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Reemergence of *Brucella melitensis* in Wildlife, France

To the Editor: Brucellosis is a worldwide zoonosis caused by *Brucella* spp. France has been free of bovine, ovine, and caprine brucellosis (caused by *B. abortus* or *B. melitensis*) since 2003 (1). In early 2012, an outbreak of bovine and human brucellosis caused by *B. melitensis* biovar 3 (*Bme13*) occurred in a French Alp massif (mountainous region), where the last reported outbreak occurred in 1999 (online Technical Appendix Figure, <http://wwwnc.cdc.gov/EID/article/20/9/13-1517-Techapp1.pdf>) (2). This outbreak suggested the

persistence or reemergence of *Brucella* spp. in livestock.

An extensive investigation was conducted that involved 40 animal herds with direct links to the outbreak. Six months later, blood samples from each adult animal in any herd (12,116 animals in 205 herds) that grazed during the summer of 2012 in the massif underwent serologic analysis. However, no other case was identified in this population (online Technical Appendix Table 1). Therefore, a potential wildlife source was investigated.

Wild ruminants in the study area were the following species: hunted red deer (*Cervus elaphus*), roe deer (*Capreolus capreolus*), chamois (*Rupicapra rupicapra*), and protected Alpine ibex (*Capra ibex*). Although *B. abortus* and *B. suis* have been reported in numerous wildlife species (3), *B. melitensis* has rarely been isolated from wildlife, and only sporadic cases of infection have been reported in Europe, in chamois and Alpine ibex in the Alps (4,5) and in Iberian ibex (*Capra pyrenaica hispanica*) in the Pyrenees (6). These cases were considered to be caused by spillover from domestic ruminants, which suggests that these wild species are unable to sustain the infection (3).

We conducted our investigation during the fall–winter of 2012–2013 in the entire massif where the outbreak occurred. Blood, lung, spleen, and testes or uterus samples were obtained from all hunted animals. French Authorities authorized the killing of 12 seropositive or diseased Alpine ibex with clinical signs of brucellosis (i.e., arthritis or orchitis) among 30 captured animals.

All serum samples were tested according to requirements of the World Organisation for Animal Health for diagnosis of brucellosis in small ruminants by using by the Rose Bengal test (RBT) and the complement fixation test (CFT) (7), and by indirect ELISA (IDEXX, Montpellier, France) and competitive ELISA

(cELISA; Ingenasa, Madrid, Spain). When blood samples were unsuitable for RBT or CFT or were missing, a lung extract was tested by only the 2 ELISAs. Culture was only performed on samples from seropositive animals (online Technical Appendix Table 1) (8). If bacteriologic results were negative, a *Brucella* genus–specific real-time PCR was also used (9).

A total of 129 hunted ruminants (55 chamois, 30 red deer, 44 roe deer) were tested. No clinical signs were observed, except for arthritis in the knee of 1 chamois. All ruminants were seronegative except for the chamois, which showed positive results in the RBT, CFT, and cELISA, and 1 red deer, which showed a weakly positive result in the cELISA, but negative results by culture and real-time PCR. *Bme13* was isolated from the chamois (online Appendix Table 1).

Among 289 Alpine ibex observed in the massif, 24 were killed (22 randomly sampled animals that showed 2 diagnostic lesions at necropsy [arthritis in the knee and mammary abscesses] and 2 diseased animals [arthritis in the knee and orchitis]), and samples from these animals were subjected to serologic analysis. Ten Alpine ibex (including the 2 diseased animals) showed positive results in the RBT, CFT, and both ELISAs, and 2 showed positive results only for both ELISAs. Thus, the prevalence of *B. melitensis* in randomly captured animals was 45% (10/22; 95% CI 24.6%–66.3%) (online Technical Appendix Table 1).

Bme13 was isolated from 5 of 11 seropositive Alpine ibex (1 Alpine ibex was killed in an avalanche). Three seropositive but culture-negative ibex showed positive results by PCR (online Technical Appendix Table 2). Multilocus variable number tandem repeat analysis showed similarity among all strains isolated in this study and strains isolated from local domestic outbreaks >13 years ago (10).

Although persistence of *B. melitensis* in wild ruminants has not been reported, and these animals are considered an epidemiologic dead-end reservoir (3), the unexpected prevalence observed ($\approx 50\%$) suggests that Alpine ibex could be the source of bovine brucellosis reemergence in the study area in France. Strict surveillance policies have prevented infection of domestic livestock with *B. melitensis* in the study area since 1999. However, cohabitation of domestic and wild ruminants on pastures during the summer is rare but possible. Clinical signs and lesions observed in chamois and Alpine ibex are consistent with those reported for chamois and Alpine ibex with brucellosis (4,5). However, positive cultures were obtained from conventional target organs (knee, testes, and lymph nodes) but also from urogenital fluids, which indicates the potential for excretion of the organism.

The fact that births occur during periods and in places where female Alpine ibex are not in close contact with other wild/domestic species (because of higher altitude or rocky peaks) could explain the low transmission rate of *B. melitensis* to these animals. It also suggests that the venereal route might contribute to transmission within Alpine ibex during the mating season in winter. This report demonstrates the need for maintaining an active/reactive surveillance system for livestock and wildlife in brucellosis-free regions.

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***Clostridium tetani* Osteitis without Tetanus**

To the Editor: Posttraumatic osteoarticular infections caused by *Clostridium* spp. are rare, and their outcomes are often unfavorable because of the persistence of the bacteria in bone (1,2). In a recent series of 12 patients (2), only 1 case of posttraumatic osteoarticular infection was caused by *C. tetani* (fracture of the distal humerus with polymicrobial infection). However, no information was available about the production of tetanospasmin by the infecting strain.

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Technical Appendix

Technical Appendix Table 1. Test results for domestic and wild ruminants for infection with *Brucella melitensis*, France*

Animal	No. herds	No. animals	Serologic tests												Bacteriologic and PCR			
			RBT		CFT†		iELISA‡				cELISA§				Total positive#	Culture**	RT-PCR††	
			+/-	NR	+/-	NR	Serum		Lung¶		Serum		Lung¶					
Domestic																		
Cattle	160	8,489	31/8,453	5	11/31	0	ND ^j	ND	ND	ND	ND	ND	ND	ND	11	0/11	ND	
Sheep and goats	45	3,634	5/3,627	2	3/5	0	ND	ND	ND	ND	ND	ND	ND	ND	3	0/3	ND	
Wild																		
Roe deer‡‡§§	NA	44¶¶	0/20	24	0/13	31	0/26	18	0/16	2	0/26	18	0/16	2	0	ND	ND	
Red deer‡‡§§	NA	30	0/24	6	0/18	12	0/28	2	0/2	0	1/27	2	0/2	0	1	0/1	0/1	
Chamois‡‡§§	NA	55	1/33	21	1/29	25	0/44	11	0/11	0	1/43	11	0/11	0	1	1/0	ND	
Alpine ibex	NA	24##	12/12	0	10/6	8	12/12	0	NR	NR	12/12	0	NR	NR	12	5/6	3/3	

*RBT, Rose Bengal test; CFT, complement fixation test; iELISA, indirect ELISA; cELISA, competitive ELISA; RT-PCR, real-time PCR; +/-, no. positive/no. negative; NR, no result (samples were missing or unsuitable for testing because of poor quality or inconclusive results [hemolysis or anti-complementary activity]); ND, not determined.

†CFT – cutoff value = 20 international CFT units/mL.

‡iELISA cutoff value 110%–120%.

§cELISA: cutoff value = 40%.

¶Lung extract was tested by ELISAs only when corresponding serum was missing.

#Number of domestic animals with a positive result by RBT and CFT, number of wild animals with a positive result by either test.

**Culture was performed only for seropositive animals: on retropharyngeal, retromammary, and iliac lymph nodes from domestic animals; on spleen and testes or uterus from hunted animals; and on retropharyngeal, retromammary, and superficial inguinal or iliac lymph nodes, uterus or testes, urine (directly sampled from the bladder), vaginal or preputial swab specimens, udder; and fluid from arthritic area from necropsied alpine ibex.

††RT-PCR was performed for seropositive/culture-negative animals, on testes and spleen from red deer, and on lymph nodes from alpine ibex.

‡‡Total number of animals sampled was 88% (roe deer), 60% (red deer), and 110% (chamois) of the target populations to detect at least 5% prevalence at a 95% CI.

§§Age: chamois, 6 mo–11 y; red and roe deer, young, subadults, and adults.

¶¶44 animals were hunted but blood or lung extract from only 42 animals showed a reliable result.

##Including 2 males with clinical signs and 22 animals randomly captured.

Technical Appendix Table 2. Bacteriologic and PCR results for wild animals seropositive for infection with *Brucella melitensis*, France*

Animal	ID no.	Sex	Age, y	Serologic tests†				Bacteriologic and PCR	
				RBT	CFT	iELISA‡	cELISA§	Culture	PCR
Red deer	4932	M	Adult	–	–	– (57.15)	+ (54.64)	–¶	–¶
Chamois	7464	F	7	+	+ (213)	– (21.93)	+ (94.51)	+ #	ND
Alpine ibex	83	M	11	+	+ (80)	+ (306.97)	+ (94.23)	+ **	ND
	84	M	13	+	+ (160)	+ (334.80)	+ (95.46)	+ ††	ND
	2	F	9	+	+ (53)	+ (219.17)	+ (95.31)	–	–
	3	F	12	+	+ (40)	+ (215.97)	+ (86.72)	–	–
	4	F	11	+	+ (320)	+ (251.58)	+ (95.98)	–	+ ††
	5	F	12	–	–	+ (234.19)	+ (82.98)	–	+ §§
	13	F	9	+	+ (320)	+ (288.08)	+ (96.30)	+ ¶¶	ND
	15	F	14	+	+ (320)	+ (336.27)	+ (94.61)	+ ###	ND
	16	F	9	+	+ (160)	+ (266.81)	+ (95.72)	–	–
	24	M	11	–	–	+ (340.37)	+ (93.04)	ND***	ND***
25	F	13	+	+ (93)	+ (248.32)	+ (93.89)	–	+ §§	
26	F	8	+	+ (426)	+ (303.06)	+ (96.23)	+ †††	ND	

*ID, identification; RBT, Rose Bengal test; CFT, complement fixation test; iELISA, indirect ELISA; cELISA, competitive ELISA; –, negative; +, positive; ND, not determined.

†Values are international CFT units/mL (CFT – cutoff value = 20 international CFT units/mL) or percentages (ELISAs).

‡iELISA cutoff value was 110%–120%.

§cELISA cutoff value was 40%.

¶Spleen/testes.

#Arthritis fluid/testes.

**Arthritis fluid, spleen, preputial swab, urine, right testicle, retropharyngeal specimen, lymph nodes.

††Arthritis fluid, nuchal abscess, spleen, preputial swab, urine (directly sampled from the bladder), right testicle, retropharyngeal specimen, lymph nodes.

‡‡Retropharyngeal specimen, lymph nodes.

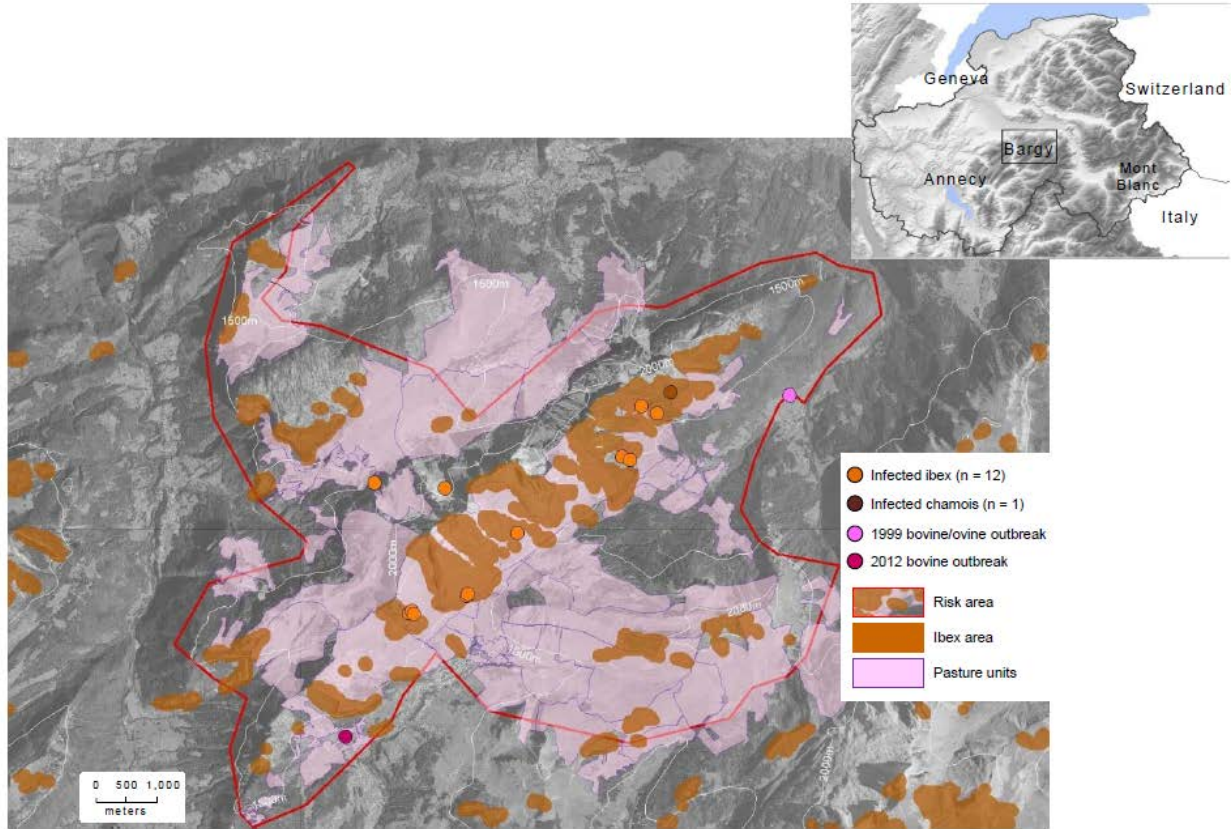
§§Iliac region, lymph nodes.

¶¶Arthritis fluids.

###Udder, mammary abscess.

***Died in avalanche.

†††Vaginal swab, uterus, retropharyngeal specimen, lymph nodes.



Technical Appendix Figure. Areas in France in which alpine ibex and chamois were found infected with *Brucella melitensis* during the study and in which brucellosis outbreaks occurred in ruminants during 1999 and 2012. The region shown is Bargy Massif, which is part of Bornes Massif in Haute-Savoie, France, and borders Switzerland and Italy. The elevation range for most of Bargy Massif is 900–2,300 m. However, the highest point has an elevation of 2,438 m. Alpine ibex areas were defined according to 3 criteria: >20% slope; region (southern, southwestern, and southeastern); and rocky substratum. The risk corresponds to the area in which the probability of cohabitation of wild and domestic ruminants is high.