Francisella tularensis, a Tier 1 select agent, is one of the most pathogenic bacteria known and the etiologic agent of the zoonotic disease tularemia (1,2). Although the various subspecies of F. tularensis share considerable genomic content (>97% nt identity), they exhibit different degrees of virulence (3). F. tularensis subspecies tularensis (also known as type A) and subspecies holarctica (also known as type B) infections can be life-threatening if untreated (1). Type A is further subdivided into subtypes A.I and A.II; clade A.I contains strains that are associated with considerably higher death rates among humans than are the other members of this pathogenic species (4). Type B infections occur throughout the Northern Hemisphere, whereas type A infections occur primarily in North America (3).

Fatal and nonfatal cases of tularemia in domestic cats (Felis catus) have been reported, as has the transmission of this disease from cats to humans (5–7). Previous serologic surveys from several geographic regions determined that 12% of the domestic cats examined had antibodies to F. tularensis (8). Cases of feline-associated tularemia in humans continue to appear in the literature. However, the relative contribution of this source of F. tularensis transmission to humans is unknown and may be underrecognized. This study was conducted to examine the relative proportion and characteristics of feline-associated F. tularensis isolates within the repository at the University of Nebraska Medical Center (UNMC).

The Study

A review was conducted of the 106 wild-type F. tularensis isolates voluntarily deposited in the UNMC collection during 1998–2012. These isolates were obtained from infected humans, animals, or ticks predominantly residing in Nebraska; however, isolates from several other regions were also included. Wild-type F. tularensis isolates had been transferred to UNMC from other locations, according to requirements of the national Select Agent Program (2). None of these isolates had been solicited from veterinary or environmental reference laboratories. Species identity for locally detected wild-type F. tularensis isolates to which humans had been exposed was confirmed by the Nebraska Public Health Laboratory in Omaha. Visible culture material was manipulated by authorized persons following select agent–approved Biosafety Level 3 criteria. For genotyping, we used the PCR-based differential insertion sequence amplification (DISA) method with the CR10 C+L+S primer set and pulsed-field gel electrophoresis (PFGE) of Pmel-digested Francisella spp. DNA, as previously described (9).

Of the 106 wild-type F. tularensis isolates in the repository, 54 (51%) were from humans for whom the source of exposure was unknown or undocumented, 29 (27%) were from cats (21) or from humans (8) with tularemia linked to infected cats, 5 (5%) were from humans with tularemia linked to infected ticks, 1 (1%) was from a human with tularemia linked to an infected rabbit, 16 (15%) were from animals with tularemia, and 1 (1%) was from an unknown host. Of the 29 isolates derived from feline-associated cases, 28 were associated with domestic cats and 1 involved a feral cat (Figure). Eight cases of human tularemia occurred through a cat bite: 7 of these cases involved adults and 1 involved a 6-year-old child. In 2 cases, the person was bitten while taking a rabbit from a cat. None of the 8 humans died.

We also analyzed a subset of the 29 F. tularensis isolates obtained from cats or humans bitten by cats. All but 2 of the 29 isolates were available for genotyping. DISA and PFGE results showed that 1 type B and 26 subtype A.I strains were responsible for the 27 cases of tularemia (Figure). Of the 8 cat bite–associated human tularemia cases, 1 was caused by a type B and 7 by subtype A.I strains. None of the feline-associated tularemia cases were caused by a subtype A.II strain.

PFGE demonstrated that the feline-derived F. tularensis A.I strains could be further divided into 4 subpopulations. For strains in 1 of these subpopulations, DISA results showed chromosomal polymorphisms in which a
1.2-kbp or a 1.5-kbp amplicon was produced. This finding is consistent with 2 unique origins for the strains, even though they were obtained from the same geographic area.

Subset analysis of Nebraska cases showed that 48% (24/50) of the wild-type \textit{F. tularensis} isolates were feline-associated, and most of those 24 isolates (71%, 17/24) were associated with tularemia cases that occurred in 1 city in eastern Nebraska. The highest number of tularemia cases in Nebraska was reported in 2012, a year noted for extreme heat and drought conditions with warm weather beginning in early January and lasting through December. No feline-associated tularemia cases in Nebraska were reported during 1998–2004; however, 18 were reported during 2005–2011.

**Conclusions**

Two different molecular methods demonstrated that \textit{F. tularensis} subtype A.I was responsible for most of the tularemia cases in cats (96%, 26/27); 1 of the 27 cases was caused by a type B strain. This finding is consistent with subtype A.I strains being found predominantly east of the 100th meridian in the United States, whereas A.II strains appear to be restricted to the west of this meridian.

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**Figure.** Molecular genotyping of \textit{Francisella tularensis} isolates obtained from infected cats or humans bitten by an infected cat, United States, 1998–2012. Genotyping was performed by using pulsed-field gel electrophoresis (PFGE) and the PCR-based differential insertion sequence amplification (DISA) assay. A dendrogram of PFGE patterns obtained with \textit{Pmel}-digested \textit{F. tularensis} isolates is shown on the left; the scale bar at the top indicates distance in relative units. The genotype-specific amplicon lengths obtained with the DISA CR10 C+L+S primer set are shown on the right. Migration profiles of the \textit{Pmel} restriction fragments from \textit{F. tularensis} chromosomal DNA were normalized to \textit{Smal}-digested \textit{Staphylococcus aureus} NCTC 8325 by using BioNumerics software (Applied Maths, Inc., Austin, TX, USA). Cluster analysis was performed by using the Dice correlation coefficient and the unweighted pair group mathematical average (UPGMA) clustering algorithm in the BioNumerics software. The DISA CR10 C+S primer pair identified the subtype A.I strains, whereas the CR10 C+L primer pair differentiated the subtype A.II and type B strains by the size of the amplicon produced. Underlining indicates \textit{F. tularensis} strains SCHU S4 (subtype A.I), WY96–3418 (subtype A.II), and ATCC 29684 (type B), which were included as references. \textit{F. tularensis} isolates NE-062508 and NE-073009 were no longer viable after storage and therefore not genotyped; these pathogens were isolated from domestic cats from eastern Nebraska (NE) in 2008 and 2009, respectively. ATCC, American Type Culture Collection; NC, North Carolina; OH, Ohio; OK, Oklahoma; WY, Wyoming.
The high percentage of feline-associated tularemia cases reported in this study demonstrates that outdoor domestic cats are frequent vectors of *F. tularensis*, particularly in areas where the bacterium is endemic. The cases reported in this study were primarily from urban areas with nearby access to veterinarians and with cat owners/caretakers motivated to treat infected animals; therefore, our findings may not accurately estimate the prevalence of this pathogen within the feral and rural domestic cat populations. These findings do emphasize the importance of taking appropriate measures to reduce the transmission of feline-associated tularemia to humans and the need for a vaccine that protects against the highly virulent pathogen *F. tularensis*.

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Dr Larson is an assistant professor in the Pathology and Microbiology Department at the University of Nebraska Medical Center in Omaha. Her research interests include the development of diagnostics and therapeutics for pathogen identification and treatment, respectively, as well as determining the molecular mechanisms that allow pathogens to replicate and persist.

**References**


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