Close Relationship of Ruminant Pestiviruses and Classical Swine Fever Virus

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To determine why serum from small ruminants infected with ruminant pestiviruses reacted positively to classical swine fever virus (CSFV)–specific diagnostic tests, we analyzed 2 pestiviruses from Turkey. They differed genetically and antigenically from known Pestivirus species and were closely related to CSFV. Cross-reactions would interfere with classical swine fever diagnosis in pigs.

Pestiviruses are enveloped viruses within the family Flaviviridae that have a highly variable single-stranded positive-sense RNA genome of ~12.3 kb (1). The genus Pestivirus comprises the established species bovine viral diarrhea virus (BVDV)–1, BVDV–2, border disease virus (BDV), and classical swine fever virus (CSFV), as well as a growing number of additional tentative Pestivirus species. CSFV is the causative agent for classical swine fever, which is notifiable to the World Organization for Animal Health because it is highly contagious and can cause great loss of pigs (2–4). For a given country, CSFV-positive status severely diminishes international trade of pigs and pig products. Accordingly, because of cross-reacting antibodies, infections of pigs (nonruminants) with ruminant pestiviruses, which occasionally occur under natural conditions, can cause serious problems with regard to serologic diagnosis of classical swine fever (5).

In Turkey, 2 pestiviruses, Aydin/04 and Burdur/05, have been isolated from a sheep and a goat with clinical signs of border disease (6). A detailed genetic and antigenic characterization revealed that these 2 isolates must be regarded as representatives of a new Pestivirus species that is closely related to CSFV and can cause serious diagnostic problems in established CSFV serology.

The Study

During 2004–2007, serum samples from 1,036 sheep and goats in Turkey were serologically screened for infection with pestiviruses of small ruminants. Of these, 11 serum samples from 7 sheep herds gave positive or doubtful re-actions in the CSFV antibody–specific ELISA (HerdChek, IDEXX) and were subjected to commonly used virus neutralization testing (VNT) (7). VNT against the 2 established CSFV strains Alfort187 (genotype 1.1) and Diepholz (genotype 2.3) and against the BDV strains Moreduin (genotype 1) and Gifhorn (genotype 3) revealed higher BDV titers in only 3 serum samples (Table 1). Equal or slightly higher titers against the CSFV reference strains became evident in 8 of the 11 serum samples, which came from 5 regions of Turkey. Further VNT analyses with the 2 previously obtained isolates, Aydin/04 and Burdur/05, demonstrated neutralizing antibody titers that were equal or higher than those against BDV and CSFV test strains. To elucidate the reason for strong serologic reactivity in CSFV assays, we genetically and antigenically characterized pestiviruses Aydin/04 and Burdur/05.

The complete genome sequence of Aydin/04 was determined as reported previously (8). The genome sequence of Burdur/05 was determined by next-generation sequencing on an Illumina MiSeq platform (2 × 250-bp paired end run, 593,328 reads) as recently described (9). Template total cellular RNA was extracted from supernatant of sheep fetal thymus cells. Of all reads, 73.9% were found to be of host origin. Of the nonhost reads, 89.9% assembled into a single sequence contig encompassing the entire pestivirus Burdur/05 genome (coverage 196-fold).

Sequence and phylogenetic analyses were performed with complete genome sequences and deduced amino acid sequences of new pestiviruses Aydin/04 (GenBank accession no. JX428945) and Burdur/05 (KM408491). For further analyses, reference sequences were obtained from GenBank (Figure 1). Genetic distances were calculated by using the Kimura 2-parameter substitution model, and phylogenetic analyses were conducted by applying the neighbor-joining method as commonly used for CSFV phylogeny (10). With the same set of sequences, a grouping scan was performed by using the SSE platform (12). Comparison of the complete coding sequences of Aydin/04 and Burdur/05 revealed a genetic distance of 16.5%. Phylogenetic analyses based on deduced polypeptide sequences showed that isolates Aydin/04 and Burdur/05 form a distinct group located between CSFV and BDV (Figure 1, panel A).
Systematic antigenic characterization was performed by using cross-neutralization assays (Table 2). For this purpose, CSFV and BDV reference strains for which homologous serum was available were tested by VNT as described (7). In general, neutralization of both isolates was more efficient when performed with different CSFV antiserum than with BDV antiserum. In addition, the Aydin-specific antiserum obtained from animal experiments neutralized

![Table 1. Serologic pestivirus testing results for sheep from different districts in Turkey, 2004–2007*](image)

<table>
<thead>
<tr>
<th>Sample</th>
<th>Province of origin</th>
<th>CSFV ELISA†</th>
<th>BVDV Neutralizing antibody titer, ND50</th>
<th>CSFV§</th>
<th>BDV§</th>
<th>Pestiviruses from Turkey</th>
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<tbody>
<tr>
<td>1</td>
<td>Antalya‡</td>
<td>+</td>
<td>&lt;5</td>
<td>320</td>
<td>226</td>
<td>28</td>
</tr>
<tr>
<td>2</td>
<td>Antalya‡</td>
<td>+</td>
<td>&lt;5</td>
<td>113</td>
<td>113</td>
<td>28</td>
</tr>
<tr>
<td>3</td>
<td>Burdur</td>
<td>+</td>
<td>&lt;5</td>
<td>40</td>
<td>640</td>
<td>113</td>
</tr>
<tr>
<td>4</td>
<td>Burdur</td>
<td>?</td>
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<td>226</td>
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<tr>
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<tr>
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<td>≤5</td>
<td>226</td>
<td>1,280</td>
<td>452</td>
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</tbody>
</table>

*BDV, border disease virus; BVDV, bovine viral diarrhea virus; CSFV, classical swine fever virus; ND50, 50% neutralizing dose; ?, doubtful result.
†HerdChek (IDEXX).
‡CSFV strains: Alfort187 (genotype 1.1), Diepholz (genotype 2.3).
§BDV strains: Moredun (genotype 1), Gifhorn (genotype 3).
¶Serum obtained from the same herd (samples 1 and 2, 6 and 7, 8–10).

Figure 1. Phylogenetic and antigenic tree displaying relatedness of pestiviruses Aydin/04 and Burdur/05 to other Pestivirus species. A) For phylogenetic analysis, deduced polyprotein sequences from GenBank were used (CSFV: J04358, GU233734, JX218094, AY568569, GQ902941, KJ619377, AY382481, AF326963, X87939, AF099102, AY578687, AY646427, BDV: AF037405, U70263, KC063426, AF144618, GQ902940, KF918753, GU270877, BVDV-1: EF101530, AF220247, M96751, AF091605, BVDV-2: AB567658, Q888866, AF502399, U18059; HoBi-like: AB871953, NC012812, giraffe-like: NC003678, KJ660072; Pronghorn and Bungowannah: NC024018, NC023176). Bootstrap values were calculated for 1,000 iterations. Only significant bootstrap values (>700) of major nodes are given in the tree. Trees were displayed by using Dendroscope (10). Scale bar indicates base substitutions per site. B) Antigenic tree based on coefficients of antigenic similarity (R values) displaying antigenic relatedness of pestiviruses Aydin/04 and Burdur/05 to representative CSFV and BDV strains. R values <25 indicate significant antigenic differences as representing >4-fold differences in titers. R values >25 are considered not significant and are therefore not drawn to scale. Boldface indicates pestiviruses circulating among sheep and goat herds in Turkey. BDV, border disease virus; BVDV, bovine viral diarrhea virus; CSFV, classical swine fever virus; G, genotype.
the CSFV reference strains with titers higher than those for the BDV strains (Table 2). Because no experimental infection with Burdur/05 has been performed, Burdur/05-specific antiserum was not available; however, close antigenic relatedness of both isolates was demonstrated by the high neutralization titers of the Aydin-specific antiserum for isolate Burdur/05 (Table 2). To quantify and to depict the antigenic relatedness, we calculated coefficients of antigenic similarity (R values) as described previously (13). An antigenic tree graphically displaying the R values clearly shows 2 distinct clades, one representing CSFV and the other comprising BDV strains (Figure 1, panel B). Furthermore, Aydin/04 is antigenically more closely related to CSFV than to BDV, but it also clearly differs from these 2 pestivirus species.

Because of their close relationship to CSFV, it was of particular interest to determine the ability of these ruminant pestiviruses to infect pigs and induce clinical disease. Therefore, 3 clinically healthy and pestivirus uninfected weaner 6 weeks of age) piglets were inoculated with $1 \times 10^6$ 50% tissue culture infectious doses of isolate Aydin/04 and given a booster of $3 \times 10^7$ 50% tissue culture infectious doses 2 weeks later. Pigs showed no clinical signs of disease, no fever, no platelet or leukocyte depletion, and no viremia (data not shown). For all 3 animals, strong seroconversion was found (50% neutralizing titer of serum for homologous virus was 240–640 on postinoculation day 77).

**Conclusions**

Several new genetically diverse groups of pestiviruses have emerged in domestic livestock and wild animals, adding to the continuously growing list of approved and tentative pestivirus species (1). According to phylogenetic analyses of short partial genome sequences, 2 pestivirus isolates, Aydin/04 and Burdur/05, recently circulating in sheep and goat herds in different regions of Turkey, were classified as novel members of the BDV species (6). However, the data from this study demonstrate that these novel Aydin-like pestiviruses are representatives of a new pestivirus species, genetically and antigenically located between CSFV and BDV (Figure 1). The genetic distance of 16.5% between these isolates indicates that distinct ruminant Aydin-like pestiviruses circulate in different regions of Turkey. For some genomic regions, both ruminant pestivirus isolates display an even higher similarity to CSFV than to BDV (Figure 2). The close genetic relatedness to CSFV is in line with the antigenic characterization by cross-neutralization assays as depicted in the antigenic tree (Figure 1, panel B). This finding is in contrast with findings for pestivirus isolates from Tunisia, another group of ruminant pestiviruses genetically closely related to CSFV but antigenically more closely related to BDV (14). The close antigenic relationship to CSFV explains the observed strong cross-reactivity of serum from sheep and goat in CSFV-specific ELISAs, even when the variable E2 protein is used as diagnostic antigen (Table 1). In routine diagnosis, questionable ELISA results are further investigated by VNT against CSFV and other pestiviruses (e.g., BVDV and BDV). Usually, VNT titers are highest for the homologous pestivirus species. Remarkably, even if representatives of the Aydin-like pestiviruses were included as test strains in the VNT, CSFV infection still could not be ruled out.

Although these novel pestiviruses are the closest known relatives of CSFV, experimental infection of pigs with Aydin/04 did not result in detectable viremia and clinical signs. Nevertheless, these ruminant pestiviruses are candidates for a switch to porcine hosts after ongoing virus evolution, which would have severe consequences for serologic diagnosis of classical swine fever, affecting control and monitoring programs performed in many parts of the world.

**Acknowledgments**

We are grateful to Alexandra Beckmann, Gabriele Müller, and Karin Ruthenberg for excellent technical assistance and to Monika Berg, Holger Mosch, and Günter Thiem for their outstanding support in the animal experiment.
This work was supported by the Directorate-General for Health and Consumers of the European Commission and by grants BE-2333/2-1 and -2 of the German Research Council (Deutsche Forschungsgemeinschaft). P.B. was supported by a Heisenberg professorship (BE-2333-1/1 and 1/2) from the German Research Council. Sample material was gained in the framework of the projects TÜBİTAK VHAG 2099 and AU BAP 2005 0810 071. The funding sources had no influence on the study design or data presented.

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References


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