infection is caused by a novel cluster of *Emmonsia*-like species (1); involves fungemia; appears to be associated with immunosuppression, including renal transplant (7–9) and orthotopic liver transplantation and HIV (10); and has a high case-fatality rate.

The timing of this infection raised concern for a donor-derived infection. However, we confirmed with the United Network for Organ Sharing (https://www.unos.org/) that no other organ recipients from the same donor had a similar posttransplant infection. Reported soil and rodent exposure for the patient and previous granulomatous disease identified by pretransplant chest imaging raised the possibility that his infection was a reactivation of a latent infection.

The unfamiliar mold isolated from the patient’s pleural fluid was initially identified as a contaminant, and the patient died despite favorable in vitro antifungal susceptibilities. In immunosuppressed patients with a compatible clinical syndrome, fungi isolated from a sterile site should be identified. More cases of *Emmonsia*-like infections will probably be diagnosed as laboratories use sequencing to identify uncommon fungal pathogens.

Dr. Kappagoda is a clinical assistant professor at Stanford University School of Medicine, Stanford, CA. Her research interests are neglected tropical diseases and endemic fungal infections.

References


Address for correspondence: Shanthi Kappagoda, Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, 300 Pasteur Dr, Lane Bldg, Rm L154, Stanford, CA 94305-5107, USA; email: skappago@stanford.edu

Outbreak of Legionnaire’s Disease Caused by *Legionella pneumophila* Serogroups 1 and 13

Toshiro Kuroki,¹ Junko Amemura-Maekawa,¹ Hitomi Ohya, Ichiro Furukawa, Miyuki Suzuki, Tomoka Masaoka, Kastuhiro Aikawa, Kazumi Hibi, Masatomo Morita, Ken-ichi Lee, Makoto Ohnishi, Fumiaki Kura

Author affiliations: Kanagawa Prefectural Institute of Public Health, Kanagawa, Japan (T. Kuroki, H. Ohya, I. Furukawa, M. Suzuki, T. Masaoka, K. Aikawa, K. Hibi); National Institute of Infectious Diseases, Tokyo, Japan (J. Amemura-Maekawa, M. Morita, K. Lee, M. Ohnishi, F. Kura)

DOI: http://dx.doi.org/10.3201/eid2302.161012

In Japan, hot springs and public baths are the major sources of legionellosis. In 2015, an outbreak of Legionnaires’ disease occurred among 7 patients who had visited a spa house. Laboratory investigation indicated that *L. pneumophila* serogroup 1 and 13 strains caused the outbreak and that these strains were genetically related.

¹These authors contributed equally to this article.
baths (2). Among Legionella species, L. pneumophila serogroup 1 accounts for most human infections (3). Legionellosis outbreaks caused by a combination of L. pneumophila serogroup 1 or other serogroups have rarely been reported. We report an outbreak of Legionnaires’ disease caused by L. pneumophila strains of serogroup 1 and serogroup 13.

During June 1–17, 2015, the Health Centers in Kanagawa and Shizuoka Prefectures, Japan, were notified of 7 cases of Legionella pneumonia. All patients with pneumonia were admitted to 1 of 5 hospitals. All patients were male (mean age 66.3 years), 4 had diabetes mellitus, and 1 had hepatic cirrhosis and liver cancer. Diagnosis of pneumonia at the hospitals was based on clinical presentation and immunochromatographic detection of L. pneumophila serogroup 1 antigen in urine. All patients recovered and were discharged.

In epidemiologic interviews, all 7 patients stated that they had visited a spa house in Odawara, Kanagawa, Japan, before illness onset. The latent period was not accurately determined because 5 of the 7 patients frequently visited this spa house and some patients had visited it again after illness onset. The spa house had 7 circulating systems, including filtration and heating components, and 9 baths for men.

Sputum samples from 5 urinary antigen–positive patients and environmental samples from the spa house were collected for epidemiologic investigations and cultured for Legionella at the Kanagawa Prefectural Institute of Public Health. L. pneumophila serogroup 1 was detected in sputum from 4 patients (Table), and L. pneumophila serogroups 1 and 13 were detected in sputum from 1 patient (patient 2). Baths 1–5, but not baths 6–9, contained L. pneumophila. Epidemiologic investigation and laboratory results revealed that failure to adequately chlorinate the bath water and the circulating systems resulted in colonization of Legionella at the spa house.

Pulsed-field gel electrophoresis (PFGE) comparison (4) of clinical and environmental isolates revealed that the L. pneumophila serogroup 1 strains produced 3 PFGE profiles: A and B, with a 1-band difference, and C (Table). Patient 7 was infected with 2 L. pneumophila serogroup 1 isolates possessing PFGE profiles A and B. The PFGE profile B of L. pneumophila serogroup 13 was identical to that of serogroup 1. We determined the sequence type (ST) of L. pneumophila strains (5,6) and identified 4 new STs: ST2113, ST2114, ST2115, and ST2121. ST2114 differed from ST2121 by only 1 nt in neuA and differed from ST2113 (serogroup 13) by only 2 alleles (mip and neuA), suggesting that these 3 STs are closely related and that 1 of 3 strains (ST2113, ST2114, or ST2121) may be derived from another by homologous recombination. All examined isolates lacked the lag-1 gene, a virulence-associated marker (7).

By using whole-genome sequencing, we applied core-genome multilocus sequence typing (cgMLST) with 50 genes (8) different from the 7 genes in sequence-based typing, thereby confirming the sequence-based typing data (Table). ST2114 and ST2115 isolates were each divided into 2 cgMLST profiles. cgMLST profile b differed from profile a by only 1 nt on lpg1503 and differed from profile d by 5 nt on lpg0812, near the lipopolysaccharide coding region, suggesting that the strain of profile d may be derived from another by homologous recombination. However, the remaining cgMLST profiles (c, e, and f) from strains not isolated from patients differed from profile b by 30, 42, and 43 alleles, respectively.

Multiple L. pneumophila strains with different genetic characteristics exist in the environment and pose infection risks (9). Among the strains studied, dual infections with L. pneumophila serogroup 1 and serogroup 13 strains (patient 2) and L. pneumophila serogroup 1 strains with different

<table>
<thead>
<tr>
<th>Serogroup</th>
<th>PGFE profile†</th>
<th>ST‡</th>
<th>Sample source</th>
<th>cgMLST profile§</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>A</td>
<td>ST2114</td>
<td>6, 10, 21, 3, 17, 14, 9</td>
<td>Patients 1, 7</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bath 1 (bath water)</td>
<td>a, b</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bath 2 (spout swab)</td>
<td>b</td>
</tr>
<tr>
<td>1</td>
<td>A</td>
<td>ST2114</td>
<td>6, 10, 21, 3, 17, 14, 9</td>
<td>Bath 1 (bath water)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bath 1 (bath water)</td>
<td>a</td>
</tr>
<tr>
<td>1</td>
<td>B</td>
<td>ST2114</td>
<td>6, 10, 21, 3, 17, 14, 9</td>
<td>Patients 2, 5, 7</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bath 1 (bathub swab)</td>
<td>b, b, b</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bath 2 (bath water)</td>
<td>b</td>
</tr>
<tr>
<td>1</td>
<td>C</td>
<td>ST1447</td>
<td>6, 10, 20, 13, 9, 4, 11</td>
<td>Bath 3 (hair trap debris)</td>
</tr>
<tr>
<td>13</td>
<td>B</td>
<td>ST2113</td>
<td>6, 10, 21, 10, 17, 14, 209</td>
<td>Patient 2</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bath 1 (bath water)</td>
<td>b</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bath 2 (bath water)</td>
<td>d</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bath 2 (spout swab)</td>
<td>Not done</td>
</tr>
<tr>
<td>10</td>
<td>D</td>
<td>ST2115</td>
<td>7, 10, 17, 3, 13, 14, 207</td>
<td>Bath 4 (spout swab)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Bath 5 (bathub swab)</td>
<td>e</td>
</tr>
</tbody>
</table>

*cgMLST, core genome multilocus sequence typing; PFGE, pulsed-field gel electrophoresis; ST, sequence type.
†Profiles A and B were obtained from clinical and environmental samples. Profiles C and D were obtained from environmental samples only.
‡New STs from this study were assigned ST2113, ST2114, ST2115, and ST2121.
§Each profile letter indicates a tentative cgMLST profile of 1 strain.
¶Concentrations of L. pneumophila in bath water were 800 CFU/L in bath 1, and 1,100 CFU/L in bath 2.
genomic subtypes (patient 7) were detected. Results from 3 genetic methods revealed that \textit{L. pneumophila} serogroup 1 and 13 strains are closely related, although the serogroups differ. Results of this study were consistent with the hypothesis that multiple infections are more likely with less virulent strains and more likely in persons with medical conditions predisposing them to Legionnaires’ disease (10).

Our study of this outbreak suggests that the spa house was colonized by several \textit{L. pneumophila} strains that were genetically related despite belonging to different serogroups and that 2 strains caused infection. Further analysis of the divergence of outbreak strains in genomes related to \textit{Legionella} serogroup and sequence types is ongoing. This analysis clarifies the in-depth genetic relations among \textit{L. pneumophila} strains, such as recombination sites and periods required for divergence. We recommend that the spa house provide high quality management and effective infection control practices according to an infection control manual (e.g., completion of documentation relating to infection control practices and training of employees) and that customers be aware of the sanitary status of spa houses.

Acknowledgments

We appreciate the help of the officers at Odawara Health and Welfare Center, who provided information on epidemiological studies.

This work was partly supported by Health and Labour Sciences Research (grant H25-kenki-009 to F. K.).

Dr. Kuroki is a director of the Department of Microbiology at Kanagawa Prefectural Institute of Public Health, Kanagawa, Japan. His research interests include epidemiology of protozoan and bacterial infectious diseases.

References


Address for correspondence: Toshiro Kuroki, Department of Microbiology, Kanagawa Prefectural Institute of Public Health, 1-3-1 Shimomachiya, Chigasaki, Kanagawa, 253-0087, Japan; email: kuroki.gcg3@pref.kanagawa.jp

Diphyllobothrium nihonkaiense Tapeworm Larvae in Salmon from North America

Roman Kuchta, Mikuláš Oros, Jayde Ferguson, Tomáš Scholz

Author affiliations: Biology Centre of the Academy of Sciences of the Czech Republic, České Budějovice, Czech Republic (R. Kuchta, T. Scholz); Slovak Academy of Sciences, Košice, Slovak Republic (M. Oros); State of Alaska Department of Fish and Game, Anchorage, Alaska, USA (J. Ferguson)

DOI: http://dx.doi.org/10.3201/eid2302.161026

Diphyllobothriasis is reemerging because of global importation and increased popularity of eating raw fish. We detected \textit{Diphyllobothrium nihonkaiense} ploerocercoids in the musculature of wild pink salmon (\textit{Oncorhynchus gorbuscha}) from Alaska, USA. Therefore, salmon from the American and Asian Pacific coasts and elsewhere pose potential dangers for persons who eat these fish raw.