African Swine Fever Virus p72 Genotype IX in Domestic Pigs, Congo, 2009

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African swine fever virus p72 genotype IX, associated with outbreaks in eastern Africa, is cocirculating in the Republic of the Congo with West African genotype I. Data suggest that viruses from eastern Africa are moving into western Africa, increasing the threat of outbreaks caused by novel viruses in this region.

African swine fever (ASF) is a serious disease of domestic pigs caused by a DNA arbovirus (African swine fever virus [ASFV]) belonging to the family Asfaviridae (I). Its highly contagious nature and ability to spread over long distances make it 1 of the most feared diseases of pigs; it causes devastating effects on pig production as manifested in the Caucasus since its introduction from southeastern Africa during 2007 (2). Considerable spread of ASF has been reported in western Africa during the past 20 years, and, except for in Côte d’Ivoire, the disease remains endemic (3). Because discernible ASFV serotypes are lacking, the field strains are grouped genetically by using sequencing of the C-terminus of the p72 protein, which discriminates 22 genotypes (4,5). Genotype I is historically associated with outbreaks in western Africa, whereas viruses from southern and eastern Africa have higher heterogeneity, with all 22 known genotypes having been recorded within the region (5–7).

The Republic of the Congo, located in western-central sub-Saharan Africa, shares borders with the Cabinda enclave of Angola, the Democratic Republic of the Congo, Central African Republic, Cameroon, and Gabon. The last ASF outbreaks in Congo were reported to the World Organization for Animal Health (OIE) during 2003. Since then, the disease has been officially declared endemic but without quantitative data. Sampling and characterization of currently circulating field strains from this region of western-central Africa are needed to fully understand virus spread and maintenance. Such data will have implications for regional control in western Africa.

The Study

During August 2009, a United Nations Food and Agriculture Organization mission was undertaken with local counterparts in Congo to support development of an action plan to control ASF. A key factor in selecting the sites sampled was inclusion of the main pig-producing, marketing, and consuming areas, with a particular focus where suspected ASF outbreaks had been recently reported. From the survey conducted, 86 samples comprising serum (35 samples), whole blood (44 samples), and tissues (7 samples) were collected from 80 domestic pigs in the departments of Brazzaville, Pointe Noire, Kouilou, Bouenza, Niari, and Cuvette (Figure). These departments contain 80%–90% of the country’s pig population. Samples also were collected from Pool in southeastern Congo, where a recent hemorrhagic disease outbreak, characterized by case-fatality rates of ≥80%, had been reported. Clinical material was sent to the European Union Reference Laboratory for African Swine Fever (Centro de Investigación en Sanidad Animal, Madrid, Spain) for confirmatory diagnosis and characterization of the ASFV strain(s) responsible for the outbreak(s).

Specific ASF antibodies were detected by using the OIE-prescribed assays (8) in 7 of 35 serum samples analyzed. All positive serum samples were from animals that had survived the ASF outbreaks in Bouenza, Niari, and Pointe Noire during 2008. For ASFV genome detection, OIE-prescribed PCRs (8) were performed on DNA extracted from 28 serum samples, 44 blood samples, and 7 tissue homogenates. A positive result was obtained in 17 (21%) of samples analyzed, indicating ASF in all the departments where the survey was conducted except for Kouilou.

Subsequently, 5 hemadsorbent Congo ASFVs (Table) were isolated in porcine peripheral blood macrophages (8). Viral DNA was extracted, and 3 different sets of primers were used for ASFV genotyping. A region of 478 bp at the C-terminal end of the p72 protein and the full-length sequence of p54 gene were amplified by using primers p72U/D and 89/722, respectively (6). We compared the sequence from each of the p72 and p54 amplicons with homologous sequences representative of each previously
described p72 and p54 genotype (5,6). A rooted minimum-evolution tree was constructed by using MEGA4.0 software and the p-distance nucleotide substitution model (9). Two different ASFV phylogenetic profiles were found in western and southeastern Congo. The phylogeny inferred for the ASFV isolates from eastern Congo (Con09/Pk45, Con09/Bzz020, Con09/Abo) placed them within p72 and p54 genotype IX with viruses collected from recent outbreaks in Kenya and Uganda during 2006–2007 (6,7). By contrast, ASFV isolates from western Congo (Con09/Ni16, Con09/PN003) clustered in p72 genotype I and p54 genotype Ic (6) comprising historical isolates from western Africa, such as Ang72 and Kat67, and the first ASFV isolated from Europe, the Portuguese Lisbon57 isolate (online Appendix Figure, www.cdc.gov/EID/content/17/8/101877-appF.htm).

To provide higher resolution, the tetrameric tandem amino acid repeat sequences within central variable region (CVR) of the B602L gene were analyzed (5–7,10). Primers CVR1 (5′-ACTTTGAAACAGGAAAC (AT)AATGATG-3′) and CVR2 (5′-ATATTTTGTGATATGTGGGGTGCCTTGCT-3′) were designed in this study selected from complete genome sequences of ASFV strains available in GenBank.

Under the same thermal cycling conditions used for full p54 gene amplification, amplicons of 600–650 bp were generated from the isolates from eastern Congo, whereas the estimated size of the amplicons from western Congo was 500 bp (data not shown). Analysis of the tandem amino acid repeat sequences within the CVR demonstrated 3 different variants cocirculating in the country, 2 within the p72 type I virus genotype. The type of CVR sequence (AABNAATDBNAAAA) identified in the isolate from Pointe Noire (Con09/PN003) was identical to CVR subgroup XIII (7) that contains early isolates from Angola (Ang70, Ang72) and Portugal (Lis57). However, the Con09/Ni16, also classified within p72 and p54 genotypes I and Ic, showed a unique CVR sequence (AAAAAAAAAF) not previously described but most similar to viruses from Burundi and Kenya in CVR subgroup XXVI (7,11). As for p72 and p54 genotyping, isolates from eastern Congo were related to CVR subgroup XXIV, which contains isolates obtained during the 2006 and 2007 outbreaks in Uganda and Kenya (6,7).

Conclusions

We confirmed ASF in 5 of the 6 departments in which surveillance was conducted during August 2009 in Congo. Genotyping of 5 ASFV isolates from Congo resulted in identification of genetically distinct viruses circulating simultaneously in the country. In eastern districts of Congo, viruses were most genetically similar to those recovered from the outbreaks in Kenya and Uganda during 2006 and 2007. Therefore, genotype IX, associated with ASF outbreaks in Western Africa, was introduced into Congo in 2006–2007, subsequently circulating throughout the country until 2009.
The continuing spread of ASFV.


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Dr. Gallardo is the laboratory research coordinator of the European Reference Laboratory for African Swine Fever at the Centro de Investigación en Sanidad Animal. Her research focuses mainly on molecular characterization and diagnosis of ASFV.

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