Bartonella spp. in Rats and Zoonoses, Los Angeles, California, USA

Vijay A.K.B. Gundi, Sarah A. Billeter, Michael P. Rood, and Michael Y. Kosoy

Bartonella spp. were detected in rats (Rattus norvegicus) trapped in downtown Los Angeles, California, USA. Of 200 rats tested, putative human pathogens, B. rochalimae and B. tribocorum were found in 37 (18.5%) and 115 (57.5%) rats, respectively. These bacteria among rodents in a densely populated urban area are a public health concern.

Bartonella spp. are vector-borne bacteria associated with an increasing array of emerging zoonotic infections in humans and animals (1). Some Bartonella spp. are widely distributed among small mammals in the United States and potentially cause human health concerns because these bacteria may be associated with human diseases (2). However, limited surveys have been conducted to identify infectious agents involved in zoonotic infections in rodents within urban areas of the United States (3). Norway rats (Rattus norvegicus) have been shown to harbor several Bartonella species, including B. tribocorum, B. elizabethae, B. ratti massiliensis, B. phoceensis, B. queenslandensis, and a strain closely related to B. rochalimae (3–6), of which B. elizabethae, B. rochalimae, and B. tribocorum were implicated in human diseases (7–11).

Our study had 3 purposes. The first purpose was to investigate prevalence of Bartonella spp. infections in rodent populations in downtown Los Angeles, California, USA. The second purpose was to evaluate genetic diversity of Bartonella spp. in blood of urban rats by analyzing variations of the citrate synthase (gltA) gene. The third purpose was to compare rates of detection of Bartonella spp. infections in rats between culture and molecular assays.

Author affiliations: Centers for Disease Control and Prevention, Fort Collins, Colorado, USA (V.A.K.B. Gundi, S.A. Billeter, M.Y. Kosoy); and Los Angeles County Department of Public Health, Baldwin Park, California, USA (M.P. Rood)

DOI: http://dx.doi.org/10.3201/eid1804.110816

The Study

Rats were sampled during 2003–2007 at 16 sites in downtown Los Angeles in a rodent management/disease surveillance program. The rats were captured by using Tomahawk live traps (Tomahawk Live Trap Co., Hazelhurst, WI, USA) and anesthetized with CO₂. Blood samples were obtained by cardiac puncture, transferred to sterile cryovials, placed on dry ice, and preserved at –70°C until testing at the Bartonella Laboratory of the Centers for Disease Control and Prevention (Fort Collins, CO, USA).

Genomic DNA was extracted from animal blood according to the blood protocol of the QIAamp DNA Mini Kit (QIAGEN, Valencia, CA, USA). Primers CS140f and CS443f (12) and CS781f and CS1137n (13), including newly designed primers CS120f-5′-TTTCACTTATGATCCTGGCTT-3′and CS1210r-5′-GATCTYTCATTCTTCTTTCCA-3′, which amplify DNA fragments ranging from 380 to 1,091 bp encompassing a 327-bp-specific zone (between bp positions 801 and 1127) of the gltA gene (14), were used in different combinations.

Eight PCRs were performed for each sample. PCR products were purified by using the Qiaquick PCR Purification Kit (QIAGEN, Germantown, MD, USA). The same primers were used for DNA sequencing. The obtained DNA sequences were trimmed to 327 bp between positions 801 and 1127 because most Bartonella spp. gltA sequences available in GenBank are limited to this size.

Details of procedures used for isolation of Bartonella spp. from mammalian blood have been described (2). Rat blood was diluted 1:4 in brain–heart infusion medium containing 5% fungizone and pipetted onto heart infusion agar plates containing 10% rabbit blood. Plates were incubated aerobically at 35°C in an atmosphere of 5% CO₂ for ≤4 weeks and monitored for bacterial growth at least 1× per week after initial plating. Colonies were subpassaged onto fresh agar plates until a pure culture, free from contamination, was obtained.

Analysis of DNA sequences and phylogenetic relationships was performed by using MEGA4 (www.megasoftware.net/) and the neighbor-joining method with the Kimura 2-parameter distance model. Stability of inferred phylogeny was assessed by using bootstrap analysis of 1,000 randomly generated trees. DNA sequences obtained in this study were deposited in GenBank under accession nos. JF429450–JF429625.

A total of 200 R. norvegicus rats were trapped in 16 sites in downtown Los Angeles. Bartonella DNA was detected in 135 (67.5%) of 200 rat blood samples. PCR-positive blood samples were subsequently cultured for viable Bartonella organisms. Fifty-nine (43.7%) of 135 blood samples were confirmed as bacteremic for bartonellae by culturing.

A total of 176 sequences were obtained either directly from blood samples (117 sequences) or from pure cultures
Among 176 gltA sequences, 23 genotypes with ≥1 nt difference were found, and sequence similarity between genotypes ranged from 85.3% to 99.7%. These 23 genotypes were closely related to B. tribocorum (n = 16), B. rochalimae (n = 2), B. queenslandensis (n = 1), or 4 potentially novel genotypes. A total of 130 DNA sequences (JF429450–JF429579) obtained from 115 rats were grouped into 16 genotypes (1–16). These genotypes showed 98.2%–99.7% sequence similarity with each other and were genetically related to B. tribocorum IB506 (AJ005494) (97.9%–100% identity). All 16 genotypes clustered with B. tribocorum with a high bootstrap value, as shown in the phylogenetic tree (online Appendix Figure). The B. queenslandensis group consisted of a single genotype (genotype 17) from 4 sequences (JF429580–JF429583) from 4 rats. This genotype demonstrated a sequence similarity to B. queenslandensis AUST/NH12 (EU111800) of 99.0% for the gltA gene (online Appendix Figure).

Conclusions

In this study, 3 novel Bartonella genogroups identified in 4 R. norvegicus rats were not genetically related to any known Bartonella spp. and might represent novel Bartonella spp. Genotypes 18 (JF429586) and 19 (JF429587) formed a Bartonella genogroup with 95% similarity to B. tribocorum as the closest species. Likewise, genotypes 20 (JF429584) and 21 (JF429585) have 91.4% and 92.6% sequence identities, respectively, with B. claridgeiae, which is their closest related species (online Appendix Figure).

Of 135 PCR-positive blood samples, 59 yielded cultures of 3 Bartonella spp.: B. tribocorum (n = 54), B. rochalimae (n = 4), and B. queenslandensis (n = 1). The number of CFU per 100 μL of blood varied: 800–1,400 CFU for B. tribocorum, 160–240 CFU for B. rochalimae, and 100 CFU for B. queenslandensis. Although 4 rats were positive by PCR for B. queenslandensis, this organism was cultured from only 1 of these animals.

Our study reports detection and identification of Bartonella spp. in urban rats in downtown Los Angeles, California. We demonstrated that R. norvegicus in downtown Los Angeles can serve as reservoirs of several Bartonella spp., such as B. rochalimae, B. tribococorum, B. queenslandensis, and possibly 3 additional novel species. Some genotypes identified in rats showed a high level of similarity (≥98.8%) with a B. tribocorum isolate obtained from a febrile patient in Thailand (GenBank accession no. GQ225706) (11). Another species (B. rochalimae) was isolated from a patient with splenomegaly who had traveled to South America (14). This bacterium has also been isolated from dogs, foxes, rats, shrews, gerbils, and raccoons, suggesting that multiple reservoirs may be involved in maintenance of this species. One Bartonella genotype found in 4 rats in this study was 99.0% similar to B. queenslandensis (GenBank accession no. EU111800), which was originally isolated from R. fuscipes rats in Australia (15).

Because most identified Bartonella species have been reported as human infectious agents elsewhere, the finding that they were circulating among rodents in a densely populated urban area is of serious public health concern. In this context, further studies should be conducted on a larger collection of rodents and clinical human samples to determine evolutionary, genetic, and pathogenic relationships.

Acknowledgment

We thank Robert Flores for assistance with rat trapping and processing.

This study was supported in part by an appointment to the Emerging Infectious Diseases Fellowship Program administered by the Association of Public Health Laboratories and funded by the Centers for Disease and Control and Prevention.

Dr Gundi is an Emerging Infectious Diseases Fellow at the Bartonella Laboratory, Division of Vector-Borne Diseases, Centers for Disease Control and Prevention, Fort Collins, Colorado. His research interests include tropical and emerging infectious diseases.

References


Address for correspondence: Michael Y. Kosoy, Centers for Disease Control and Prevention, 3150 Rampart Rd, Fort Collins, CO 80521, USA; email: mck3@cdc.gov