### Acknowledgments

We thank the National Research Council of Thailand and the Agricultural Research Development Agency for financial support (PRP6005020450) and the Center for Advanced Studies for Agriculture and Food, Kasetsart University Institute for Advanced Studies. We also thank Chulalongkorn University for financial support to the Center of Excellence for Emerging and Re-emerging Infectious Diseases in Animals, the Thailand Research Fund for financial support to the Thailand Research Fund Senior Scholar (to A.A.) (RTA5780006), and the National Science and Technology Development Agency (P-15-50004).

Dr. Surachetpong is an assistant professor at the Faculty of Veterinary Medicine, Kasetsart University, Bangkok. His research interests are emerging viruses and immunology in aquatic animals.

### References

- Crane M, Hyatt A. Viruses of fish: an overview of significant pathogens. Viruses. 2011;3:2025–46. http://dx.doi.org/10.3390/ v3112025
- Keawcharoen J, Techangamsuwan S, Ponpornpisit A, Lombardini ED, Patchimasiri T, Pirarat N. Genetic characterization of a betanodavirus isolated from a clinical disease outbreak in farm-raised tilapia *Oreochromis niloticus* (L.) in Thailand. J Fish Dis. 2015;38:49–54. http://dx.doi.org/10.1111/jfd.12200
- Shlapobersky M, Sinyakov MS, Katzenellenbogen M, Sarid R, Don J, Avtalion RR. Viral encephalitis of tilapia larvae: primary characterization of a novel herpes-like virus. Virology. 2010;399:239–47. http://dx.doi.org/10.1016/j.virol.2010.01.001
- Eyngor M, Zamostiano R, Kembou Tsofack JE, Berkowitz A, Bercovier H, Tinman S, et al. Identification of a novel RNA virus lethal to tilapia. J Clin Microbiol. 2014;52:4137–46. http://dx.doi.org/10.1128/JCM.00827-14
- Ferguson HW, Kabuusu R, Beltran S, Reyes E, Lince JA, del Pozo J. Syncytial hepatitis of farmed tilapia, *Oreochromis niloticus* (L.): a case report. J Fish Dis. 2014;37:583–9. http://dx.doi.org/10.1111/jfd.12142
- Kurita J, Nakajima K, Hirono I, Aoki T. Polymerase chain reaction (PCR) amplification of DNA of red seam bream iridovirus (RSIV). Fish Pathology. 1998;33:17–23. http://dx.doi.org/10.3147/ jsfp.33.17
- Bacharach E, Mishra N, Briese T, Zody MC, Kembou Tsofack JE, Zamostiano R, et al. Characterization of a novel orthomyxo-like virus causing mass die-offs of tilapia. MBio. 2016;7:e00431–16. http://dx.doi.org/10.1128/mBio.00431-16
- Müller R, Poch O, Delarue M, Bishop DH, Bouloy M. Rift Valley fever virus L segment: correction of the sequence and possible functional role of newly identified regions conserved in RNAdependent polymerases. J Gen Virol. 1994;75:1345–52. http://dx.doi.org/10.1099/0022-1317-75-6-1345
- Poch O, Sauvaget I, Delarue M, Tordo N. Identification of four conserved motifs among the RNA-dependent polymerase encoding elements. EMBO J. 1989;8:3867–74.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. Mol Biol Evol. 2013;30:2725–9. http://dx.doi.org/10.1093/molbev/mst197

Address for correspondence: Win Surachetpong, Faculty of Veterinary Medicine, Kasetsart University, Bangkok, Thailand 10900; email: fvetwsp@ku.ac.th

# Endemic Hantavirus in Field Voles, Northern England

### Anna G. Thomason, Michael Begon, Janette E. Bradley, Steve Paterson, Joseph A. Jackson

DOI: https://dx.doi.org/10.3201/eid2306.161607

Author affiliations: University of Salford, Salford, UK (A.G. Thomason, J.A. Jackson); University of Liverpool, Liverpool, UK (M. Begon, S. Paterson); University of Nottingham, Nottingham, UK (J.E. Bradley)

We report a PCR survey of hantavirus infection in an extensive field vole (*Microtus agrestis*) population present in the Kielder Forest, northern England. A Tatenale virus–like lineage was frequently detected ( $\approx$ 17% prevalence) in liver tissue. Lineages genetically similar to Tatenale virus are likely to be endemic in northern England.

**R**ecently a new vole-associated hantavirus (Tatenale virus) was discovered in northern England (1), but only from an individual *Microtus agrestis* field vole. Previously only hantaviruses from murine-associated lineages (Seoul virus [SEOV] and SEOV-like viruses) had been reported in the United Kingdom, despite the abundance of potential vole hosts in the mainland United Kingdom and the endemicity of vole-associated hantavirus lineages (Puumala virus [PUUV] and Tula virus) in mainland Europe (2). Here we present data suggesting that the Tatenale virus lineage is endemic in northern England.

European hantaviruses are of public health significance because they are a causative agent of hemorrhagic fever with renal syndrome (HFRS). In the United Kingdom, HFRS cases have primarily been attributed to SEOV-like viruses on the basis of serologic tests. SEOV antibodies have been detected in both humans and Norway rats (*Rattus norvegicus*) in Northern Ireland and Yorkshire (3,4), and seropositivity in humans correlates with domestic or occupational exposure to rats (3,5). However, in the United Kingdom, HFRS cases with serologic cross-reactivity to PUUV (3), which might share antigenic determinants with Tatenale virus, have occurred.

To investigate the endemicity of hantavirus in field voles in the United Kingdom, we surveyed the extensive field vole population in the Kielder Forest, Northumberland ( $\approx$ 230 km distant from the locality where Tatenale virus was discovered). All sampled sites were grassy, clear-cut areas (adjacent to forest stands) where field voles were prevalent. Fieldwork was approved by the University of Liverpool Animal Welfare Committee and conducted subject to UK home office project license PPL 70 8210. Following the

#### RESEARCH LETTERS

capture and processing of animals as previously described (6), we extracted viral RNA from 48 livers using a QIAamp Viral RNA Mini Kit (QIAGEN, Manchester, UK) and converted to cDNA using a High-Capacity RNA-to-cDNA Kit (Applied Biosystems, Warrington, UK). Hantaviruses were detected by PCR amplification of a fragment of the genomic L segment encoding RNA polymerase, following the strategy outlined by Klempa et al. (7).

PCR-positive results were recorded for 16.7% of voles (8/48) in total; positive voles were recorded at 3 of the 5 sampled sites and throughout the survey period, March–September 2015 (Figure, panel A; online Technical Appendix Table, https://wwwnc.cdc.gov/EID/article/23/6/16-1607-Techapp1.pdf). Three positive samples from different individual voles were sequenced (in both directions from

independent replicate PCRs) by Sanger sequencing (Source BioScience, Rochdale, UK). The 380-bp sequence was identical in all 3 positive vole samples, with the exception of a single nucleotide polymorphism at position 145 (adenine, 2 voles; guanine, 1 vole; GenBank accession nos. KY751731, KY751732). The recovered sequences were similar to but divergent from Tatenale virus (86.0%–86.3% identity at the nucleotide level and 95.9%–96.7% identity at the amino acid level; online Technical Appendix Figures 1, 2). Phylogenetic analysis of the L segment demonstrated this level of divergence was comparable to the divergence among many western European lineages of PUUV (Figure, panel B; online Technical Appendix Figure 3).

Taken together with the original report, these data are sufficient to suggest that Tatenale-like hantavirus



**Figure**. Investigation of Tatenale virus–like hantavirus lineages in the United Kingdom. A) Locations on the UK mainland where Tatenale virus–like hantavirus lineages have been found: Tattenhall, Cheshire, the site where Tatenale virus was discovered (*1*); and Kielder Forest, Northumberland. Kielder Forest sample sites are indicated in the inset (Geordies Knowe [GRD]: 55°11′1.61″N, 2°35′3.05″W; Cheese Sike [CHE]: 55°13′8.39″N, 2°32′26.50″W; Scaup [SCP]: 55°15′44.18″N, 2°32′41.05″W). B) Bayesian phylogenetic tree for the hantavirus genomic large segment (318-bp fragment with no missing data), showing relationships among Tatenale virus-like lineages and other relevant lineages. Bold represents the Tatenale virus–like lineage found in this study; either sequence reported here produces the same tree. Phylogenetic analysis was conducted by using a general time reversible plus gamma plus invariant sites model within MrBayes (*8*) software using Markov chain Monte Carlo chain lengths of 1 million and a strict clock. We estimated substitution models using MrModelTestV2 (9). Sequences are represented by the taxonomic names, strain (if >1 is included), and GenBank accession numbers. The tree is drawn to scale with node values representing the posterior probabilities. Scale bar represents nucleotide substitutions per site. ADLV, Adler virus; CATV, Catacamas virus; HOKV, Hokkaido virus; KBRV, Khabarovsk virus; LNV, Laguna Negra virus; PHV, Prospect Hill virus; PUUV, Puumala virus; TULV, Tula virus.

lineages are widespread and common in northern England. Furthermore, the considerable sequence divergence between samples in Cheshire and Northumberland is consistent with a long-standing endemicity in northern England. Given that PUUV has never been recorded in the United Kingdom (2,10), the possibility should be considered that a Tatenale-like virus could have been responsible for some of the HFRS cases that have occurred here. More studies are needed to confirm whether other common rodents in the United Kingdom are hosts for this virus and to further characterize its phyletic relationships and zoonotic potential. Cross-reactivity of the sera from Tatenale-like virus-infected individuals to antigens of other relevant hantaviruses should be determined to inform future serologic surveys and the diagnosis of human HFRS cases.

#### Acknowledgments

We thank Rebecca Barber, Lukasz Lukomski, Steve Price, and Ann Lowe for their assistance.

This work was funded by the Natural Environment Research Council, United Kingdom (research grant NE/L013517/2).

Ms. Thomason is a doctoral student at the School of Environment and Life Sciences at the University of Salford, Manchester, United Kingdom. Her interests are in the ecological dynamics of infectious disease in wildlife.

#### References

- Pounder KC, Begon M, Sironen T, Henttonen H, Watts PC, Voutilainen L, et al. Novel hantavirus in wildlife, United Kingdom. Emerg Infect Dis. 2013;19:673–5. http://dx.doi.org/ 10.3201/eid1904.121057
- Bennett E, Clement J, Sansom P, Hall I, Leach S, Medlock JM. Environmental and ecological potential for enzootic cycles of Puumala hantavirus in Great Britain. Epidemiol Infect. 2010;138:91–8. http://dx.doi.org/10.1017/S095026880999029X
- Jameson LJ, Newton A, Coole L, Newman EN, Carroll MW, Beeching NJ, et al. Prevalence of antibodies against hantaviruses in serum and saliva of adults living or working on farms in Yorkshire, United Kingdom. Viruses. 2014;6:524–34. http://dx.doi.org/10.3390/v6020524
- McKenna P, Clement J, Matthys P, Coyle PV, McCaughey C. Serological evidence of hantavirus disease in Northern Ireland. J Med Virol. 1994;43:33–8. http://dx.doi.org/10.1002/ jmv.1890430107
- Jameson LJ, Taori SK, Atkinson B, Levick P, Featherstone CA, van der Burgt G, et al. Pet rats as a source of hantavirus in England and Wales, 2013. Euro Surveill. 2013;18:18.
- Jackson JA, Begon M, Birtles R, Paterson S, Friberg IM, Hall A, et al. The analysis of immunological profiles in wild animals: a case study on immunodynamics in the field vole, *Microtus agrestis*. Mol Ecol. 2011;20:893–909. http://dx.doi.org/10.1111/ j.1365-294X.2010.04907.x
- Klempa B, Fichet-Calvet E, Lecompte E, Auste B, Aniskin V, Meisel H, et al. Hantavirus in African wood mouse, Guinea. Emerg Infect Dis. 2006;12:838–40. http://dx.doi.org/10.3201/ eid1205.051487

- Ronquist F, Huelsenbeck JP. MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics. 2003;19:1572–4. http://dx.doi.org/10.1093/bioinformatics/btg180
- 9. Nylander J. MrModeltest v2. Uppsala: Evolutionary Biology Centre, Uppsala University; 2004.
- Pether JV, Lloyd G. The clinical spectrum of human hantavirus infection in Somerset, UK. Epidemiol Infect. 1993;111:171–5. http://dx.doi.org/10.1017/S095026880005679X

Address for correspondence: Joseph A. Jackson, University of Salford, School of Environment and Life Sciences, Peel Building, Salford M5 4WT, UK; email: J.A.Jackson@Salford.ac.uk

## Measles Cases during Ebola Outbreak, West Africa, 2013–2106

Francesca Colavita,<sup>1</sup> Mirella Biava,<sup>1</sup> Concetta Castilletti, Serena Quartu, Francesco Vairo, Claudia Caglioti, Chiara Agrati, Eleonora Lalle, Licia Bordi, Simone Lanini, Michela Delli Guanti, Rossella Miccio, Giuseppe Ippolito, Maria R. Capobianchi, Antonino Di Caro; and the Lazzaro Spallanzani Institute for Research and Health Care Ebola Virus Disease Sierra Leone Study Group<sup>2</sup>

Author affiliations: Lazzaro Spallanzani Institute for Research and Health Care, Rome, Italy (F. Colavita, M. Biava, C. Castilletti, S. Quartu, F. Vairo, C. Caglioti, C. Agrati, E. Lalle, L. Bordi, S. Lanini, G. Ippolito, M.R. Capobianchi, A. Di Caro); Emergency Nongovernmental Organization, Milan, Italy (M. Delli Guanti, R. Miccio)

DOI: https://dx.doi.org/10.3201/eid2306.161682

The recent Ebola outbreak in West Africa caused breakdowns in public health systems, which might have caused outbreaks of vaccine-preventable diseases. We tested 80 patients admitted to an Ebola treatment center in Freetown, Sierra Leone, for measles. These patients were negative for Ebola virus. Measles virus IgM was detected in 13 (16%) of the patients.

The Ebola virus disease (EVD) outbreak in West Africa during 2013–2016 was one of the worst public health disasters in recent history; it caused >28,646 cases and 11,323

<sup>&</sup>lt;sup>1</sup>These authors contributed equally to this article. <sup>2</sup>Members of this group are listed at the end of this article.