

# Significant Decrease in Pertactin–Deficient *Bordetella pertussis* Isolates, Japan

## Technical Appendix

Technical Appendix Table 1. Characteristics of *Bordetella pertussis* isolates, Japan, 2005–2016\*

Isolate	Isolation year	Patient age	Vaccine status	Origin (district)	Pertactin production	<i>prn</i> gene	<i>prn</i> allele	MLVA type
BP285	2005	2 mo	0	Kanto	–	ΔSS	1	186
BP291	2005	1 y	3	Chubu	–	ΔSS	1	186
BP292	2005	1 mo	0	Chubu	–	ΔSS	1	226
BP293	2005	4 mo	1	Chubu	–	ΔSS	1	186
BP299	2005	5 mo	0	Kyusyu	–	ΔSS	1	186
BP302	2005	1 mo	0	Kinki	–	1598–	1	226
						1599::IS481		
BP307	2006	2 mo	0	Chubu	–	1598–	1	186
						1599::IS481		
BP310	2006	11 y	Unknown	Tohoku	–	ΔSS	1	194
BP313	2006	58 y	Unknown	Kanto	–	1598–	1	186
						1599::IS481		
BP322	2006	1 mo	0	Kanto	–	ΔSS	1	186
BP323	2006	1 y	Unknown	Kanto	–	ΔSS	1	186
BP332	2006	35 y	Unknown	Kinki	–	ΔSS	1	194
BP333	2006	30 y	Unknown	Kinki	–	ΔSS	1	194
BP334	2006	0 y	0	Kinki	–	ΔSS	1	194
BP314	2007	3 mo	0	Kanto	–	ΔSS	1	186
BP318	2007	1 y	0	Kanto	–	ΔSS	1	186
BP466	2008	33 y	Unknown	Kanto	–	ΔSS	1	186
BP469	2008	8 y	Unknown	Kanto	–	ΔSS	1	186
BP346	2009	2 mo	0	Chubu	–	ΔSS	1	186
BP351	2009	4 mo	Unknown	Shikoku	–	ΔSS	1	186
BP458	2009	70 y	Unknown	Kanto	–	ΔSS	1	186
BP459	2009	5 mo	Unknown	Kanto	–	ΔSS	1	186
BP460	2009	6 mo	Unknown	Kanto	–	ΔSS	1	186
BP357	2010	12 y	Unknown	Kanto	–	ΔSS	1	186
BP358	2010	10 y	Unknown	Kanto	–	ΔSS	1	186
BP359	2010	Unknown	Unknown	Kanto	–	ΔSS	1	186
BP360	2010	Unknown	Unknown	Kanto	–	ΔSS	1	186
BP361	2010	10 y	Unknown	Kanto	–	ΔSS	1	186
BP363	2010	Unknown	Unknown	Kanto	–	ΔSS	1	186
BP365	2010	Unknown	Unknown	Kanto	–	ΔSS	1	186
BP366	2010	6 y	Unknown	Kanto	–	ΔSS	1	186
BP378	2011	21 y	Unknown	Kyusyu	–	ΔSS	1	186
BP390	2011	4 y	Unknown	Kinki	–	ΔSS	1	316
BP394	2011	5 mo	0	Kyusyu	–	<i>prn</i>	2	27
						(transcriptional down–regulation)		
BP395	2011	6 mo	0	Kinki	–	ΔSS	1	186
BP397	2011	7 y	Unknown	Kinki	–	ΔSS	1	186
BP398	2011	4 mo	0	Kinki	–	ΔSS	1	225
BP400	2011	5 y	Unknown	Kinki	–	ΔSS	1	186
BP401	2011	9 y	4	Kinki	–	ΔSS	1	314
BP403	2011	2 mo	0	Kinki	–	ΔSS	1	315
BP417	2011	35 y	4	Kanto	–	ΔSS	1	186
BP437	2011	1 mo	0	Kinki	–	ΔSS	1	186
BP438	2011	2 mo	0	Kinki	–	ΔSS	1	186
BP451	2011	3 mo	0	Kinki	–	ΔSS	1	186
BP452	2011	7 mo	Unknown	Kinki	–	ΔSS	1	186
BP410	2012	1 mo	0	Tohoku	–	ΔSS	1	186
BP412	2012	26 y	Unknown	Shikoku	–	ΔSS	1	186

Isolate	Isolation year	Patient age	Vaccine status	Origin (district)	Pertactin production	<i>prn</i> gene	<i>prn</i> allele	MLVA type
BP416	2012	3 mo	0	Kanto	-	ΔSS	1	186
BP418	2012	5 y	4	Kanto	-	ΔSS	1	186
BP440	2012	10 y	4	Kinki	-	ΔSS	1	186
BP442	2012	2 y	0	Kinki	-	ΔSS	1	316
BP447	2012	28 y	Unknown	Kyusyu	-	ΔSS	1	186
BP478	2012	5 y	Unknown	Kinki	-	ΔSS	1	186
BP480	2013	2 mo	0	Kinki	-	ΔSS	1	224
BP481	2013	1 mo	0	Kinki	-	ΔSS	1	186
BP510	2015	1 mo	0	Kinki	-	ΔSS	1	186
BP533	2015	1 mo	0	Kinki	-	245-246::IS481	2	27
BP535	2015	3 mo	0	Kyusyu	-	ΔSS	1	186
BP550	2016	7 y	4	Kanto	-	ΔSS	1	186
BP283	2005	9 y	Unknown	Kanto	+	<i>prn</i>	2	27
BP284	2005	1 mo	0	Chubu	+	<i>prn</i>	1	186
BP289	2005	6 mo	0	Kinki	+	<i>prn</i>	2	27
BP290	2005	1 y	0	Chugoku	+	<i>prn</i>	2	27
BP294	2005	10 mo	1	Kyusyu	+	<i>prn</i>	2	27
BP296	2005	6 y	0	Kyusyu	+	<i>prn</i>	2	27
BP297	2005	3 mo	0	Chugoku	+	<i>prn</i>	2	22
BP298	2005	3 mo	0	Kinki	+	<i>prn</i>	2	69
BP300	2005	2 mo	0	Kyusyu	+	<i>prn</i>	2	26
BP301	2005	11 mo	0	Kyusyu	+	<i>prn</i>	2	27
BP303	2005	1 mo	0	Chugoku	+	<i>prn</i>	1	224
BP306	2006	10 y	4	Chubu	+	<i>prn</i>	2	69
BP311	2006	1 y	Unknown	Tohoku	+	<i>prn</i>	1	187
BP335	2006	0 y	Unknown	Kinki	+	<i>prn</i>	1	186
BP312	2007	4 mo	0	Kanto	+	<i>prn</i>	2	27
BP316	2007	53 y	Unknown	Tohoku	+	<i>prn</i>	1	234
BP317	2007	11 y	Unknown	Tohoku	+	<i>prn</i>	1	234
BP324	2007	65 y	Unknown	Kanto	+	<i>prn</i>	2	27
BP327	2007	10 y	Unknown	Shikoku	+	<i>prn</i>	1	229
BP330	2007	6 mo	0	Chubu	+	<i>prn</i>	2	27
BP336	2007	0 y	2	Kinki	+	<i>prn</i>	2	27
BP337	2007	43 y	Unknown	Kinki	+	<i>prn</i>	2	27
BP343	2007	0 y	Unknown	Kanto	+	<i>prn</i>	2	95
BP331	2008	1 y	0	Chubu	+	<i>prn</i>	2	27
BP338	2008	11 y	2	Kinki	+	<i>prn</i>	2	27
BP339	2008	8 y	Unknown	Kinki	+	<i>prn</i>	2	27
BP340	2008	29 y	Unknown	Kinki	+	<i>prn</i>	2	27
BP341	2008	6 y	Unknown	Kinki	+	<i>prn</i>	1	187
BP342	2008	0 y	Unknown	Kinki	+	<i>prn</i>	2	27
BP344	2008	5 y	Unknown	Chubu	+	<i>prn</i>	2	27
BP345	2008	4 y	Unknown	Chubu	+	<i>prn</i>	2	27
BP464	2008	3 mo	0	Kanto	+	<i>prn</i>	2	27
BP465	2008	27 y	Unknown	Kanto	+	<i>prn</i>	2	27
BP467	2008	8 mo	Unknown	Kanto	+	<i>prn</i>	2	27
BP347	2009	3 y	0	Shikoku	+	<i>prn</i>	9	27
BP348	2009	41 y	Unknown	Chubu	+	<i>prn</i>	2	27
BP349	2009	2 mo	0	Kyusyu	+	<i>prn</i>	1	186
BP350	2009	13 y	4	Chubu	+	<i>prn</i>	1	186
BP352	2009	1 mo	0	Shikoku	+	<i>prn</i>	1	34
BP353	2009	12 y	Unknown	Kyusyu	+	<i>prn</i>	2	22
BP354	2009	1 mo	0	Kyusyu	+	<i>prn</i>	1	311
BP355	2009	6 mo	Unknown	Kinki	+	<i>prn</i>	1	27
BP356	2010	2 mo	0	Kinki	+	<i>prn</i>	ND	221
BP362	2010	3 y	Unknown	Kyusyu	+	<i>prn</i>	2	27
BP364	2010	8 y	Unknown	Kanto	+	<i>prn</i>	2	27
BP367	2010	9 y	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP368	2010	5 mo	Unknown	Kyusyu	+	<i>prn</i>	1	186
BP369	2010	15 y	Unknown	Kyusyu	+	<i>prn</i>	2	27
BP371	2010	1 mo	0	Kyusyu	+	<i>prn</i>	1	186
BP462	2010	6 mo	Unknown	Kanto	+	<i>prn</i>	2	27
BP470	2010	1 mo	0	Kanto	+	<i>prn</i>	2	27
BP376	2011	2 mo	0	Chubu	+	<i>prn</i>	2	27
BP377	2011	1 mo	0	Chugoku	+	<i>prn</i>	2	27
BP380	2011	11 y	Unknown	Kyusyu	+	<i>prn</i>	2	27
BP388	2011	3 y	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP389	2011	3 mo	1	Kanto	+	<i>prn</i>	2	27

Isolate	Isolation year	Patient age	Vaccine status	Origin (district)	Pertactin production	<i>prn</i> gene	<i>prn</i> allele	MLVA type
BP391	2011	5 y	Unknown	Kanto	+	<i>prn</i>	2	27
BP392	2011	4 mo	Unknown	Kanto	+	<i>prn</i>	ND	27
BP393	2011	11 mo	Unknown	Kyusyu	+	<i>prn</i>	2	27
BP396	2011	1 mo	0	Kinki	+	<i>prn</i>	1	187
BP399	2011	2 y	Unknown	Kinki	+	<i>prn</i>	ND	27
BP402	2011	1 mo	0	Kinki	+	<i>prn</i>	ND	27
BP404	2011	10 y	Unknown	Kyusyu	+	<i>prn</i>	2	27
BP405	2011	14 y	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP406	2011	1 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP407	2011	6 mo	Unknown	Kyusyu	+	<i>prn</i>	2	27
BP430	2011	1 y	3	Kinki	+	<i>prn</i>	2	27
BP431	2011	2 mo	0	Kinki	+	<i>prn</i>	9	27
BP432	2011	8 y	0	Kinki	+	<i>prn</i>	2	27
BP433	2011	5 mo	0	Kinki	+	<i>prn</i>	2	27
BP436	2011	4 mo	0	Kinki	+	<i>prn</i>	ND	27
BP453	2011	6 mo	0	Kinki	+	<i>prn</i>	1	186
BP463	2011	3 mo	1	Kanto	+	<i>prn</i>	2	27
BP471	2011	2 mo	0	Kanto	+	<i>prn</i>	2	27
BP493	2011	Unknown	Unknown	Kanto	+	<i>prn</i>	ND	27
BP494	2011	0 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP501	2011	Unknown	Unknown	Kinki	+	<i>prn</i>	ND	27
BP408	2012	5 mo	Unknown	Kanto	+	<i>prn</i>	ND	27
BP409	2012	4 mo	Unknown	Tohoku	+	<i>prn</i>	2	27
BP411	2012	17 y	Unknown	Tohoku	+	<i>prn</i>	ND	27
BP413	2012	10 y	4	Kanto	+	<i>prn</i>	2	27
BP414	2012	1 mo	0	Chubu	+	<i>prn</i>	2	27
BP415	2012	8 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP420	2012	4 mo	0	Kanto	+	<i>prn</i>	2	27
BP421	2012	5 mo	Unknown	Kanto	+	<i>prn</i>	ND	27
BP422	2012	8 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP423	2012	8 y	Unknown	Kanto	+	<i>prn</i>	2	27
BP424	2012	9 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP425	2012	7 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP426	2012	8 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP427	2012	4 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP428	2012	10 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP429	2012	12 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP434	2012	7 y	Unknown	Kanto	+	<i>prn</i>	2	27
BP435	2012	7 y	Unknown	Kanto	+	<i>prn</i>	ND	27
BP439	2012	3 mo	Unknown	Kinki	+	<i>prn</i>	2	27
BP441	2012	3 mo	Unknown	Kinki	+	<i>prn</i>	ND	186
BP443	2012	3 mo	Unknown	Kinki	+	<i>prn</i>	2	27
BP444	2012	1 mo	0	Tohoku	+	<i>prn</i>	3	27
BP445	2012	1 mo	0	Tohoku	+	<i>prn</i>	2	27
BP446	2012	2 mo	0	Tohoku	+	<i>prn</i>	2	27
BP448	2012	2 mo	0	Kinki	+	<i>prn</i>	2	27
BP449	2012	4 mo	2	Kinki	+	<i>prn</i>	2	27
BP450	2012	1 mo	0	Kinki	+	<i>prn</i>	2	27
BP454	2012	11 y	Unknown	Tohoku	+	<i>prn</i>	9	27
BP455	2012	1 mo	Unknown	Tohoku	+	<i>prn</i>	2	27
BP456	2012	0 mo	0	Tohoku	+	<i>prn</i>	2	27
BP472	2012	1 mo	0	Kanto	+	<i>prn</i>	2	27
BP477	2012	2 mo	0	Kinki	+	<i>prn</i>	ND	27
BP495	2012	0 y	Unknown	Tohoku	+	<i>prn</i>	ND	27
BP496	2012	Unknown	Unknown	Kanto	+	<i>prn</i>	ND	27
BP513	2012	3 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP514	2012	4 mo	≥1	Kyusyu	+	<i>prn</i>	ND	27
BP515	2012	5 mo	≥1	Kyusyu	+	<i>prn</i>	ND	26
BP538	2012	0 mo	0	Tohoku	+	<i>prn</i>	ND	27
BP473	2013	29 y	≥1	Shikoku	+	<i>prn</i>	ND	27
BP474	2013	31 y	Unknown	Kyusyu	+	<i>prn</i>	ND	186
BP475	2013	8 y	Unknown	Kyusyu	+	<i>prn</i>	ND	28
BP479	2013	10 y	0	Kinki	+	<i>prn</i>	ND	27
BP482	2013	4 mo	Unknown	Kanto	+	<i>prn</i>	ND	27
BP483	2013	52 y	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP497	2013	3 mo	Unknown	Kanto	+	<i>prn</i>	ND	27
BP498	2013	5 y	Unknown	Tohoku	+	<i>prn</i>	ND	27
BP516	2013	2 mo	0	Kyusyu	+	<i>prn</i>	ND	27

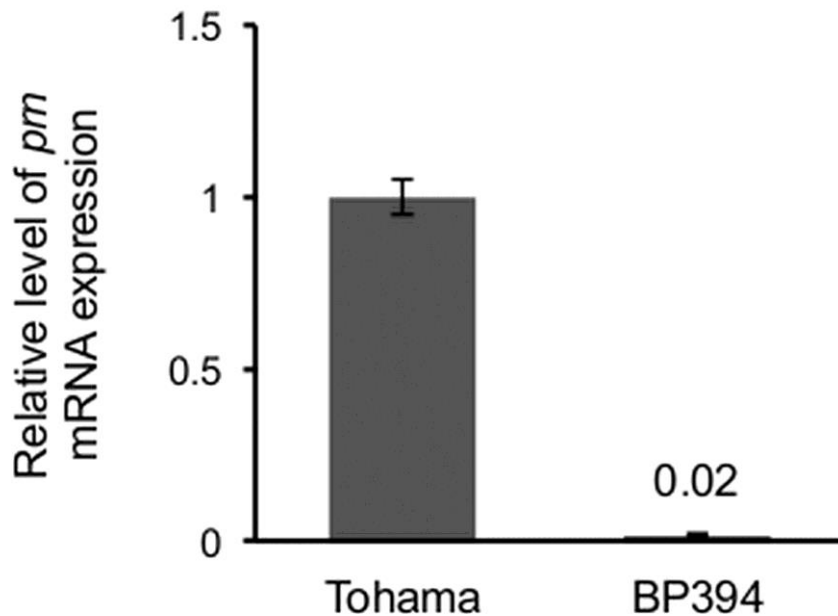
Isolate	Isolation year	Patient age	Vaccine status	Origin (district)	Pertactin production	<i>prn</i> gene	<i>prn</i> allele	MLVA type
BP484	2014	1 mo	0	Kyusyu	+	<i>prn</i>	ND	28
BP485	2014	13 y	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP486	2014	1 mo	0	Shikoku	+	<i>prn</i>	ND	27
BP487	2014	1 mo	0	Kanto	+	<i>prn</i>	ND	27
BP488	2014	1 mo	0	Chugoku	+	<i>prn</i>	ND	27
BP489	2014	5 y	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP490	2014	3 mo	0	Chugoku	+	<i>prn</i>	ND	27
BP491	2014	8 y	4	Kinki	+	<i>prn</i>	ND	27
BP499	2014	3 mo	Unknown	Kinki	+	<i>prn</i>	ND	27
BP500	2014	30 y	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP502	2014	1 mo	0	Kanto	+	<i>prn</i>	ND	27
BP503	2014	1 mo	0	Kanto	+	<i>prn</i>	ND	27
BP505	2014	36 y	Unknown	Kinki	+	<i>prn</i>	ND	27
BP506	2014	10 y	Unknown	Hokkaidou	+	<i>prn</i>	ND	27
BP517	2014	1 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP518	2014	1 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP519	2014	8 y	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP520	2014	13 y	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP521	2014	6 y	≥1	Kyusyu	+	<i>prn</i>	ND	27
BP522	2014	6 y	≥1	Kyusyu	+	<i>prn</i>	ND	27
BP523	2014	8 y	≥1	Kyusyu	+	<i>prn</i>	ND	27
BP524	2014	4 mo	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP525	2014	1 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP526	2014	2 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP527	2014	2 mo	0	Kyusyu	+	<i>prn</i>	ND	29
BP532	2014	4 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP507	2015	3 mo	Unknown	Kinki	+	<i>prn</i>	ND	78
BP508	2015	1 mo	0	Chubu	+	<i>prn</i>	ND	27
BP509	2015	32 y	Unknown	Kyusyu	+	<i>prn</i>	ND	29
BP511	2015	2 mo	0	Kyusyu	+	<i>prn</i>	ND	96
BP512	2015	8 y	4	Kinki	+	<i>prn</i>	ND	27
BP528	2015	1 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP529	2015	2 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP530	2015	3 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP531	2015	3 mo	Unknown	Kyusyu	+	<i>prn</i>	ND	27
BP534	2015	0 mo	0	Kyusyu	+	<i>prn</i>	ND	27
BP536	2015	3 mo	1	Shikoku	+	<i>prn</i>	ND	27
BP537	2015	2 mo	0	Kinki	+	<i>prn</i>	ND	31
BP539	2015	1 mo	0	Tohoku	+	<i>prn</i>	ND	32
BP540	2015	1 mo	0	Tohoku	+	<i>prn</i>	ND	27
BP541	2015	6 mo	0	Kinki	+	<i>prn</i>	ND	27
BP551	2015	12 y	3	Chubu	+	<i>prn</i>	2	27
BP542	2016	9 y	4	Hokkaidou	+	<i>prn</i>	2	27
BP543	2016	10 y	Unknown	Kinki	+	<i>prn</i>	2	27
BP544	2016	1 mo	0	Kyusyu	+	<i>prn</i>	1	34
BP545	2016	13 y	4	Chubu	+	<i>prn</i>	2	27
BP549	2016	9 y	4	Hokkaidou	+	<i>prn</i>	2	27
BP555	2016	6 mo	3	Chubu	+	<i>prn</i>	2	28
BP556	2016	8 y	4	Kanto	+	<i>prn</i>	2	27

\*, negative; +, positive; ND, not determined; MLVA, multilocus variable number tandem repeat analysis.

Technical Appendix Table 2. Primers used in this study of *Bordetella pertussis* isolates, Japan, 2008–2016\*

Designation	Primer name	Sequence (5' to 3')
Conventional PCR	SS-defect-F5	CTCTGTCACGCATTGACAAC
	SS-outerR	CTCGGCCGCGGGATTTTCTA
	IS481-combine-F	GCAGACGCCACTAGGTGTGA
	IS481-combine-R	AAAGGTCGCCGCGCTGCCTA
qRT PCR	qprn-F	ATCGTCAAGACCGGTGAGCG
	qprn-R	CTGACGGCCGCTTACCTTGA
	qrecA-F	CCAATGTGGTCGACAAGTCC
	qrecA-R	ATGGCCATTCCTTGTGCTC

\*qRT–PCR, quantitative reverse transcription PCR.



Technical Appendix Figure. Lack of *pm* transcript expression in pertactin–deficient *Bordetella pertussis* isolate BP394. The pertactin–deficient isolate was cultured for 3 days on Bordet–Gengou agar. Total RNA was isolated by using the RNeasy Mini Kit (QIAGEN, Hilden, Germany) and treated with RNase–Free DNase (QIAGEN) to degrade contaminating DNA. Total RNA (0.1  $\mu$ g) was reverse–transcribed into cDNA by using the PrimeScript RT Master Mix with random hexamers (TaKaRa Bio, Inc., Shiga, Japan). Relative levels of *pm* and *recA* transcripts were determined using SYBR Premix Ex *TaqII* (TaKaRa Bio) with the ABI PRISM 7500 Fast Real–Time PCR System (Applied Biosystems, Waltham, MA, USA). Quantitative reverse transcription–PCR (qRT–PCR) conditions were 10 s at 95°C, followed by 40 cycles of 95°C for 3 s, and 60°C for 30 s. Primer sets qprn–F/qprn–R and qrecA–F/qrecA–R were used for *pm* and *recA* amplification, respectively (Technical Appendix Table 2). The relative *pm* transcript level was calculated using the  $\Delta\Delta$ Ct method and was normalized to that of *recA*. The *recA* transcript was used as an internal control for each sample. Data are presented as -fold- changes in expression compared with

those observed in *B. pertussis* strain Tohama. The mean  $\pm$  SDs of results obtained from 3 separate experiments are shown.