preliminary results. We propose proactive vaccination of Ethiopian wolves across their distribution as an effective and urgently needed strategy to protect the species from extinction. This program should be part of an integrated disease control plan that also includes controlling disease in domestic dogs, limiting contact between dogs and wolves, and conducting policy and education interventions to reduce the size and roaming behavior of local dog populations (2).

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The animal care and use protocols for the ethical handling of Ethiopian wolves were approved by the Ethiopian Wildlife Conservation Authority and the University of Oxford’s Local Ethical Review Process (Zoology ERC; case no. ZERC040905) and adhere to the United Kingdom’s ASPA regulations (1986).

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References

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High Abundance and Genetic Variability of Atypical Porcine Pestivirus in Pigs from Europe and Asia

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Atypical porcine pestivirus (APPV) was recently reported to be associated with neurologic disorders in newborn piglets. Investigations of 1,460 serum samples of apparently healthy pigs from different parts of Europe and Asia demonstrate a geographically wide distribution of genetically highly variable APPV and high APPV genome and antibody detection rates.

Pestiviruses are highly variable RNA viruses within the family Flaviviridae. The recently discovered atypical porcine pestivirus (APPV) is capable of inducing neurologic disorder in its host, like other pathogens of this family (e.g., tick-borne encephalitis virus, Zika virus). Several recently published reports demonstrate that APPV is a prominent cause of virus-induced congenital tremor in pigs (1–4). Serum samples from healthy but viremic animals can induce birth of clinically affected offspring when experimentally transferred to sows during gestation (1,2). So far, APPV presence has been reported from the United States, some countries within Europe, and China (2,4–7).

The economic relevance of APPV-related losses in pig production remains to be determined, but estimation revealed a drop in reproductive performance of 10% in an affected farm (4). Early data from the United States and Germany suggested a relatively high abundance (2.4%–22%) of APPV genomes in apparently healthy pigs (3,6,8) that likely play an important epidemiologic role as virus carriers. We investigated APPV genome and antibody abundance in healthy pigs from different parts of Europe and Asia to provide insight into genetic diversity of this novel pathogen.

We tested 1,460 serum samples from Germany, Great Britain, Italy, Serbia, Switzerland, mainland China, and Taiwan by using an APPV-specific PCR and an indirect APPV ELISA, as previously described (3,9). The sample set comprised 369 serum samples from Germany that were previously screened for the presence of APPV genomes (3). For our study, serum samples were taken from apparently healthy pigs within the framework of national veterinary health management in concordance with national legal and ethical regulations.

For APPV genome detection, we conducted a PCR targeting the nonstructural protein (NS) 3 encoding region and confirmed specificity of amplification by gel electrophoresis (3). We detected APPV genomes in domestic pigs from all investigated regions. In total, 130 (8.9%) of the 1,460 tested samples were APPV genome positive (Figure). Genome detection rates ranged from 2.3% (2/86 samples from Great Britain) to 17.5% (35/200 samples from Italy). Moreover, we demonstrated that APPV was abundant in Asia; we detected the APPV genome in 11/219 samples (5%) from mainland China and 22/200 samples (11%) from Taiwan.

We used individual samples with high genome loads to generate amplicons in a seminested PCR and subsequently performed Sanger sequencing (FlexiRun, LGC Genomics, Berlin, Germany). We generated 20 different APPV NS3 sequences from apparently healthy pigs of all countries (sequences deposited into GenBank under accession nos. MF279213–32). Genetic differences reflect geographic origin to a low degree (Figure); genetic variability even within a country is remarkably high (e.g., Germany and Italy). Genetic analyses including sequence data obtained from samples of diseased piglets revealed no correlation of pathogenicity with certain genetic variants (Figure).

In addition to the NS3 fragments, we determined APPV complete coding sequences (CDS) from 1 sample from a healthy pig from China (deposited into GenBank under accession no. MF167292) and 2 samples (accession nos. MF167290 and MF167291) obtained from pigs during outbreaks of congenital tremors in Germany (3,9). We performed next-generation sequencing as previously described (3). The outbreak isolates from Germany were almost identical (0.2% genetic distance) and were similar to an isolate from northern Germany (accession no. LT594521). The APPV from China had a unique 93-nt deletion in the NS5A encoding region. A similar genome (97.9% identity) is lacking this deletion (Guangxi Province; accession no. KY652092). The biological relevance of the deletion remains elusive, but classical pestiviruses show a remarkable genetic tolerance in this genomic region (10).

The sequence data we obtained reveal a high genetic variability (up to 21% genetic distance), which is comparable to that of classical swine fever virus (online Technical Appendix, https://wwwnc.cdc.gov/EID/article/23/12/17-0951-Techapp1.pdf).

We applied an indirect APPV E\textsuperscript{NS} antibody ELISA, as described (9), and classified the serologic status into low (S/p\textless;0.5), intermediate (0.5\textless;S/p\textless;1.0), or highly (S/p\textless;1.0) reactive. Due to the lack of reference material and a standard assay, we could not determine test parameters (e.g., sensitivity, specificity) at this stage. Nevertheless, the ELISA was a valuable tool for detecting seroconversion in infected pigs; \textgeq;60% of the animals showed intermediate to high reactivity in the antibody ELISA (Figure), which is in line with high APPV genome detection rates. We detected similar frequencies of APPV antibody–positive samples for each region, independent of the genome detection rates (Figure). We found most of the viral genomes (\textapprox;86%) in samples with intermediate or low antibody status; few (\textapprox;14%) of the highly antibody-positive animals were viremic at the same time. This...
observation might indicate a degree of protection provided by the induced antibodies. Of the 40% of the pigs that were antibody negative, 10% were genome positive; possible explanations are that serum samples were taken either from acutely infected animals before induction of a detectable antibody response or from persistently infected animals lacking a specific humoral immune response due to a specifically acquired immunotolerance, a well-known consequence of intrauterine pestivirus infections.

Our findings indicate that the recently discovered APPV is abundant on several continents. APPV must be regarded as a pig pathogen of likely worldwide relevance.

Figure. Detection rates of APPV genome and antibodies and genetic variability in Europe and Asia. A) APPV antibody status in pigs from parts of Europe and Asia. The region of origin, the number of investigated samples, and the absolute numbers of APPV genome–positive samples in dependence on the serologic category (low, intermediate, or high APPV antibody status) are shown in the central circle. B) Phylogenetic tree based on a 400-nt fragment in the nonstructural protein 3 encoding region. We calculated genetic distances using the Kimura 2-parameter model. We performed phylogenetic analysis by the neighbor-joining method including 1,000 iterations for bootstrap analysis. Only bootstrap values ≥700 are indicated. Bold indicates sequences generated in this study; asterisks indicate sequences from piglets with congenital tremor. Accession numbers for reference sequences from GenBank are shown in brackets. Scale bar indicates nucleotide substitutions per site. APPV, atypical porcine pestivirus; CH, Switzerland; CN, China; GB, Great Britain; GER, Germany; IT, Italy; RS, Serbia; TWN, Taiwan.
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References


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Human Case of Streptococcus suis Disease, Ontario, Canada

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We report a case of Streptococcus suis human disease in Ontario, Canada, caused by a serotype 2 strain genotypically similar to those commonly isolated from pigs in North America. Initially, the isolate was misidentified as a viridans group Streptococcus. Human S. suis infections may be underdiagnosed in North America.

Streptococcus suis is a zoonotic agent responsible for both sporadic and outbreak human disease in several Asian countries (1–3). However, human S. suis infections are less frequent in Western countries, and particularly in North America (3,4). We describe a severe human S. suis infection in Ontario, Canada.

The patient, a 69-year-old male farmer, was brought to the emergency department of a rural community hospital in southwestern Ontario after being found unresponsive by his
Technical Appendix

**Technical Appendix Table.** Genetic distances (in %) of APPV obtained from different geographic regions*

<table>
<thead>
<tr>
<th>Region</th>
<th>USA-Kansas</th>
<th>China-CQ</th>
<th>Austria</th>
<th>China-GD2</th>
<th>Ger-Bavaria</th>
<th>Ger-NRW</th>
<th>USA-Iowa</th>
<th>Netherlands</th>
</tr>
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<tbody>
<tr>
<td>China-GD1</td>
<td>19.2</td>
<td>19.1</td>
<td>20.2</td>
<td>21.3</td>
<td>20.8</td>
<td>20.9</td>
<td>18.8</td>
<td>18.1</td>
</tr>
<tr>
<td></td>
<td>(19.7)</td>
<td>(20.0)</td>
<td>(19.7)</td>
<td>(19.6)</td>
<td>(19.5)</td>
<td>(19.5)</td>
<td>(20.1)</td>
<td>(19.9)</td>
</tr>
<tr>
<td>USA-Kansas</td>
<td>–</td>
<td>13.9</td>
<td>13.2</td>
<td>14.9</td>
<td>12.6</td>
<td>12.3</td>
<td>13.5</td>
<td>14.8</td>
</tr>
<tr>
<td></td>
<td></td>
<td>(13.8)</td>
<td>(13.4)</td>
<td>(13.2)</td>
<td>(13.2)</td>
<td>(13.4)</td>
<td>(13.7)</td>
<td>(13.6)</td>
</tr>
<tr>
<td>China-CQ</td>
<td>–</td>
<td>9.5</td>
<td>9.8</td>
<td>10.4</td>
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<td>10.1</td>
<td>8.9</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td>(10.6)</td>
<td>(10.4)</td>
<td>(10.4)</td>
<td>(10.7)</td>
<td>(10.9)</td>
<td>(10.6)</td>
</tr>
<tr>
<td>Austria</td>
<td>–</td>
<td>6.1</td>
<td>3.6</td>
<td>8.9</td>
<td>11.0</td>
<td>8.9</td>
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</tr>
<tr>
<td></td>
<td></td>
<td>(7.4)</td>
<td>(7.2)</td>
<td>(10.7)</td>
<td>(10.7)</td>
<td>(10.2)</td>
<td></td>
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<tr>
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<td>10.7</td>
<td>11.7</td>
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<td>(10.4)</td>
<td>(9.8)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ger-Bavaria</td>
<td>–</td>
<td>11.3</td>
<td>11.6</td>
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<td>(10.5)</td>
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<tr>
<td>Ger-NRW</td>
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<td>(9.8)</td>
</tr>
<tr>
<td>USA-Iowa</td>
<td>–</td>
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<td></td>
<td></td>
<td></td>
<td>8.9</td>
<td>(9.5)</td>
</tr>
</tbody>
</table>

*Genetic distances (Kimura 2-parameter substitution model) of a fragment in the NS3 encoding region were compared with distances among complete coding sequences (indicated by numbers in parentheses). Reference sequences were obtained from GenBank (KX950761, KR011347, KX778724, KY624591, KU041639, KU194229). CDS of APPV Ger-NRW_L277 (MF167291) and China-CQ (MF167292) were obtained within this study. Highly similar sequences harboring more than 95% genetic similarity were excluded from analysis. CQ, Chongqing; GD, Guangdong; Ger, Germany; NRW, North Rhine Westphalia.