We calculated rates of foodborne and waterborne infections reported to the health department in Victoria, Australia, during 2000–2009 for elderly residents of long-term care facilities (LTCFs) and the community. We used negative binomial regression to estimate incidence rate ratios, adjusting for age, sex, and reporting period. We analyzed 8,277 infections in elderly persons. Rates of campylobacteriosis, legionellosis, listeriosis, toxigenic *Escherichia coli* infections, and shigellosis were higher in community residents, and rates of *Salmonella* infection were higher in LTCF residents. Each year, 61.7 *Campylobacter* infections were reported per 100,000 LTCF residents, compared with 97.6 per 100,000 community residents. LTCF residents were at higher risk for *S. enterica* serotype Typhimurium associated with outbreaks. Rates of foodborne infections (except salmonellosis) were similar to or lower for LTCF residents than for community residents. These findings may indicate that food preparation practices in LTCFs are safer than those used by elderly persons in the community.

Infectious disease incidence varies with age, and elderly persons are considered more vulnerable than younger persons to foodborne and waterborne infections (1,2). In many countries, elderly persons unable to care for themselves live in long-term care facilities (LTCFs) where they receive assistance with meals, daily living, and health care (3). Food preparation practices and various exposures in LTCFs may modify the risk of foodborne and waterborne infections in these residents (4,5) when compared with elderly persons living in the community, who may have less safe food preparation practices (6–8).

A variety of pathogens transmitted by food or water, including *Campylobacter* sp., *Clostridium perfringens*, *Cryptosporidium* sp., *Legionella* spp., and *Shigella* sp., and various serotypes of *Salmonella enterica* can infect humans (9,10). Foodborne and waterborne infections predominantly manifest in elderly persons as gastroenteritis but, depending on the infectious agent, can result in pneumonia, bacteremia, and meningitis (11,12). Elderly persons can become infected by ingesting contaminated water or food or, as with *Legionella* spp., inhaling contaminated aerosols (13). Some infections are predominantly foodborne; others can be acquired from infected persons or animals or through contact with contaminated environments (4).

These agents can manifest as outbreaks in facilities, leading to community concern about the safety of residents (14,15). Although most outbreaks of gastroenteritis in LTCFs are spread from person to person and are generally mild (16), such outbreaks do result in higher case-fatality rates (CFRs) among residents (17). As a result, regulatory agencies in many countries have mandated programs to manage food safety in facilities. To prevent legionellosis in residents, health agencies commonly provide advice about disinfection of hot water systems that can be reservoirs for *Legionella* spp. (13).

Few studies have compared the incidence of infections caused by agents that can be transmitted by contaminated food or water consumed by elderly persons living in LTCFs and in the community. One study in the United States estimated that the lower limit of the death rate for...
nursing home residents from gastroenteritis of unknown etiology was 38.91 (95% CI 38.55–39.27) per 100,000 persons per year, compared with an estimated upper limit of 8.50 (95% CI 8.47–8.53) per 100,000 persons ≥65 years of age living in the community (18). Little examination has been done of the incidence of sporadic foodborne or waterborne diseases in institutionalized elderly persons. To address this gap, we estimated rates of reported infection in persons ≥65 years of age living in Victoria, Australia, infected with any of 7 different pathogens according to whether they lived in a government-subsidized LTCF or in the community, and we examined the effect of age on incidence of disease. These pathogens were *Campylobacter* sp., *Cryptosporidium* sp., *Legionella* spp., *Listeria* sp., *Salmonella enterica*, Shiga toxin–producing *Escherichia coli* (STEC), and *Shigella* sp.

## Methods

### Infectious Disease Surveillance

Victoria is 1 of 6 states and 2 territories in Australia. Public health legislation mandates that all physicians and pathology laboratories in Victoria report cases of notifiable conditions under the Public Health and Wellbeing Regulations 2009 (www.health.vic.gov.au/ideas/notifying/whatto) to the state’s Department of Health. Health department staff members enter details about reported cases into a database. Among the 64 conditions notifiable as of April 14, 2010, a total of 14 were enteric diseases that can be transmitted by contaminated food or water and 1 was *Legionella* infection that could potentially be transmitted by inhalation of contaminated water. Surveillance for these diseases has remained essentially unchanged in Victoria since the early 1990s, except for cryptosporidiosis, for which reporting was voluntary until 2001, when notification became mandatory by law.

We analyzed data on all cases of campylobacteriosis, cryptosporidiosis, legionellosis, listeriosis, shigellosis, salmonellosis, and STEC infection that were reported to Victoria’s Department of Health during January 1, 2000–December 31, 2009. Surveillance officers recorded whether cases were part of an outbreak or occurred in persons who had traveled overseas during their incubation period, which varied for different diseases. In addition, to identify information that was not recorded by surveillance officers, we reviewed surveillance data for all LTCF residents. More specifically, we identified where ≥2 cases of the same pathogen occurred within the same facility within 2 weeks, and we recoded these cases as outbreak associated. Surveillance officers also recorded whether case-patients had died of the disease or of other concurrent conditions within the weeks after infection during public health follow-up.

### Data Analysis

We categorized reported cases by residential status of the patient. An LTCF resident was a person who had a residential address of a government-subsidized LTCF, and a community resident was a person living in a private residence in Victoria (online Technical Appendix, wwwnc.cdc.gov/EID/pdfs/11-0311-Techapp.pdf). We excluded from analysis cases in persons without a valid address because we assumed the address was missing completely at random. We also excluded case-patients residing in privately funded facilities (supported residential services) catering to elderly or disabled persons or persons with psychiatric illness or dementia because they were not included in the denominator of LTCF residents. We counted the annual number of foodborne and waterborne infections, including those from epidemiologically important serotypes and species, in LTCF and community residents during the 10-year period. We obtained age-specific annual denominator data for residents of government-subsidized LTCFs from annual reports prepared by the Australian Institute of Health and Welfare (19). Denominator data for community residents were calculated by subtracting annual age-specific estimates of LTCF residents from estimated resident populations prepared by the Australian Bureau of Statistics (www.abs.gov.au/). We calculated annual rates of notification for different diseases by residential status and compared them with the total rate of notifications for the state. We calculated CFRs by dividing the number of deaths from the disease in the different groups by the total number of cases of disease, including cases for which death status was unknown, for the 10-year period. To account for age differences in LTCF and community residents, we calculated age-adjusted relative risks (RRs) for death from infection with different pathogens by using Mantel-Haenszel methods.

To estimate incidence rate ratios (IRRs), we used a negative binomial regression model of the annual count of foodborne and waterborne infections by the period of notification (2000–2004 and 2005–2009), sex, 3 categories of age (65–74, 75–84, and ≥85 years), and residence (LTCF and community). The number of persons living in LTCFs or the community in each age group for each year was entered into the model as an offset. We used robust variance estimation suited to longitudinal or clustered data to account for possible clustering from outbreaks. We assessed model fit by examining the distribution of standardized Pearson residuals. To assess the effect on incidence, we repeated regression models excluding travel-associated cases and including only the first case for each known outbreak in LTCFs and the community.

We analyzed data by using Stata version 11.2 (Stata Corp., College Station, TX, USA).
The Australian National University human research ethics committee approved this study. Victoria’s Department of Health approved release of the data.

**Results**

**Incidence in Persons >65 Years of Age**

During January 1, 2000–December 31, 2009, a total of 8,534 cases of the 7 diseases were reported in persons >65 years of age. During data cleaning, we excluded 238 (2.8%) persons without valid residential addresses and 19 who lived in a private institution caring for elderly or disabled persons. A total of 8,277 cases were available for analysis, including 132 (1.6%) case-patients living in retirement villages.

**Infections in Institutional and Community Residents**

Rates of all reported infections in LTCF residents were similar to or lower than those in community residents, except infections from *S. enterica* (Table 1). No LTCF residents were reported with *L. longbeachae*, STEC, or *Shigella* sp. infections during the surveillance period. Among persons >65 years of age infected with *Legionella* species other than *L. longbeachae* were more likely to live in an LTCF (crude RR 1.15, 95% CI 1.1–1.2; *p* = 0.27). The reported rate of *Campylobacter* spp. was lower in LTCF residents than in community residents for all years of surveillance (Figure 1). In contrast, reports of *S. enterica* serotype Typhimurium peaked because of outbreaks during the study period, which resulted in higher rates for LTCF residents overall (Figure 2).

**Associated Death**

The CFR was highest for listeriosis and *L. longbeachae* infection, from which 17.8% and 6.7% of persons died, respectively (Table 2). The age-adjusted RR for death from *L. monocytogenes* infections was 3.5 (95% CI 1.2–10.4; *p* = 0.03) for LTCF residents. No *L. pneumophila*-associated deaths were recorded in LTCF residents. For salmonellosis, most deaths were considered to have resulted from other concurrent conditions. The age-adjusted RR for death in LTCF residents infected with any *S. enterica* serotype did not differ significantly from that in community residents (adjusted RR 2.8, 95% CI 0.8–9.1; *p* = 0.87). Death status was not routinely ascertained for persons with campylobacteriosis, although 1 death from campylobacteriosis and 11 deaths from concurrent conditions were recorded. No deaths were recorded for shigellosis or cryptosporidiosis.

**Rates of Foodborne and Waterborne Infections**

In multivariable analysis, the incidence of *S. enterica* serotype Typhimurium was higher in LTCF residents than

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**Table 1. Incidence rate for reported infections with pathogens possibly transmitted by food or water, Victoria, Australia, January 2000–December 2009**

<table>
<thead>
<tr>
<th>Pathogen</th>
<th>Persons &lt;65 y</th>
<th>Persons &gt;65 y</th>
<th>Total reports</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No. cases</td>
<td>Rate</td>
<td>LTCF residents</td>
</tr>
<tr>
<td>Campylobacter spp.</td>
<td>50,444</td>
<td>115.4</td>
<td>215</td>
</tr>
<tr>
<td>Cryptosporidium sp.</td>
<td>4,955</td>
<td>11.3</td>
<td>7</td>
</tr>
<tr>
<td>Legionella pneumophila/other</td>
<td>457</td>
<td>1.0</td>
<td>8</td>
</tr>
<tr>
<td><em>L. longbeachae</em></td>
<td>49</td>
<td>0.1</td>
<td>0</td>
</tr>
<tr>
<td>Listeria monocytogenes†</td>
<td>46</td>
<td>0.1</td>
<td>3</td>
</tr>
<tr>
<td>Salmonella enterica</td>
<td>7,204</td>
<td>16.5</td>
<td>87</td>
</tr>
<tr>
<td>serotype Typhimurium</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>S. enterica</em>, other</td>
<td>5,003</td>
<td>11.4</td>
<td>44</td>
</tr>
<tr>
<td>serotypes</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Shigella</em> sp.</td>
<td>845</td>
<td>1.9</td>
<td>0</td>
</tr>
</tbody>
</table>

*Annual rate of reported infections per 100,000 persons. LTCF, long-term care facility.
†Total listeriosis reports exclude 16 pregnancy-associated infections.
in community residents (IRR 2.3, 95% CI 1.6–3.4; p<0.001) and non-Typhimurium serotypes of *S. enterica* (IRR 1.4, 95% CI 1.0–1.9; p = 0.04) and lower for *Campylobacter* spp. (IRR 0.63, 95% CI 0.52–0.77; p<0.001) (Table 3). IRRs could not be estimated for *L. longbeachae* and STEC infections and for shigellosis because no cases occurred in LTCF residents during the surveillance period. We observed a trend of increasing rates of reported infections for cryptosporidiosis, salmonellosis, and campylobacteriosis over time during the surveillance period.

From multivariable analysis, reported incidence rates were higher in older age groups for *L. monocytogenes*, *Campylobacter* sp., and *S. enterica* serotype Typhimurium infections than in the base age group of persons 65–74 years (Table 3). The incidence of *L. pneumophila* and non-Typhimurium serotypes of *S. enterica* infections and cryptosporidiosis did not differ significantly by age group.

**Accounting for Travel and Outbreaks**

During their incubation period, 105 (1.3%) of the 8,277 persons with notifiable infections traveled internationally; all were community residents. Among community residents, 16 (37.2%) of the 43 with shigellosis and 48 (8.7%) of the 552 with non-Typhimurium serotypes of *S. enterica* had traveled internationally, compared with 0 of those with *L. monocytogenes*, *L. longbeachae*, and STEC infections. Only 35 (0.6%) of the 6,207 community residents infected with *Campylobacter* sp. were recorded as traveling overseas before infection.

During the study period, 42 separate outbreaks of *S. enterica* serotype Typhimurium occurred in persons ≥65 years of age. There were also 36 outbreaks of non-Typhimurium serotypes of *S. enterica*, 14 of *L. pneumophila*/other, 10 of *Campylobacter* spp. infections, and 2 of *Shigella* sp. In total, 189 (2.4%) of 7,913 cases in community residents were recorded as outbreak associated. In LTCF residents, 111 (30.5%) of the 364 cases were outbreak associated, including 68 (78.2%) of the 87 *S. enterica* serotype Typhimurium and 33 (15.3%) of the 215 *Campylobacter* spp. infections. No cases of cryptosporidiosis, listeriosis, or STEC infection were recorded as outbreak associated in either LTCF residents or community residents.

When we repeated multivariable models, excluding travel-associated infections and including a single case for each identified outbreak, the incidence rate for *S. enterica* serotype Typhimurium was similar in LTCF residents and community residents (IRR 0.91, 95% CI 0.64–1.29; p = 0.59). For infections with non-Typhimurium serotypes of *S. enterica*, the incidence was higher in LTCF residents than in community residents (IRR 1.4, 95% CI 1.0–2.0; p = 0.05). After adjustment for travel and outbreaks, the incidence rate was lower for LTCF residents with *Campylobacter* spp. (IRR 0.57, 95% CI 0.48–0.68; p<0.001). For *L. pneumophila*, the incidence rate was lower, but not significantly, for LTCF residents (IRR 0.63, 95% CI 0.29–1.3; p = 0.23).

**Discussion**

Rates of foodborne and waterborne infections among LTCF residents were lower than or similar to rates among community residents, except for salmonellosis, which was higher. In particular, rates of campylobacteriosis in LTCF residents were consistently lower throughout the entire study period, which was unexpected because incidence of this infection is universally high (20). Despite the high

<table>
<thead>
<tr>
<th>Pathogen</th>
<th>Died of disease</th>
<th>Died of concurrent condition</th>
<th>Death status unknown</th>
<th>No. cases</th>
<th>Case-fatality rate*</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Legionella pneumophila</em>/other</td>
<td>11</td>
<td>1</td>
<td>169</td>
<td>301</td>
<td>4.0</td>
</tr>
<tr>
<td><em>L. longbeachae</em></td>
<td>2</td>
<td>1</td>
<td>0</td>
<td>45</td>
<td>6.7</td>
</tr>
<tr>
<td><em>Listeria monocytogenes</em></td>
<td>4</td>
<td>9</td>
<td>14</td>
<td>73</td>
<td>17.8</td>
</tr>
<tr>
<td><em>Salmonella enterica</em> serotype Typhimurium</td>
<td>3</td>
<td>8</td>
<td>183</td>
<td>672</td>
<td>1.6</td>
</tr>
<tr>
<td><em>S. enterica</em>, other serotypes</td>
<td>0</td>
<td>7</td>
<td>177</td>
<td>596</td>
<td>1.2</td>
</tr>
<tr>
<td>Shigella toxin–producing <em>Escherichia coli</em></td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>12</td>
<td>8.3</td>
</tr>
<tr>
<td><em>Shigella</em> sp.</td>
<td>0</td>
<td>0</td>
<td>10</td>
<td>43</td>
<td>NA</td>
</tr>
</tbody>
</table>

*Per 100 cases. Case-fatality rate for the 10-year study period was calculated by dividing the total number of deaths among case-patients by all case-patients, including those with unknown death status. NA, not applicable.
incidence of campylobacteriosis, outbreaks are rare in Australia, possibly because of the high dose required to cause infection and because foods causing infection are thought to become contaminated through cross-contamination (16,20). In Australia, Campylobacter infections are the most common cause of bacterial foodborne disease, with contaminated chicken meat causing \( \approx 30\% \) of all infections each year (21–23).

The lower incidence of campylobacteriosis might result from the highly regulated food hygiene system for LTCFs. In 1998, Victoria was the first Australian state to implement mandatory food safety programs for food service settings, and those serving vulnerable populations require independent auditing (24). These programs might have resulted in better understanding and practices by LTCF staff about food storage, cooking, and cross-contamination than by elderly persons in their own homes. Campylobacter infections in elderly community residents have been associated with risk possibly from cross-contamination during food preparation (25). Even though Campylobacter infections are associated with travel, a case–control study in Australia found that only 23 (2.8\%) of 833 persons \( \geq \)5 years of age with campylobacteriosis had traveled overseas during the week before illness (21). When we accounted for known travel history and outbreak-associated cases, the IRR for Campylobacter spp. infections in LTCF residents was lowered.

S. enterica is a common cause of foodborne and waterborne outbreaks in LTCFs (17,26,27), a finding that our study confirmed. We found that outbreaks of S. enterica serotype Typhimurium infections accounted for the higher incidence of these infections, but not for non-Typhimurium serotype infections, in LTCF residents. Sources of outbreaks in LTCFs often are not identified, although eggs are commonly suspected as the cause in S. enterica serotype Typhimurium–associated outbreaks (17). Although residents are at higher risk for outbreak-associated disease, ascertainment of cases is biased in the institutional setting because of the common living environment, centralized access to health care, and collection of specimens by public health staff. During outbreaks, public health investigators often collect fecal specimens from LTCF residents with diarrhea, which would not occur for elderly persons in the community. Because surveillance is well established in Victoria, LTCFs are more likely than persons in the community to report outbreaks (16).

We did not find any evidence to suggest that living in an LTCF increased a person’s risk for legionellosis, despite the occasional occurrence of outbreaks and sporadic cases in this setting (13,28). LTCF residents reported with legionellosis were more likely to be infected with Legionella species other than with L. longbeachae. L. longbeachae is associated with gardening and potting mix, so we expected the incidence of this infection to be low in LTCF residents (29). The incidence of listeriosis was similar in LTCF and community residents. Given the food safety program requirements in facilities in Victoria, LTCF residents plausibly could be exposed to lower concentrations of L. monocytogenes in food, compared with community residents who may keep food longer, have poorer food preparation practices, and eat foods considered higher risk for transmitting foodborne pathogens (8).

Different clinical investigative approaches for LTCF and community residents with potential foodborne and waterborne disease might account for some of our findings. Although clinicians might elect not to collect specimens when LTCF residents have diarrheal illness, we think it more likely that reporting is more complete in LTCFs. Most of the diseases in our study are serious illnesses, and infected persons would have severe gastrointestinal and extraintestinal symptoms lasting for several days or weeks (3,4). In a case–control study of campylobacteriosis...
in Australia, 41% of case-patients had bloody diarrhea, and 75% had fever; both of these symptoms are strong predictors for physicians ordering laboratory tests (30,31).

We were unable to control for potential confounding factors, such as concurrent conditions and factors that might predispose for infection. Many elderly persons with concurrent conditions live in the community, but the health status of LTCF residents is likely to be lower, and they are likely to be more frail. We would have partly controlled for frailty through inclusion of age in our multivariable model because elderly persons in institutions are the oldest and the most frail in society (32). In addition, our study was underpowered to detect an effect for diseases where notification rates were very low. The potential bias in the final estimates from lack of control of confounding would be more likely to result in increased incidence rates in LTCF residents. However, except for salmonellosis, infection rates were higher in community residents.

Our findings should not be overinterpreted because our study was a retrospective record–based study in which we manually coded surveillance data and were unable to validate case-patients’ addresses. It is possible that we were unable to correctly identify residential status of case patients from addresses. In some instances, residents were recorded as living at addresses where an LTCF and retirement village were on the same grounds, making determining whether a person lived in the facility difficult. Similarly, some persons might have been infected after moving into an LTCF, but the address on a pathology report still recorded their residential address in the community where they had previously lived. However, in Victoria, physicians and laboratories were required to report these infections, making it unlikely that both sources of notification would incorrectly report the residential address. For Campylobacter infections, however, physicians report only 50% of notifications, with the remainder coming from laboratories (33).

Elderly community residents might receive meals from organizations that provide community support. In addition, elderly residents of LTCF might eat food that has been prepared outside the facility during excursions or brought in by visitors, which could result in exposure to foodborne pathogens. For both groups, these alternative routes of exposure would modify the risk for infection so that it did not truly reflect the risk in their place of residence.

The strength of our approach was that we consistently coded addresses without regard to disease-causing agent, yet we observed distinct differences in reported incidence from disease to disease. Our findings were consistent with what we know about these diseases, such as increasing incidence in older persons for diseases such as listeriosis. The CFRs were consistent with reports in the literature for elderly persons, although we assessed deaths only short term (i.e., in the weeks after infection) (34,35). In general, elderly persons have more severe outcomes from foodborne infections than do younger persons (4,18). Large-scale studies that used population-based registers have demonstrated that enteric diseases contribute to more deaths than recognized from short-term follow-up, even when controlling for concurrent conditions (36,37).

In our study, rates of surveillance reports for most infections in persons ≥65 years of age were similar to or lower than for persons <65 years of age, a finding that contradicts the common statement that elderly persons are at higher risk for foodborne disease. However, we did find that the CFRs were high for some infections and that LTCF residents were affected more severely. We believe that our findings can be generalized to other Australian states and territories with similar rates of infection and methods of surveillance (16,22). Other investigators could repeat this study by using record-linkage to compare their findings with our findings.

We observed a lower incidence of reported Campylobacter spp. infection in LTCF residents, which provides some reassurance for food safety regulators and the aged care industry. Our study highlights that most foodborne and waterborne infections are rare in elderly residents of LTCF and the community, but that these infections do cause occasional deaths. Primary research is needed into the specific causes of foodborne and waterborne infections in elderly persons in the community and in institutional settings that particularly accounts for the effect of concurrent conditions. In our study, elderly LTCF residents had an incidence of foodborne and waterborne infections that was similar to or lower than that that for elderly persons living in the community, except for S. enterica infections.

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Use of trade names is for identification only and does not imply endorsement by the Public Health Service or by the US Department of Health and Human Services.
Foodborne and Waterborne Infections in Elderly Community and Long-Term Care Facility Residents, Victoria, Australia

Technical Appendix

Coding Surveillance Data for Residential Status

Surveillance in Victoria, Australia

In Victoria, Australia, doctors and laboratories were required to notify cases of 64 different infections of public health concern under the Public Health and Wellbeing Regulations 2009 (www.health.vic.gov.au/ideas/notifying/whatto) to the Department of Health, Victoria. Doctors reported cases on handwritten forms or entered details about infected patients online (see www.health.vic.gov.au/notifying). Information provided by doctors included: case-patient’s name, date of birth, sex, ethnicity, residential address, infecting agent, date illness began, clinical comments, and contact details for the notifying doctor. Treating doctors were required to sign the notification form. Doctors were able to send, fax, or enter details online or report cases over the phone. Pathology laboratories provided copies of reports to treating doctors about patients infected with diseases on the notification list to the Department of Health. These reports included the patient’s name, date of birth, sex, residential address, infecting agent, date of specimen collection, and date a test result was positive.

Department of Health staff members entered details about notified cases onto a database—the Notifiable Infectious Diseases Surveillance System. Where there were discrepancies on duplicate reports from doctors and laboratories, information from the treating doctor’s report were entered onto the database. Infections were prioritized for follow-up by public health officers, with more serious or transmissible infections classified as urgent. On the basis of this, public health staff entered additional information on the surveillance database, such as exposure information, vaccination status, any treatment or prophylaxis, and whether the case was part of an outbreak.
Recording of Addresses

The surveillance dataset recorded address details for cases across 9 fields, that were, in order:

1. Address prefix—for recording information before street number, such as “unit 10.”
2. Street number—recording the street number of the address.
3. Address suffix—records additional information to the street number.
4. Street name—street where the case-patient lived.
5. Address 2—additional information about the address.
6. Address 3—additional information about the address.
7. Suburb—city, town, or suburb where the case-patient lived.
8. State—records the case-patients’ residence by state or territory and whether the case-patient was an overseas resident.
9. Postcode—postal code corresponding to the address.

Assessing Exposure

To assess exposure, 1 investigator (M.D.K.) assessed each address field for all records for names matching that of government-subsidized LTCFs. Names of government-subsidized facilities were obtained from a list held by the Victorian Department of Health in 2010. Where there was doubt about whether a facility named in a case-patient’s record address was an LTCF, the name was checked using a publicly available online aged care guide (www.agedcareguide.com.au/) and the name searched using Google and location searched on Google maps. If any of the following rules were satisfied, a case-patient was coded as living in an LTCF:

1. If the name of a nursing home was included in any of the first 6 address fields and the suburb was recorded as having an LTCF of the same name on the LTCF list or online aged care guide or
2. If the case-patient’s address matched exactly, in terms of street number and street name, that of a recognized government-subsidized LTCF or
3. If the case-patient’s address did not specify a street number but listed a street name where the LTCF list and Internet aged care guide included a facility at that street without a street number recorded or
4. If a Web search on the facility identified that there had been an LTCF of the same name and address at the time of the case-patient’s infection but was not currently in operation in 2010.

If a typical name of facilities of any type was mentioned in the first 6 fields and the name was not identified in the LTCF facility lists, a Google search and Google map search were conducted to identify whether the name referred to a retirement village or Supported Residential Service. In instances where the case-patient’s address mentioned a “retirement village” but was the same as a recognized LTCF, we searched Google to determine whether there was a retirement village onsite at the same address. If so, the case-patient was coded as living at a “retirement village.” All case-patients living in “retirement villages” were considered “community residents” for the purposes of this study.

Exclusions

In this study, case-patients were excluded if they were not part of the study population according to the following criteria.

1. Nonresidents of Victoria or Australia according to the residential address or country of residence were excluded from the study.

2. Case-patients for which information in any of the first 6 address fields was insufficient to assess whether they lived in an LTCF or the community were not included. Case-patients with a residential address listed as “RMB” (Rural Mail Box) followed by a number were coded as having a valid address. RMB indicated that a person received mail in a rural area at a designated mail box where defined street numbers were not in use. Case-patients with a residential address of PO Box (Post Office Box) followed by a number were coded as missing a valid residential address because it gave no indication of where a case-patient lived.

3. Resident of a “supported residential service”—privately run facilities providing care for people who were old, disabled, or suffering from dementia or psychiatric illness (see www.health.vic.gov.au/srs/index.htm) were not included in the study.