

Close Relationship of Ruminant Pestiviruses and Classical Swine Fever Virus

Alexander Postel, Stefanie Schmeiser,
Tuba Cigdem Oguzoglu, Daniela Indenbirken,
Malik Alawi, Nicole Fischer, Adam Grundhoff,
Paul Becher

To determine why serum from small ruminants infected with ruminant pestiviruses reacted positively to classical swine fever virus (CSFV)-specific diagnostic tests, we analyzed 2 pestiviruses from Turkey. They differed genetically and antigenically from known *Pestivirus* species and were closely related to CSFV. Cross-reactions would interfere with classical swine fever diagnosis in pigs.

Pestiviruses are enveloped viruses within the family *Flaviviridae* that have a highly variable single-stranded positive-sense RNA genome of ≈ 12.3 kb (1). The genus *Pestivirus* comprises the established species bovine viral diarrhea virus (BVDV)-1, BVDV-2, border disease virus (BDV), and classical swine fever virus (CSFV), as well as a growing number of additional tentative *Pestivirus* species. CSFV is the causative agent for classical swine fever, which is notifiable to the World Organisation of Animal Health because it is highly contagious and can cause great loss of pigs (2–4). For a given country, CSFV-positive status severely diminishes international trade of pigs and pig products. Accordingly, because of cross-reacting antibodies, infections of pigs (nonruminants) with ruminant pestiviruses, which occasionally occur under natural conditions, can cause serious problems with regard to serologic diagnosis of classical swine fever (5).

In Turkey, 2 pestiviruses, Aydin/04 and Burdur/05, have been isolated from a sheep and a goat with clinical signs of border disease (6). A detailed genetic and antigenic characterization revealed that these 2 isolates must be regarded as representatives of a new *Pestivirus* species that is closely related to CSFV and can cause serious diagnostic problems in established CSFV serology.

Author affiliations: University of Veterinary Medicine, Hannover, Germany (A. Postel, S. Schmeiser, P. Becher); Ankara Üniversitesi Veteriner Fakültesi Viroloji Anabilim Dalı Dışkapı, Ankara, Turkey (T.C. Oguzoglu); Heinrich Pette Institute—Leibniz Institute for Experimental Virology, Hamburg, Germany (D. Indenbirken, M. Alawi, A. Grundhoff); University Medical Center Hamburg-Eppendorf, Hamburg (M. Alawi, N. Fischer); German Center for Infection Research Hamburg-Lübeck-Borstel, Hamburg (N. Fischer, A. Grundhoff); German Center for Infection Research Hannover-Braunschweig, Hannover (P. Becher)

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The Study

During 2004–2007, serum samples from 1,036 sheep and goats in Turkey were serologically screened for infection with pestiviruses of small ruminants. Of these, 11 serum samples from 7 sheep herds gave positive or doubtful reactions in the CSFV antibody-specific ELISA (HerdChek, IDEXX) and were subjected to commonly used virus neutralization testing (VNT) (7). VNT against the 2 established CSFV strains Alfort187 (genotype 1.1) and Diepholz (genotype 2.3) and against the BDV strains Moredun (genotype 1) and Gifhorn (genotype 3) revealed higher BDV titers in only 3 serum samples (Table 1). Equal or slightly higher titers against the CSFV reference strains became evident in 8 of the 11 serum samples, which came from 5 regions of Turkey. Further VNT analyses with the 2 previously obtained isolates, Aydin/04 and Burdur/05, demonstrated neutralizing antibody titers that were equal or higher than those against BDV and CSFV test strains. To elucidate the reason for strong serologic reactivity in CSFV assays, we genetically and antigenically characterized pestiviruses Aydin/04 and Burdur/05.

The complete genome sequence of Aydin/04 was determined as reported previously (8). The genome sequence of Burdur/05 was determined by next-generation sequencing on an Illumina MiSeq platform (2 × 250-bp paired end run, 593,328 reads) as recently described (9). Template total cellular RNA was extracted from supernatant of sheep fetal thymus cells. Of all reads, 73.9% were found to be of host origin. Of the nonhost reads, 89.9% assembled into a single sequence contig encompassing the entire pestivirus Burdur/05 genome (coverage 196-fold).

Sequence and phylogenetic analyses were performed with complete genome sequences and deduced amino acid sequences of new pestiviruses Aydin/04 (GenBank accession no. JX428945) and Burdur/05 (KM408491). For further analyses, reference sequences were obtained from GenBank (Figure 1). Genetic distances were calculated by using the Kimura 2-parameter substitution model, and phylogenetic analyses were conducted by applying the neighbor-joining method as commonly used for CSFV phylogeny (11). With the same set of sequences, a grouping scan was performed by using the SSE platform (12). Comparison of the complete coding sequences of Aydin/04 and Burdur/05 revealed a genetic distance of 16.5%. Phylogenetic analyses based on deduced polyprotein sequences showed that isolates Aydin/04 and Burdur/05 form a distinct group located between CSFV and BDV (Figure 1, panel A).

Table 1. Serologic pestivirus testing results for sheep from different districts in Turkey, 2004–2007*

Sample	Province of origin	CSFV ELISA†	Neutralizing antibody titer, ND ₅₀						
			BVDV NADL	CSFV‡		BDV§		Pestiviruses from Turkey	
				Alfort187	Diepholz	Moredun	Gifhorn	Aydin	Burdur
1	Antalya¶	+	<5	320	226	28	57	905	905
2	Antalya¶	+	<5	113	113	28	40	226	160
3	Burdur	+	<5	40	640	113	226	226	226
4	Burdur	?	<5	226	226	160	226	640	453
5	Konya	?	<5	<80	<40	160	113	113	57
6	Konya¶	+	<5	160	905	320	160	320	452
7	Konya¶	?	<5	≤28	14	20	10	40	40
8	Kütahya¶	+	<5	≤15	≤15	160	640	57	113
9	Kütahya¶	+	<5	≤15	453	640	640	113	57
10	Kütahya¶	+	<5	28	160	160	160	226	80
11	Sanliurfa	+	<5	226	<1,280	<320	160	452	452

*BDV, border disease virus; BVDV, bovine viral diarrhea virus; CSFV, classical swine fever virus; ND50, 50% neutralizing dose; ?, doubtful result.

†HerdChek (IDEXX).

‡CSFV strains: Alfort187 (genotype 1.1), Diepholz (genotype 2.3).

§BDV strains: Moredun (genotype 1), Gifhorn (genotype 3).

¶Serum obtained from the same herd (samples 1 and 2, 6 and 7, 8–10).

Systematic antigenic characterization was performed by using cross-neutralization assays (Table 2). For this purpose, CSFV and BDV reference strains for which homologous serum was available were tested by VNT as described

(7). In general, neutralization of both isolates was more efficient when performed with different CSFV antiserum than with BDV antiserum. In addition, the Aydin-specific antiserum obtained from animal experiments neutralized

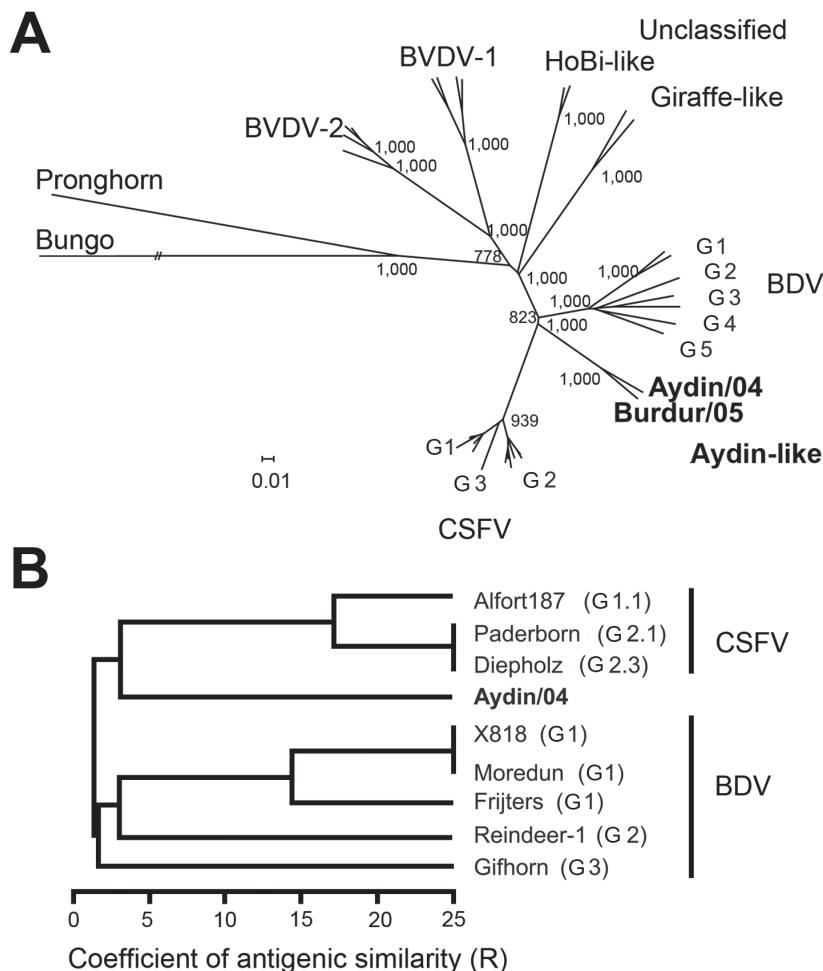


Figure 1. Phylogenetic and antigenic tree displaying relatedness of pestiviruses Aydin/04 and Burdur/05 to other *Pestivirus* species. A) For phylogenetic analysis, deduced polyprotein sequences from GenBank were used (CSFV: J04358, GU233734, JX218094, AY568569, GQ902941, KJ619377, AY382481, AF326963, X87939, AF099102, AY578687, AY646427; BDV: AF037405, U70263, KC963426, AF144618, GQ902940, KF918753, GU270877; BVDV-1: EF101530, AF220247, M96751, AF091605; BVDV-2: AB567658, GQ888686, AF502399, U18059; HoBi-like: AB871953, NC012812; giraffe-like: NC003678, KJ660072; Pronghorn and Bungowannah: NC024018, NC023176). Bootstrap values were calculated for 1,000 iterations. Only significant bootstrap values (≥ 700) of major nodes are given in the tree. Trees were displayed by using Dendroscope (10). Scale bar indicates base substitutions per site. B) Antigenic tree based on coefficients of antigenic similarity (R values) displaying antigenic relatedness of pestiviruses Aydin/04 and Burdur/05 to representative CSFV and BDV strains. R values < 25 indicate significant antigenic differences as representing > 4 -fold differences in titers. R values > 25 are considered not significant and are therefore not drawn to scale. Boldface indicates pestiviruses circulating among sheep and goat herds in Turkey. BDV, border disease virus; BVDV, bovine viral diarrhea virus; CSFV, classical swine fever virus; G, genotype.

Table 2. Antigenic relationships determined by cross-neutralization of serum raised against different CSFV and BDV reference strains*

Strain	CSFV†				BDV‡					
	Alfort187	Diepholz	Paderborn	Aydin	Burdur	Moredu	Frijters	X818	Reindeer	Gifhorn
Alfort187	1,810	452	113	20	80	5	10	≤3.5	10	12
Diepholz	381	1,280	452	80	135	57	28	10	48	28
Paderborn	34	160	452	≤3.5	14	28	5	8.4	14	7,1
Aydin	40	40	20	761	135	8.4	14	8.4	34	24
Moredu	≤3.5	4.2	≤3.5	≤3.5	≤3.5	190	4.2	95	≤3.5	≤3.5
Frijters	14	20	17	14	67	113	761	269	134	12
X818	14	160	95	24	30	905	381	2,560	381	226
Reindeer	8.4	17	8.4	4.2	17	80	8.4	48	2,560	40
Gifhorn	14	80	67	28	80	30	40	67	134	5,120

*Boxes indicate virus neutralization test titers for homologous serum; boldface indicates results for Aydin/04 and Burdur/05. BDV, border disease virus; CSFV, classical swine fever virus.

†CSFV strains: Alfort187 (genotype 1.1), Diepholz (genotype 2.3), Paderborn (genotype 2.1).

‡BDV strains: Moredu (genotype 1), Frijters (genotype 1), X818 (genotype 1), Reindeer (genotype 2), Gifhorn (genotype 3).

the CSFV reference strains with titers higher than those for the BDV strains (Table 2). Because no experimental infection with Burdur/05 has been performed, Burdur/05-specific antiserum was not available; however, close antigenic relatedness of both isolates was demonstrated by the high neutralization titers of the Aydin-specific antiserum for isolate Burdur/05 (Table 2). To quantify and to depict the antigenic relatedness, we calculated coefficients of antigenic similarity (R values) as described previously (13). An antigenic tree graphically displaying the R values clearly shows 2 distinct clades, one representing CSFV and the other comprising BDV strains (Figure 1, panel B). Furthermore, Aydin/04 is antigenically more closely related to CSFV than to BDV, but it also clearly differs from these 2 pestivirus species.

Because of their close relationship to CSFV, it was of particular interest to determine the ability of these ruminant pestiviruses to infect pigs and induce clinical disease. Therefore, 3 clinically healthy and pestivirus uninfected weaner (6 weeks of age) piglets were inoculated with 1×10^6 50% tissue culture infectious doses of isolate Aydin/04 and given a booster of 3×10^7 50% tissue culture infectious doses 2 weeks later. Pigs showed no clinical signs of disease, no fever, no platelet or leukocyte depletion, and no viremia (data not shown). For all 3 animals, strong seroconversion was found (50% neutralizing titer of serum for homologous virus was 240–640 on postinoculation day 77).

Conclusions

Several new genetically diverse groups of pestiviruses have emerged in domestic livestock and wild animals, adding to the continuously growing list of approved and tentative pestivirus species (1). According to phylogenetic analyses of short partial genome sequences, 2 pestivirus isolates, Aydin/04 and Burdur/05, recently circulating in sheep and goat herds in different regions of Turkey, were classified as novel members of the BDV species (6). However, the data from this study demonstrate that these novel Aydin-like pestiviruses are representatives

of a new pestivirus species, genetically and antigenically located between CSFV and BDV (Figure 1). The genetic distance of 16.5% between these isolates indicates that distinct ruminant Aydin-like pestiviruses circulate in different regions of Turkey. For some genomic regions, both ruminant pestivirus isolates display an even higher similarity to CSFV than to BDV (Figure 2). The close genetic relatedness to CSFV is in line with the antigenic characterization by cross-neutralization assays as depicted in the antigenic tree (Figure 1, panel B). This finding is in contrast with findings for pestivirus isolates from Tunisia, another group of ruminant pestiviruses genetically closely related to CSFV but antigenically more closely related to BDV (14). The close antigenic relationship to CSFV explains the observed strong cross-reactivity of serum from sheep and goat in CSFV-specific ELISAs, even when the variable E2 protein is used as diagnostic antigen (Table 1). In routine diagnosis, questionable ELISA results are further investigated by VNT against CSFV and other pestiviruses (e.g., BVDV and BDV). Usually, VNT titers are highest for the homologous pestivirus species. Remarkably, even if representatives of the Aydin-like pestiviruses were included as test strains in the VNT, CSFV infection still could not be ruled out.

Although these novel pestiviruses are the closest known relatives of CSFV, experimental infection of pigs with Aydin/04 did not result in detectable viremia and clinical signs. Nevertheless, these ruminant pestiviruses are candidates for a switch to porcine hosts after ongoing virus evolution, which would have severe consequences for serologic diagnosis of classical swine fever, affecting control and monitoring programs performed in many parts of the world.

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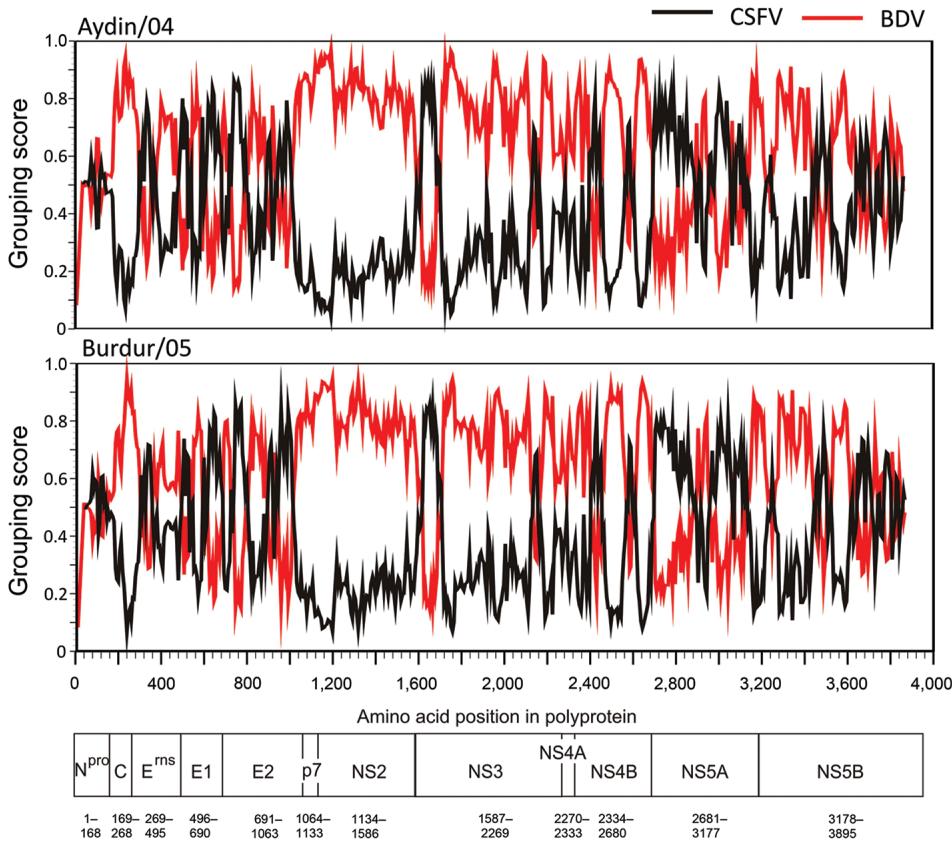


Figure 2. Amino acid similarity of pestiviruses Aydin/04 and Burdur/05 to representative CSFV and BDV polyprotein sequences. The same CSFV and BDV polyprotein sequences as in Figure 1 were used for analysis. Grouping scan was performed with the SSE software platform as described previously, by using a window of 200 aa with 20-aa increments (12). For calculation of genetic distances, the Kimura 2-parameter model was applied. Borders of the mature viral proteins in the polyprotein of Aydin/04 are given below. BDV, border disease virus; CSFV, classical swine fever virus; C, core protein; E, envelope protein; rms, ribonuclease secreted; N^{pro}, N-terminal autoprotease; NS, nonstructural protein; p7, protein p7.

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Dr. Postel is a veterinarian and head of the Laboratory for Molecular Biology of the European Union and World Organisation for Animal Health Reference Laboratory for Classical Swine Fever at the Institute of Virology of the University of Veterinary Medicine in Hannover, Germany. His research interests are molecular evolution of pestiviruses, characterization of novel pestivirus isolates, and diagnosis and control of classical swine fever.

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Address for correspondence: Paul Becher, EU and OIE Reference Laboratory for Classical Swine Fever, Institute of Virology, Department of Infectious Diseases, University of Veterinary Medicine Hannover, Buenteweg 17, 30559 Hannover, Germany; email: paul.becher@tiho-hannover.de

The image shows a screenshot of the CDC's Facebook page. At the top, there's a navigation bar with the Facebook logo and a search bar. Below that, a large banner promotes the 'Solve the Outbreak' iPad app, featuring a graphic of a network diagram on a tablet. A sign-up banner for 'New outbreaks! CDC is on Facebook' is overlaid on the right side of the banner, with 'Sign Up' and 'Log In' buttons. Below the banner, the CDC profile information is visible, including the name 'CDC', a verified checkmark, and the text '263,397 likes · 3,144 talking about this'. The 'About' section describes the CDC as a Government Organization dedicated to protecting health and promoting quality of life. Below this, there are sections for 'Photos', 'Likes' (showing 263k), 'Vital Signs', and 'Welcome'. A 'Highlights' dropdown menu is visible. The main content area shows a post from CDC shared a link about heatwave safety tips, and a section for 'Recent Posts by Others on CDC' featuring posts from Carol Ferguson, Thomas Roles, and Najim Samourai.

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