nuttalli parasites are as virulent as E. histolytica parasites in animal models, it remains unclear whether they are as virulent in humans (4). We recommend that caretakers of NHPs be screened on a regular basis and be provided with appropriate medications if needed.

Acknowledgments
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References

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Alaria alata Mesocercariae among Feral Cats and Badgers, Denmark

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To the Editor: The digenean trematode Alaria alata is considered an emerging zoonotic parasite in Europe because of increased findings in wild boars during Trichinella inspection. No human illness caused by A. alata mesocercariae (infective larvae) has been reported, but concern remains because the closely related North American species A. americana has caused illnesses among humans, including 1 death (1).

In Denmark, high prevalence of A. alata trematodes in final hosts has been shown (2), but limited data on potential paratenic hosts are available. Therefore, samples from 406 domestic pigs, 130 wild boars, 9 badgers, and 99 cats were collected by convenience sampling during October 2013–September 2014. We used pig and wild boar samples from multiple geographic areas of Denmark were leftover tissue samples from ongoing Trichinella spp. surveillance. Badgers had died naturally or were hit by vehicles (8 from Jutland, 1 from Zealand) and collected as part of a wildlife monitoring program. Cats (all from Zealand) were either feral (n = 92) or domesticated (n = 7) and had been euthanized as part of a national control program. Carcasses were necropsied in our laboratory; we collected 30 g of tissue samples according to Riehn et al. (3). All samples were analyzed by the modified A. alata mesocercariae migration technique (3). In brief, the sample was cut into ≈0.5-cm edge pieces, wrapped in gauze, and suspended for 90 min by 2 wooden sticks in a conical glass with ≈300 mL of water (46°C–48°C). Approximately 15 mL of sediment was collected from the bottom of the glass by suction using a glass pipette and examined by microscopy (magnification ×20).

A. alata mesocercariae were isolated from 3 cats and 6 badgers (online Technical Appendix Table 1, http://wwwnc.cdc.gov/EID/article/20/10/14-1817-Techapp1.pdf). All 3 cats were female (2 pregnant, 1 lactating); prevalence was significantly higher in pregnant or lactating females (3/12) than other intact females (0/24) (p = 0.031 by Fisher exact test). This finding might be related to increased exposure because an increase in predation by the cats during pregnancy and lactation to meet higher protein and energy demand. However, because A. marciannae mesocercariae can be
transmitted through milk in cats (4), lactating females may also be predisposed to an increased chance for *A. alata* mesocercariae reaching their offspring. Examination of the intestines of all cats by sedimentation and counting technique (5) revealed no *A. alata* adults. Although *A. alata* adults have been found in cats in Uruguay (6), reports from Europe are lacking, and thus it is still uncertain whether cats can act as amphiparatenic or final hosts. Natural infection of cats with other *Alaria* spp. has been reported in the United States (7), indicating biologic differences among *Alaria* spp.

Zoonotic risk for *A. alata* infection through ingestion of cat meat is probably minimal in Europe but may be important in Asia and South America, where cats are occasionally consumed. Badgers are, however, sometimes consumed as game meat or road kill meat in Europe. In Russia, 10.6% of trichinellosis outbreaks during 1998–2002 were caused by consumption of badger meat (8). Thus, the zoonotic potential of infections in this animal, although a protected species, should not be ignored.

Negative findings in domestic pigs and wild boars in this study may reflect underestimation because those samples were below the recommended 30 g and often taken from sites that are not typically infected with mesocercariae (3). A follow-up study with better sampling strategy would be of value to determine the risk for *A. alata* transmission from domestic pigs and wild boars.

Identification of isolated mesocercariae was confirmed by PCR and sequencing of a fragment (332 bp) of the mitochondrial cytochrome c oxidase subunit 1 gene (cox1) (9). By neighbor-joining analysis (10), the consensus cox1 sequences were compared with the trematode *Neodiplostomum seoulense* (outgroup), 1 *A. alata* isolate from a Danish red fox, and all 7 cox1 sequences of *Alaria* spp. available in GenBank as of October 2014. (Sequences from this study have been deposited into GenBank under accession nos. KP123417–KP123422 [badgers] KP123423–KP123425 [feral cats].) The inferred phylogenetic tree (Figure) showed marked genetic variation among *A. alata* isolates from Denmark and other parts of Europe but no apparent separation of most *A. alata* isolates from Europe based on host species or country, except for that from badger 1 (online Technical Appendix Table 2). This animal originated from Northern Jutland, where host and parasite populations are geographically isolated by a large fjord separating the region from the rest of the country. The marked genetic variation within cox1 sequences suggests the usefulness of this marker, but additional genetic markers should be included in future studies to explore the genetic flow of *A. alata* within natural hosts.

In conclusion, *A. alata* mesocercariae seem to favorably infect pregnant or lactating cats, thereby increasing the chance of vertical transmission. Further, detection of *A. alata* infection in numerous badgers suggests potential high zoonotic risk associated with ingestion of such exotic meat. These results should, however, be interpreted with caution because of the small sample size and unknown efficacy of the modified *A. alata* mesocercariae migration technique.

**Acknowledgments**

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**References**

LETTERS

Human Infections with *Pseudoterranova cattani* Nematodes, Chile

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To the Editor: Anisakidosis is an emerging foodborne zoonosis caused by nematode larvae of the Anisakinae subfamily, which includes the genera *Anisakis*, *Pseudoterranova*, and *Contracecum*. In natural cycles, anisakid larvae are transmitted to marine mammals or piscivorous birds when they eat raw saltwater fish or squid. In the human incidental host, larvae attach to the mucosa of the gastrointestinal tract, causing clinical features ranging from asymptomatic carriage to severe abdominal pain with complications, such as gastric perforation. Microscopical diagnosis is hampered by the lack of distinguishing morphologic characteristics in larval stages. Recently, molecular genetic techniques have shown that the main species, *Anisakis simplex* and *Pseudoterranova decipiens*, are in fact species groups with distinct geographic and biologic characteristics. The *P. decipiens* complex consists of at least 6 sibling species. We report 4 human infections with *P. cattani* diagnosed during 2012–2014.

The case-patients were adults 22–59 years of age; 2 were female, and all lived in Santiago, Chile. Additional anamnestic and clinical data were available for 3 patients: all spontaneously regurgitated the parasites without having other gastrointestinal complaints. All 3 reported eating ceviche, a dish made of raw marine fish marinated in lemon juice. One patient reported a tingling sensation and coughs before the expulsion of a highly motile larva. The patient was awaiting oral surgery after a bicycle accident and had eaten the last raw fish dish 2 weeks previously. Initially, parasites were identified by morphologic criteria. Larvae were 20 mm long, were of whitish to reddish color, and had 3 anterior lips. Molecular identification, DNA samples were extracted by using a DNeasy Blood and Tissue Kit (QIAGEN K.K., Tokyo, Japan). The rRNA gene containing 2 internal transcribed spacer (ITS) regions was amplified by PCR using primers NC5 and NC2, as previously described. PCR products were sequenced by using a BigDye Terminator Cycle Sequencing Kit (Applied Biosystems Inc., Foster City, CA, USA) on an automated sequencer (ABI3100, Applied Biosystems). Sequence similarities were determined by a BLAST search of DDBJ (http://blast.ddbj.nig.ac.jp/top-j.html). The GENETYX-WIN program version 7.0 (Software Development Co., Tokyo, Japan) facilitated sequence alignment and comparison. Within the 4 ITS sequences of amplicons obtained, all were 100% identical, and alignment with the other *P. cattani* sequence differed only in 1 nt. ITS sequences of 2 isolates are available in GenBank (accession nos. KF781284 and KF781285). All *P. cattani* sequences showed a previously described

1These authors contributed equally to this article.
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Technical Appendix

Technical Appendix Table 1. Information on domestic animals and wildlife examined for Alaria alata by using modified Alaria alata migration technique, Denmark October 2013–September 2014*

<table>
<thead>
<tr>
<th>Host species</th>
<th>Origin</th>
<th>Examination period</th>
<th>Sample type</th>
<th>No. examined</th>
<th>Prevalence (%) (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pigs Outdoor</td>
<td>Z</td>
<td>Oct 2013–Mar 2014</td>
<td>Diaphragm or foreleg</td>
<td>260</td>
<td>0</td>
</tr>
<tr>
<td>ND</td>
<td>Z, F, J, ND</td>
<td>Oct 2013–Mar 2014</td>
<td>Diaphragm or foreleg</td>
<td>146</td>
<td>0</td>
</tr>
<tr>
<td>Wild boars</td>
<td>Z</td>
<td>Oct 2013–Mar 2014</td>
<td>Diaphragm or foreleg</td>
<td>39</td>
<td>0</td>
</tr>
<tr>
<td>Farmed</td>
<td>J</td>
<td>Nov 2013–Jan 2014</td>
<td>Diaphragm, foreleg, or intercostal muscles</td>
<td>58</td>
<td>0</td>
</tr>
<tr>
<td>ND</td>
<td>ND</td>
<td>Oct 2013–Mar 2014</td>
<td>Diaphragm or foreleg</td>
<td>33</td>
<td>0</td>
</tr>
<tr>
<td>Badgers</td>
<td>J</td>
<td>Feb–Sep 2014</td>
<td>Tissue sample mixture†</td>
<td>9</td>
<td>66.7 (29.9–92.5)</td>
</tr>
<tr>
<td>Cats</td>
<td>Z</td>
<td>Mar–May 2014</td>
<td>Tissue sample mixture†</td>
<td>99</td>
<td>3.0 (0.6–8.6)</td>
</tr>
</tbody>
</table>

*Z, Zealand; F, Funen; J, Jutland; ND, no data available.
†30 g tissue sample mixture of diaphragm, tongue, and masticatory muscles were used.

Technical Appendix Table 2. Characteristics of Alaria alata mesocercariae–positive animal tissue samples identified by using modified Alaria alata migration technique, Denmark

<table>
<thead>
<tr>
<th>Identity code</th>
<th>Species</th>
<th>Sex</th>
<th>Origin</th>
<th>No. of mesocercariae per 30 g sample</th>
<th>GenBank accession number</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cat 43</td>
<td>Cat</td>
<td>Female (pregnant)</td>
<td>Zealand</td>
<td>23</td>
<td>KP123423</td>
</tr>
<tr>
<td>Cat 100</td>
<td>Cat</td>
<td>Female (pregnant)</td>
<td>Zealand</td>
<td>2</td>
<td>KP123424</td>
</tr>
<tr>
<td>Cat 120</td>
<td>Cat</td>
<td>Female (lactating)</td>
<td>Zealand</td>
<td>12</td>
<td>KP123425</td>
</tr>
<tr>
<td>Badger 1</td>
<td>Badger</td>
<td>Not recorded</td>
<td>Northern Jutland</td>
<td>1*</td>
<td>KP123417</td>
</tr>
<tr>
<td>Badger 2</td>
<td>Badger</td>
<td>Female</td>
<td>Central Jutland</td>
<td>2</td>
<td>KP123418</td>
</tr>
<tr>
<td>Badger 4</td>
<td>Badger</td>
<td>Female</td>
<td>Southern Jutland</td>
<td>1</td>
<td>KP123419</td>
</tr>
<tr>
<td>Badger 5</td>
<td>Badger</td>
<td>Male</td>
<td>Central Jutland</td>
<td>2</td>
<td>KP123420</td>
</tr>
<tr>
<td>Badger 6</td>
<td>Badger</td>
<td>Male</td>
<td>Central Jutland</td>
<td>8</td>
<td>KP123421</td>
</tr>
<tr>
<td>Badger 9</td>
<td>Badger</td>
<td>Female</td>
<td>Zealand</td>
<td>2</td>
<td>KP123422</td>
</tr>
</tbody>
</table>

*30 g tissue sample mixture of diaphragm, tongue, and masticatory muscles were used.