Hepatitis E Virus Outbreak in Monkey Facility, Japan

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An outbreak of hepatitis E virus occurred in an outdoor monkey breeding facility in Japan during 2004–2006. Phylogenetic analysis indicated that this virus was genotype 3. This virus was experimentally transmitted to a cynomolgus monkey. Precautions should be taken by facility personnel who work with monkeys to prevent infection.

Wild or reared monkeys have been used as disease models in animal facilities worldwide. Because disease caused by hepatitis E virus (HEV) is a zoonosis (1–4), monkeys might be infected. We examined the prevalence of antibodies against HEV in serum and fecal samples collected from monkeys in animal facilities at the Primate Research Institute of Kyoto University in Japan for 6 years (2004–2009). We found that spontaneous infection and transmission of HEV occurred in a monkey facility.

The Study

There are 9 monkey colonies (A–I) at the Primate Research Institute of Kyoto University. Colonies A–G contained Japanese monkeys (Macaca fuscata), and colonies H and I contained rhesus monkeys (Macaca mulatta). Each colony was bred in a separate outdoor breeding facility. A total of 588 monkey serum samples were collected during September–November 2004–2009 and tested for IgG and IgM against HEV and for HEV RNA by ELISA or reverse transcription PCR (RT-PCR) as described (5–7). Samples from colonies G and F were collected during 2004–2006, whereas in 2009 samples were collected from colonies A, C, D, and I.

The prevalence of IgG against HEV was 0% in 2004, 20.0% in 2005, and 78.5% in 2006, followed by a gradual decrease to 35.9% in 2009 (Table 1). The prevalence of IgM against HEV increased from 0% in 2004 to 2.5% in 2005 and to 6.6% in 2006, and then decreased to 1.1% in 2007 and 0% in 2008 and 2009.

IgG against HEV was not detected in any of the 9 colonies in 2004, indicating that HEV infection did not occur before October 2004. However, in 2005, the prevalence of IgG reached 100% in colony D and 20% in colony G (Figure 1). ELISA titers were high, ranging from 0.293 to 1.641 in colony D and from 0.230 to 0.845 in colony G. These results suggested that HEV infection occurred after October 2004 in the monkey facility. The prevalence of IgG was higher in colony D than in colony G, and IgM was not detected in colony D, suggesting that HEV infection occurred earlier in colony D than in colony G. These colonies adjoined each other, indicating that the first HEV infection occurred in colony D and was then transmitted to colony G. Colonies A, C, D, E, and H each had an IgG prevalence of 90%–100%, and colonies B and G had an IgG prevalence >80% in 2006 (Figure 1). These results demonstrated that infection spread over a large area, except for colony F, during 2005 and 2006.

To compare the kinetics of IgG formation during 2004–2009, serum samples from 25 monkeys whose peak ELISA optical density (OD) values for IgG against HEV were each higher than 1.0 were selected. In most monkeys, OD values for IgG increased rapidly and then decreased gradually year by year. The kinetic pattern of monkey M1543 was different from those of other monkeys that had high OD values (2.568–2.738). IgM was detected exclusively in this monkey in 2006 (OD value 0.620).

Serum samples from the 25 monkeys were used to detect HEV RNA by RT-PCR. Four serum samples were positive for HEV RNA; all were from the same monkey (M1543) from which samples were collected in 2006, 2007, 2008, and 2009. Nucleotide sequences of 348 bp coding the partial open reading frame 2 showed 100% identity. This result indicated that monkey M1543 was infected persistently with HEV and produced virus continuously.

To examine whether HEV was present in feces, 2 fecal samples were collected from monkey M1543 in September and November 2009 for detection of HEV RNA. Both samples were positive for HEV RNA. Nucleotide sequences of these samples were identical to those detected from serum samples.

Table 1. Prevalence of IgG and IgM against hepatitis E virus in monkeys at monkey facility, Japan, 2004–2009

<table>
<thead>
<tr>
<th>Year</th>
<th>IgG</th>
<th>IgM</th>
</tr>
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<tbody>
<tr>
<td>2004</td>
<td>0/110</td>
<td>0/110</td>
</tr>
<tr>
<td>2005</td>
<td>24/120 (20.0)</td>
<td>3/120 (2.5)</td>
</tr>
<tr>
<td>2006</td>
<td>96/121 (78.5)</td>
<td>8/121 (6.6)</td>
</tr>
<tr>
<td>2007</td>
<td>73/96 (76.0)</td>
<td>1/96 (1.1)</td>
</tr>
<tr>
<td>2008</td>
<td>47/90 (52.2)</td>
<td>0/90</td>
</tr>
<tr>
<td>2009</td>
<td>18/51 (35.3)</td>
<td>0/51</td>
</tr>
</tbody>
</table>

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Primers were designed on the basis of sequences of swine HEV (GenBank accession no. AB248522), and RT-PCR was performed to amplify the viral genome except for the N terminus noncoding region. This strain was designated the monkey HEV Inuyama strain (JQ026407). Phylogenetic analysis of its genome indicated that this strain belongs to HEV genotype 3 (Figure 2). Infectivity of the monkey HEV strain was examined ex vivo with a human hepatocarcinoma cell line (PLC/PRF/5), and in vivo with 2 HEV-negative cynomolgus monkeys. Both experiments showed that the virus was infectious (online Technical Appendix Figures 1 and 2, wwwnc.cdc.gov/EID/pdfs/12-0884-Techapp.pdf).

A total of 94 human serum samples were collected from staff of the Primate Research Institute and subjected to ELISA for detection of IgG and IgM against HEV. All serum samples were negative for IgM against HEV, but the prevalence of IgG was 6.9% in 2007, 9.7% in 2008, and 11.8% in 2009, although differences among these years were not significant (p>0.05) (Table 2). No HEV RNA was detected in serum samples, and none of the staff had symptomatic hepatitis E during the 6-year study.

Conclusions
We conducted long-term monitoring of HEV infection in monkeys and report natural infection and transmission of HEV in a monkey facility. We sought to determine the source of the HEV outbreak and where HEV was intro-
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Dr Yamamoto is an associate professor at the University of Toyama, Toyama, Japan. His research interests are zoonoses, especially surveys of simian herpes B virus and hepatitis E virus infections of laboratory animals; and mouse and monkey Mx genes.

References


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Technical Appendix

Experimental Infection of 2 Cynomolgus Monkeys with Hepatitis E Virus, Japan

Growth of Monkey Hepatitis E Virus on PLC/PRF/5 Cells

A hepatocarcinoma cell line (PLC/PRF/5) was cultured in a 25-cm² culture bottle containing 5 mL of Dulbecco modified Eagle medium and 10% fetal calf serum. A 10% suspension of a monkey fecal specimen (2 mL) was placed on PLC/PRF/5 cells. After adsorption at 37°C for 1 h, cells were washed 3x with phosphate-buffered saline and 8 mL of maintenance medium (medium 199; Invitrogen, Carlsbad, CA, USA) containing 2% (vol/vol) heat-inactivated fetal calf serum and 10 mmol/L MgCl₂. Culture medium was replaced every 3 days and used for detection of hepatitis E virus (HEV) antigen and HEV RNA.

Infectivity of Monkey HEV

Two cynomolgus monkeys (Macaca fascicularis), M3471 (21-year-old male) and M4530 (13-year-old male), were inoculated intravenously with 2 mL of suspension (titers of HEV RNA were unknown). Fecal samples were collected daily and used for detection of HEV RNA. Serum samples were collected weekly to detect HEV RNA, HEV-specific antibodies, and levels of alanine aminotransferase and aspartate aminotransferase. All monkey experiments were reviewed and conducted according to the Guides for Animal Experiments performed at the National Institute of Infectious Diseases (code 510001). Primates were housed individually in biosafety level 2 facilities.

Cell Culture of Monkey HEV

To examine infectivity of the monkey HEV Inuyama strain in fecal samples, a fecal suspension from monkey M1543 was placed on PLC/PRF/5 cells. HEV antigen and RNA in cell culture supernatant were detected by ELISA and reverse transcription PCR, respectively. HEV antigen was detected on day 68, and the optical density value increased exponentially until
peaking on day 110 postinfection (Technical Appendix Figure 1). HEV RNA was detected on day 50 postinfection, and the nucleotide sequence of 348-bp PCR product showed 100% identity with that of the original fecal specimen. Extensive virus growth was found constantly in the second passage, as indicated by earlier detection of HEV antigen and RNA in the culture medium (Technical Appendix Figure 1). These results indicated that monkey M1543 secreted infectious HEV.

**Transmission of Monkey HEV from Japanese Monkeys to Cynomolgus Monkeys**

Infectivity of the monkey HEV Inuyama strain was also confirmed by an animal experiment. Two cynomolgus monkeys (M3471 and M4530) were inoculated with 2 mL of fecal suspension, and biochemical, serologic, and virologic markers were monitored (Technical Appendix Figure 2). HEV RNA was detected within 1 week after infection in serum samples from both monkeys. HEV RNA was detected in feces on days 7–12 postinfection in monkey M3471 and on days 8–11 postinfection in monkey M4530. Increases in IgG and IgM titers were found in these animals, indicating that the HEV strain isolated from the Japanese monkey was infectious. However, neither cynomolgus monkeys showed increased alanine aminotransferase and aspartate aminotransferase levels during the experiment.

![Technical Appendix Figure 1. Replication of monkey hepatitis E virus (HEV) in a hepatocarcinoma cell line, Japan. PLC/PRF/5 cells were infected with a monkey fecal specimen positive for HEV RNA or culture supernatant. Culture supernatants were collected every 3 days and used for detection of HEV antigen by ELISA. Circles indicate fecal specimen–infected PLC/PRF/5 cells, and triangles indicate culture supernatant–infected PLC/PRF/5 cells. Supernatant was collected on day 120 from fecal specimen–infected PLC/PRF/5 cells.](image-url)
Technical Appendix Figure 2. Kinetics of biochemical, serologic, and virologic markers after challenge of 2 cynomolgus monkeys with hepatitis E virus (HEV), Japan. Monkeys M3471 (A) and M4530 (B), which were negative for HEV antibody, were infected with monkey HEV. HEV RNA in serum and fecal samples was monitored by reverse transcription PCR. Circles indicate IgG in serum, triangles indicate IgM in serum, squares indicate alanine aminotransferase levels, and diamonds indicate aspartate aminotransferase levels. -, negative; +, positive; OD$_{492}$, optical density at 492 nm.