL. The immunocompetent 46-year-old woman (case-patient 5) had a viral load of 4.80 log₁₀ genomes/g from a postmortem lung biopsy specimen.

**Discussion**

We have demonstrated that the HAdV-14p1 strain first identified in the United States is now circulating in Ireland and is associated with substantial illness and with 3 deaths: a BMT recipient, a neonate, and (most notably) an immunocompetent, apparently otherwise healthy adult. The role of HAdV-14p1 in the deaths remains to be elucidated because 2 of the 3 deaths occurred in substantially immunocompromised patients. Conversely, other patients who were also immunocompromised recovered unremarkably, and fatal severe pneumonia and ARDS are known to occur in immunocompetent adults infected with other HAdV serotypes (24). Nevertheless, of concern in the cohort described here is the third death, which occurred in a previously well patient. The immunocompetent woman was a smoker, and smoking has been identified as an independent risk factor possibly facilitating transmission of HAdV-14p1 (11). However, the low rate of infections (5%) observed by Esposito et al. (11) in household contacts of persons infected with the emerging virus suggests that infection is unlikely to spread in the community. Efficient transmission of HAdV-14p1 appears to require close physical contact, as supported by the observation that in barracked communities used for military trainees in the United States, antibody titers demonstrated recent exposure after entry to the facility (25). Of interest in the HAdV-14p1 cases identified in the present study is the higher frequency of male patients (78%) infected;
this observation has been reported by other groups (19,26), although the numbers identified in our study are insufficient to make definitive conclusions. Previous reports have noted that prior vaccination with HAdV-7, a subspecies B1 virus, of military recruits may confer cross-protection, presumably from heterotypic immunity, and that this vaccine could potentially be used to reduce HAdV-14p1 disease severity (27,28). Trei et al. have reported on the spread of HAdV-14 in 2007 from a large military training facility in Texas to secondary sites across the United States and to South Korea, despite the institution of active surveillance for acute respiratory disease and prevention and control measures (29). Despite evidence of decreased spread from military personnel to family members outside the training population, many recent military graduates may have been asymptomatic and incubating the virus, which presents a possible scenario for the subsequent dissemination of this virus.

Our findings suggest that clinicians and other health care workers should consider HAdV-14p1 in the differential diagnosis of community-acquired pneumonia. No evidence indicates that HAdV-14p1 infection is substantially more severe than that by other HAdV serotypes; however, we propose increased surveillance for this emerging agent in Europe and elsewhere in immunologically naive populations. In addition, we suggest the retrospective investigation of HAdV-positive specimens for HAdV-14p1, particularly from untyped viruses from patients who had severe disease and who died without an established etiology to help determine the time and location of reemergence of this HAdV in Europe.

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Dr Carr is a clinical scientist in the NVRL, University College Dublin, Ireland. His research interests include the molecular epidemiology and pathogenesis of respiratory viruses infecting humans.

References


Address for correspondence: Michael J. Carr, National Virus Reference Laboratory, University College Dublin, Dublin 4, Ireland; email: michael.carr@ucd.ie