In the United Kingdom, ≈1 of 2,000 persons could be infected with variant Creutzfeldt-Jakob disease (vCJD). Therefore, risk of transmission of vCJD by medical procedures remains a major concern for public health authorities. In this study, we used in vitro amplification of prions by protein misfolding cyclic amplification (PMCA) to estimate distribution and level of the vCJD agent in 21 tissues from 4 patients who died of clinical vCJD and from 1 asymptomatic person with vCJD. PMCA identified major levels of vCJD prions in a range of tissues, including liver, salivary gland, kidney, lung, and bone marrow. Bioassays confirmed that the quantitative estimate of levels of vCJD prion accumulation provided by PMCA are indicative of vCJD infectivity levels in tissues. Findings provide critical data for the design of measures to minimize risk for iatrogenic transmission of vCJD.

Prion diseases, or transmissible spongiform encephalopathies (TSEs), are fatal neurodegenerative disorders that occur naturally in sheep (scrapie), cattle (bovine spongiform encephalopathy [BSE]), and humans (Creutzfeldt-Jakob disease [CJD]). A key event in the pathogenesis of TSEs is the conversion of the normal cellular prion protein (PrPC, encoded by the PRNP gene) into an abnormal disease-associated isoform (PrPSc) in tissues of infected animals. PrPSc is completely degraded after controlled digestion with proteinase K in the presence of nondenaturing detergents. In contrast, PrPSc is N terminally truncated under the same conditions, resulting in a proteinase K–resistant prion (PrPRes) (1). In 1996, a new form of CJD, termed variant CJD (vCJD) was identified in the United Kingdom. vCJD is believed to result from zoonotic transmission of the BSE agent, probably as a consequence of dietary exposure to BSE-contaminated meat products (2,3). The total number of clinical cases of vCJD thus far identified is limited (227 patients worldwide at the time of writing this article). However, the estimated prevalence of asymptomatic vCJD in populations exposed to the BSE agent is uncertain (4).

In the United Kingdom, 32,441 appendix samples collected during surgery from patients born during 1941–1985 have been tested for abnormal prion protein accumulation by using immunohistochemical analysis. This study reported a vCJD prevalence estimate of 1/2,000 in persons in these age cohorts (95% CI 1/3,500–1/1,250) (5). No comparable data are available concerning the prevalence of asymptomatic vCJD in other countries, although BSE exposure is known to have occurred in several countries in continental Europe, as judged by cases of vCJD that are not attributable to exposure in the United Kingdom (http://www.cjd.ed.ac.uk/documents/worldfigs.pdf).

Over the past 2 decades, several studies have reported on the distribution of the vCJD agent in tissues of infected patients (6–8). Most of these studies did not detect the vCJD agent outside the nervous system (central, peripheral, and autonomic) and lymphoid tissues. However, the sensitivity of detection techniques for PrPRes used in these investigations was limited.

Protein misfolding cyclic amplification (PMCA) is believed to mimic prion replication in vitro, but in an accelerated form, which enables amplification of minute amounts of PrPSc and prion infectivity (9). In PMCA, a PrPSc-containing substrate is combined with a seed that might contain otherwise undetectable amounts of PrPSc. After repeated cycles of incubation and sonication, the
amount of PrPSc increases to levels at which they can be
detected by using conventional biochemical techniques.
Recently, our group and others have shown that PMCA
can detect endogenous vCJD agent in patient biologic fluid
such as urine and blood (10,11).

In this study, we evaluated the relative sensitivity of
PMCA versus that of bioassay in mice for detection of
the vCJD agent. We estimated by using PMCA the level
of vCJD prions in 21 tissues collected from 4 patients
who died of symptomatic vCJD and from a patient with
asymptomatic vCJD. We also determined whether vCJD
prion levels, as estimated by using PMCA, were consis-
tent with infectious titers, as estimated by bioassay with
transgenic mice.

Methods

Ethics Statements

All animal experiments were performed in compliance
with institutional and French national guidelines and in
accordance with the European Community Council Direc-
tive 86/609/EEC. Animal experiments that were part of this
study (national registration no. 01734.01) were approved
by the local ethic committee of the Ecole Nationale Véteri-
naire de Toulouse (Toulouse, France). Mouse inoculations
were performed under anesthesia with isofluorane. Mice that
displayed clinical signs of disease were anesthetized
with isofluorane before being humanely killed by inhala-
tion of CO2.

Human samples were obtained from the United King-
dom National CJD Research and Surveillance Unit Brain
and Tissue Bank, which is part of the Medical Research
Council Edinburgh Brain Bank (Edinburgh, Scotland,
UK). Tissue samples were pseudo-anonymized by using
a Brain Bank reference number. All case-patients in the
United Kingdom provided informed consent. Use of sam-
ples in this study was approved by the East of Scotland
Research Ethics Service for the Edinburgh Brain Bank
(16/ES/0084).

vCJD and Control Patients

We investigated tissues from 4 clinical vCJD case-pat-
ients (vCJD-1–vCJD-4) and 1 asymptomatic person with
vCJD who had received a transfusion of packed eryth-
rocytes from a donor who subsequently died from vCJD
(12). Tissues from 2 non–vCJD–affected patients were
used as controls. For case-patients who provided appro-
priate consent, the entire PRNP gene coding sequence
was established to exclude pathogenic mutations in this
gene (13,14).

Mouse Bioassays

Bioassays were performed by using mice expressing bo-
vine PrP (tgBov-tg110) as described (15,16). These mice
were observed daily and their neurologic status was as-
essed weekly. When clinically progressive TSE symptoms
were evident, or at the end of their lifespan, the animals
were euthanized. Survival time was expressed as the mean
± SD days postinoculation of mice positive for PrPSc.
For mice that showed no clinical signs, they were humanely
killed at the end of their natural lifespan (600–800 days).
In these instances, incubation periods are reported as >600

| Table 1. Endpoint titration of reference sample from a patient
| with vCJD in tgBov mice expressing bovine prion protein* |
|-----------------|-----------------|-----------------|
| **Dilution**    | **No. positive** | **Mean ± SD,**   |
|                 | **mice/no. tested** | **incubation, d** |
| Undiluted       | 6/6             | 249 ± 2         |
| 10\(^{-1}\)      | 6/6             | 283 ± 15        |
| 10\(^{-2}\)      | 6/6             | 316 ± 21        |
| 10\(^{-3}\)      | 6/6             | 342 ± 10        |
| 10\(^{-4}\)      | 6/6             | 453 ± 66        |
| 10\(^{-5}\)      | 2/6             | 479, 495†       |
| 10\(^{-6}\)      | 1/6             | 502†            |
| 10\(^{-7}\)      | 0/6             | >700            |

* A 10% (wt/vol) homogenate was prepared by using frontal cortex from a
clinically affected patient with vCJD. Groups of 6 tgBov mice were
inoculated intracerebrally with 20 μL of serial 10-fold dilutions of this
homogenate. Mice were considered positive when abnormal prion protein
deposition was detected in the brain. vCJD, variant Creutzfeldt-Jakob
disease. †Dilutions at which <50% of mice were positive.

| Table 2. Endpoint titration of PMCA seeding activity in a reference brain sample from a patient with vCJD* |
|------------------|------------------|------------------|
| **Amplification round** | **Reference vCJD 10% brain homogenate dilution series** | **no. of positive PMCA reactions/no. reactions conducted** |
|                  | **10\(^{-2}\)** | **10\(^{-3}\)** | **10\(^{-4}\)** | **10\(^{-5}\)** | **10\(^{-6}\)** | **10\(^{-7}\)** | **10\(^{-8}\)** | **10\(^{-9}\)** | **10\(^{-10}\)** |
| 1                 | 6/6             | 6/6             | 0/6             | 0/6             | 0/6             | 0/6             | 0/6             | 0/6             | 0/6             |
| 2                 | 6/6             | 6/6             | 5/6             | 3/6             | 0/6             | 0/6             | 0/6             | 0/6             | 0/6             |
| 3                 | 6/6             | 6/6             | 6/6             | 6/6             | 3/6             | 0/6             | 0/6             | 0/6             | 0/6             |
| 4                 | 6/6             | 6/6             | 6/6             | 6/6             | 6/6             | 5/6             | 2/6             | 0/6             | 0/6             |
| 5                 | 6/6             | 6/6             | 6/6             | 6/6             | 6/6             | 5/6             | 2/6             | 0/6             | 0/6             |
| 6                 | 6/6             | 6/6             | 6/6             | 6/6             | 6/6             | 5/6             | 2/6             | 0/6             | 0/6             |

*A 10% (wt/vol) homogenate was prepared by using frontal cortex from a symptomatic patient with vCJD (same homogenate as in Table 1) Samples were serially diluted 10-fold (10\(^{-2}\)–10\(^{-10}\)) before being used to seed PMCA reactions. Six individual replicates of each sample dilution were tested. PMCA substrate was prepared by using brains from transgenic mice overexpressing the ARQ variant of sheep prion protein. PMCA s were subjected to 8 rounds of amplification, each composed of 96 cycles (sonication for 10 s and incubation for 14.5 min at 39.5°C) in a Qsonica700 Sonicator (Qsonica LLC, Newtown, CT, USA). After each round, reaction products (1 volume) were mixed with fresh substrate (9 volumes) to seed the following round. Part of the same product was analyzed by Western blotting for abnormal PrPsc by using Sha31 antibody epitope YEDRYRE. Values are number of PMCA Western Blot–positive replicates corresponding to each round and each dilution. PMCA, protein misfolding cyclic amplification; PrPsc, proteinase K–resistant prion; vCJD, variant Creutzfeldt-Jakob disease.
days postinoculation, which corresponded to survival

time observed for ≥3 of 6 mice.

**Estimation of Infectious Prion Titors**

We estimated infectious titer in a reference 10% (wt/vol)
fron t cortex homogenate from a clinical vCJD patient by
using endpoint titration (intracerebral route) in tgBov mice.
Infectious titer (50% lethal dose/g intracerebral in tgBov
mice) was estimated by using the Spearman method.

The titer of prion infectivity in vCJD–affected patient
bone marrow samples was estimated by using the method
of Arnold et al. (17). This method uses the probability of
survival (attack rate at each dilution) and the individual
mouse incubation periods at each dilution to estimate in-
fected load and is thus able to provide more accurate es-
timation of titer than using either attack rate or incubation
period data alone.

**PMCA Reactions**

A transgenic mouse line that expresses ovine A136R
PrP variant PrP<sup>C</sup> (tgShXI) was used to prepare the PMCA
substrate as described (18,19). PMCA amplification was per-
fomed as described (11). Each PMCA experiment included
a reference vCJD sample (10% brain homogenate) as a con-
trol for the amplification efficiency. Unseeded controls (1
unseeded control for 8 seeded reactions) were also included
in each experiment. For each tested dilution of each sample,
≥4 replicates were tested in 2 independent experiments. For
each sample, the highest dilution showing ≥50% of positive
replicates (presence of detectable PrP<sup>res</sup> in the reaction as as-
sessed by using Western blotting) was determined.

**Detection of Abnormal PrP by Western Blotting and
Paraffin-Embedded Tissue Blotting**

Extraction of proteinase K–resistant abnormal PrP and
Western blotting were performed as described (11). Im-
munodetection was performed by using 2 PrP-specific
monoclonal antibodies, Sha31 (1 µg/mL) (20), and 12B2
(4 µg/mL) (21), which recognize amino acid sequences
YEDRYyre (145–152), and WGQGG (89–93), respec-
tively. Paraffin-embedded tissue blotting was performed as
described (22,23).

**Results**

**Sensitivity of vCJD Agent Detection by
PMCA and Bioassay**

To determine the relative sensitivity of PMCA, we reti-
trated a reference sample (10% cerebral cortex homog-
enate from a vCJD-affected patient) that had previously
undergone endpoint titration (IC inoculation route; Table
1) in bovine PrP–expressing mice (tgBov, intracerebral
route, 10<sup>3.7</sup> 50% lethal dose/g). PMCA substrate was
prepared by using brains from transgenic mice overexpressing the
ARQ variant of sheep prion protein. An unseeded (lane U) reaction
was included as a specificity control. PMCAS were subjected to 6
rounds of amplification, each composed of 96 cycles (sonication
for 10 s and incubation for 14.5 min at 39.5°C) in a Qsonica700
Sonicator (Qsonica LLC, Newtown, CT, USA). After each round,
reaction products (1 volume) were mixed with fresh substrate (9
volumes) to seed the following round. Part of the same product
was analyzed by Western blotting for abnormal PrP<sup>res</sup> (Sha31
antibody epitope YEDRYYRE). A sheep scrapie sample (WB cont)
was included as a control on each gel. WB, Western blot.
showed an infectious titer of $10^{7.7} \text{LD}_{50}/g$. When we took
into account the 4-fold lower amount of material used
to seed the PMCA reaction compared with material used in
mouse inoculations, we found that the PMCA protocol
used was 465 times more sensitive than the bioassay of
tgBov mice for detection of vCJD prions.

PMCA for Control and vCJD Patients
We compiled basic demographic data for vCJD and con-
tral patients (Table 3). A 10-fold dilution series of 10% homogenates from the vCJD–affected and non–vCJD–af-
fected control patients was prepared, and this series was
subjected to 4 rounds of PMCA. Amplification products
from each round were tested for PrP\(^\text{res}\) using by Western
blotting (Table 4; Figure 2).

We found that none of the reactions seeded with tissue
homogenates from non–CJD controls were positive for
PrP\(^\text{res}\) (Table 4). In contrast, PMCA reactions seeded with
tissues from the 4 symptomatic vCJD patients were posi-
tive for PrP\(^\text{res}\) (Table 4; Figure 2). As expected, among test-
ed tissues, brain homogenates (temporal cortex) showed
the highest seeding activity (highest PrP\(^\text{res}\)-positive dilu-
tion $10^{-8}$). All lymphoid organs tested also showed seed-
ing activity, but the highest PMCA-positive dilution varied

Table 3. Characteristics of 5 patients with vCJD and 2 controls in study of distribution and quantitative estimates of variant prions in tissues*

<table>
<thead>
<tr>
<th>Patient identification no.</th>
<th>Diagnosis</th>
<th>Sex</th>
<th>Year of death</th>
<th>Age, y, at death</th>
<th>Disease duration, mo</th>
<th>PRNP gene codon 129</th>
<th>PRNP gene mutations</th>
</tr>
</thead>
<tbody>
<tr>
<td>vCJD-1</td>
<td>vCJD</td>
<td>M</td>
<td>1999</td>
<td>33</td>
<td>18</td>
<td>MM</td>
<td>None detected</td>
</tr>
<tr>
<td>vCJD-2</td>
<td>vCJD</td>
<td>F</td>
<td>2000</td>
<td>17</td>
<td>18</td>
<td>MM</td>
<td>None detected</td>
</tr>
<tr>
<td>vCJD-3</td>
<td>vCJD</td>
<td>M</td>
<td>2000</td>
<td>26</td>
<td>10</td>
<td>MM</td>
<td>None detected</td>
</tr>
<tr>
<td>vCJD-A</td>
<td>Asymptomatic vCJD</td>
<td>F</td>
<td>2004</td>
<td>82</td>
<td>NA</td>
<td>NA</td>
<td>None detected</td>
</tr>
<tr>
<td>NC-1</td>
<td>No CJD (tumor, infarction, ischemia)</td>
<td>F</td>
<td>2005</td>
<td>85</td>
<td>NA</td>
<td>MM</td>
<td>No consent for sequencing</td>
</tr>
<tr>
<td>NC-2</td>
<td>No CJD (Alzheimer’s disease, infarction, ischemia)</td>
<td>F</td>
<td>2010</td>
<td>80</td>
<td>NA</td>
<td>MM</td>
<td>None detected</td>
</tr>
</tbody>
</table>

*NA, not applicable; vCJD, variant Creutzfeldt-Jakob disease.

Table 4. Protein misfolding cyclic amplification reactions seeded with tissue homogenate from vCJD and control patients*

<table>
<thead>
<tr>
<th>Tissue</th>
<th>Clinical vCJD patients, Met(<em>{129})/Met(</em>{129})</th>
<th>Preclinical vCJD patient, Met(<em>{129}/\text{Val}</em>{129})</th>
<th>Non–vCJD controls, Met(<em>{129}/\text{Met}</em>{129})</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>vCJD-1</td>
<td>vCJD-2</td>
<td>vCJD-3</td>
</tr>
<tr>
<td>Frontal cortex</td>
<td>$10^{-8}$</td>
<td>$10^{-8}$</td>
<td>$10^{-8}$</td>
</tr>
<tr>
<td>Pituitary gland</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Trigeminal ganglia</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Dorsal root ganglia</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Cervical lymph node</td>
<td>$10^{-3}$</td>
<td>$10^{-4}$</td>
<td>$10^{-4}$</td>
</tr>
<tr>
<td>Tonsil</td>
<td>$10^{-3}$</td>
<td>$10^{-4}$</td>
<td>$10^{-4}$</td>
</tr>
<tr>
<td>Appendix</td>
<td>$10^{-4}$</td>
<td>$10^{-4}$</td>
<td>$10^{-4}$</td>
</tr>
<tr>
<td>Distal ileum</td>
<td>$10^{-4}$</td>
<td>$10^{-5}$</td>
<td>$10^{-5}$</td>
</tr>
<tr>
<td>Spleen</td>
<td>$10^{-4}$</td>
<td>$10^{-4}$</td>
<td>$10^{-5}$</td>
</tr>
<tr>
<td>Thymus</td>
<td>$10^{-3}$</td>
<td>$10^{-3}$</td>
<td>$10^{-2}$</td>
</tr>
<tr>
<td>Lung</td>
<td>$10^{-3}$</td>
<td>$10^{-3}$</td>
<td>–</td>
</tr>
<tr>
<td>Heart</td>
<td>$10^{-3}$</td>
<td>$10^{-2}$</td>
<td>–</td>
</tr>
<tr>
<td>Liver</td>
<td>$10^{-3}$</td>
<td>$10^{-2}$</td>
<td>$10^{-2}$</td>
</tr>
<tr>
<td>Kidney</td>
<td>$10^{-3}$</td>
<td>$10^{-3}$</td>
<td>–</td>
</tr>
<tr>
<td>Salivary gland</td>
<td>$10^{-4}$</td>
<td>$10^{-3}$</td>
<td>$10^{-2}$</td>
</tr>
<tr>
<td>Pancreas</td>
<td>$10^{-2}$</td>
<td>–</td>
<td>$10^{-2}$</td>
</tr>
<tr>
<td>Thyroid</td>
<td>$10^{-2}$</td>
<td>–</td>
<td>$10^{-2}$</td>
</tr>
<tr>
<td>Adrenal gland</td>
<td>$10^{-3}$</td>
<td>$10^{-3}$</td>
<td>$10^{-3}$</td>
</tr>
<tr>
<td>Bone marrow</td>
<td>$10^{-4}$</td>
<td>$10^{-5}$</td>
<td>$10^{-3}$</td>
</tr>
<tr>
<td>Skeletal muscle</td>
<td>$10^{-4}$</td>
<td>$10^{-2}$</td>
<td>NA</td>
</tr>
<tr>
<td>Testis</td>
<td>–</td>
<td>NA</td>
<td>–</td>
</tr>
<tr>
<td>Ovary</td>
<td>NA</td>
<td>$10^{-4}$</td>
<td>NA</td>
</tr>
</tbody>
</table>

*PMCA reactions were seeded with 10-fold serial dilutions of 10% tissues homogenates ($10^{-2}$–$10^{-9}$) that had been collected postmortem from 4 symptomatic vCJD patients (vCJD-1–vCJD-4) or an asymptomatic vCJD-infected person (vCJD-A). At least 4 replicates of each sample dilution were tested in 2 independent PMCA experiments. Prions from patients vCJD-1–vCJD-4 were homozygous for methionine at codon 129 of the PRNP gene. Prions from patient vCJD-A were heterozygous (methionine/valine) at codon 129 of the PRNP gene. PMCA substrate was prepared by using brains from transgenic mice overexpressing the ARQ variant of sheep prion protein. Reactions seeded with tissues from 2 non–vCJD-infected control patients (NC-1 and NC-2) were included as negative controls. PMCAs were subjected to 4 rounds of amplification, each composed of 96 cycles (sonication for 10 s and incubation for 14.5 min at 39.5°C) in a Qsonica700 Sonicator (Qsonica LLC, Newtown, CT, USA). PMCA reactions were analyzed by Western blotting for proteinase K–resistant PrP by using Sha31 antibody epitope YEDRYYRE. Values are the highest dilution that resulted in a positive Western blot result in >50% of the tested replicates after 4 PMCA amplification rounds. NA, not applicable; PMCA, protein misfolding cyclic amplification; PrP, prion protein; vCJD, variant Creutzfeldt-Jakob disease; –, negative.

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According to the organs tested from 10\(^{-2}\) (thymus) to 10\(^{-6}\) (distal ileum and tonsil). Moreover, for a given lymphoid organ, \(<10^2\)-fold differences was observed in seeding activity, depending on the patient and sample tested. These data indicate that for symptomatic vCJD patients, lymphoid organs contain 10\(^{5}\)–10\(^{6}\)-fold less prion seeding activity than the same amount of brain tissue (Table 4).

Salivary gland, adrenal gland, liver, and bone marrow from the 4 symptomatic vCJD patients showed positive reactions by PMCA (Figures 2, 3). Using the highest dilution to show a positive reaction as a measure of seeding activity, we found that the vCJD agent in these tissues was 10\(^{5}\)–10\(^{6}\)-fold lower than that for the brain. PrP\(_{\text{res}}\) was also detected by PMCA reactions seeded with heart, liver, kidney, skeletal muscle, several endocrine/exocrine glands (pancreas, thyroid), and gonads, from some, but not all, of the 4 clinical vCJD patients. Positive tissues contained a level of vCJD seeding activity that was equivalent to those observed in distal ileum (i.e., 10\(^{5}\)–10\(^{6}\)-fold lower than for the brain). Irrespective of the tissue used to seed the PMCA reactions, the PrP\(_{\text{res}}\) Western blot profile for positive reactions was indistinguishable from that observed in reactions seeded with the vCJD brain control (Figure 3).
Analysis of Tissues from an Asymptomatic vCJD-Infected Person

Prion seeding activity was not detected in the brain (temporal cortex) of the asymptomatic vCJD–affected patient, who was infected with a PRNP gene codon 129 heterozygote (Met/Val\textsubscript{129}) prion (12) (Table 4; Figure 2). PMCA reactions seeded with dorsal root ganglia or trigeminal ganglia homogenates from this patient showed negative results. However, seeding activity was detected in the pituitary gland (highest PrP\textsuperscript{res}-positive dilution 10\textsuperscript{−2}). In addition, as for the symptomatic vCJD patient, PMCA amplification readily detected vCJD prions in all lymphoid organs tested from this asymptomatic person. On the basis of PMCA results, the vCJD agent load in lymphoid organs in this asymptomatic patient infected with the PRNP gene codon 129 Met/Val\textsubscript{129} prion was similar to those for patients infected with Met/Met\textsubscript{129} prions during the clinical stage of disease.

In addition to findings for lymphoid organs, prion seeding activity was detectable in certain peripheral tissues (salivary gland, lung, and liver) from this patient (Tables 4; Figures 2, 3). Certain tissues, such as bone marrow or adrenal gland, that contained a substantial prion seeding activity in the clinically affected patients showed negative results. Again, the PrP\textsuperscript{res} Western blot profile for positive reactions was indistinguishable from that observed for reactions seeded with the vCJD brain control.

vCJD Infectivity in Bone Marrow

To test whether PMCA seeding activity in peripheral tissues from vCJD patients correlated with infectivity, we inoculated bone marrow samples from the symptomatic patient into tgBov mice. Clinical TSE was observed in mice that were inoculated with each of the 4 bone marrow samples. The PrP\textsuperscript{res} Western blot profile and the PrP\textsuperscript{res} distribution pattern, as assessed by paraffin-embedded tissue blotting for brain of the bone marrow–inoculated mice, were identical to those observed in tgBov mice inoculated with the vCJD brain control sample (Figure 4).

Data obtained for mice inoculated with bone marrow samples were also used to estimate prion infectivity levels in these samples. For this purpose, we applied the method of Arnold et al. (17). This method combines the probability of survival (attack rate) and the individual mouse incubation period to provide an estimation of infectious titers. We used data corresponding to endpoint titration in tgBov mice for reference vCJD sample (frontal cortex from a clinical vCJD patient) (Table 1) to derive the relationship between prion titer of inoculum and the probability of infection and length of the incubation period (Figure 5). We found that bone marrow samples had an infectious titer that ranged from 10\textsuperscript{−2.3} LD\textsubscript{50}/g through 10\textsuperscript{−4.7} LD\textsubscript{50}/g in tgBov mice (Table 5).
These values are consistent with a 10³–10⁵ lower infectivity load in bone marrow samples than in the reference vCJD brain sample. Consistent with the PMCA results (Table 4), we found that prion load in bone marrow samples (highest PrP\textsuperscript{res-positive} dilution [10³–10⁵]) was also 10³–10⁵-fold lower than for the reference vCJD isolate (highest PrP\textsuperscript{res-positive} dilution [10³]). These results strongly support the idea that PMCA seeding activity provides a reliable estimate of the prion load in tissues from vCJD-infected patients.

**Discussion**

Most previous studies with tissue from vCJD patients have failed to identify consistent accumulation of the vCJD agent outside the nervous and lymphoreticular systems. However, data obtained in this study clearly demonstrate the presence of vCJD prions in a wide and unexpected variety of peripheral tissues.

Natural scrapie and experimental BSE in sheep are 2 models of orally transmitted prion diseases (24,25). In both diseases, the agent accumulates in the lymphoreticular system and the enteric nervous system during the early preclinical phase of the incubation period. Moreover, an early and persistent prionemia is observed in asymptomatic infected animals (26,27). These features were also observed in vCJD in humans and in view of the likely origin of vCJD (oral exposure to BSE agent), these similarities have led to a consensus that BSE and scrapie in sheep and vCJD in human have a common pathogenesis (28).

Although vCJD prions in a variety tissues, such as bone marrow, kidney, salivary gland, skeletal muscle, pancreas, liver, or heart, might be surprising, each of these tissues has already been demonstrated to accumulate prion infectivity or abnormal prion protein in TSE-infected sheep (29–33). Because low levels of infectivity have been reported in blood fractions from a vCJD-affected...
variant Creutzfeldt-Jakob Disease Prions

Figure 5. Dose–response relationship for A) incubation period and B) probability of infection of bovine PrP–expressing mice. Data were derived from an endpoint titration of 10% (wt/vol) frontal cortex homogenate from a patient with variant Creutzfeldt-Jakob disease. This homogenate was inoculated into tgBov mouse (20 µL by intracerebral [ic] route; Table 1). This procedure was used to establish a model that estimates infectious titer in a homogenate on the basis of incubation period and the probability of infection in inoculated mice. Model plots are indicated by solid lines. +, observed value. LD50, 50% lethal dose.

The patient in our study who was infected with a prion containing PRNP gene codon 129 Met/Val is 1 of only 2 identified vCJD agent–infected persons known to have died of other causes before onset clinical symptoms of vCJD, and the only person who provided consent to sample autopsy tissues for research. For this patient, all previous investigations did not detect abnormal prion protein or infectivity in peripheral tissues, the presence of vCJD agent in peripheral tissues of the asymptomatic vCJD–affected person (PRNP gene codon 129 Met/Val) or the difference between the vCJD seeding activity in lymphoreticular tissues was similar to that observed for symptomatic vCJD patients, several tissues that were positive in clinically affected patients were negative in this heterozygous asymptomatic person. These findings suggest that involvement of some peripheral tissues might occur at a later stage in the incubation period than others, or that they could involve recirculation of the agent from the central nervous system (i.e., centrifugal spread in a late stage). However, we cannot discount the possibility that these differences in tissue distribution are caused by the hematogenous route of infection in this person (as opposed to the probable oral route in patients with clinical vCJD) or the difference between the PRNP gene codon 129 genotype of the asymptomatic vCJD–affected person (PRNP gene codon 129 Met/Val) and persons with clinical vCJD (PRNP gene codon 129 Met/Met).

Irrespective of the actual explanation for these differences, the presence of vCJD agent in peripheral tissues of patients during preclinical and clinical stage of the disease.

Table 5. Bone marrow sample bioassay in bovine PrP–expressing mice (tgBov) for 4 patients with vCJD

<table>
<thead>
<tr>
<th>Sample</th>
<th>No. positive/no. inoculated mice</th>
<th>Mean ± SD incubation, d</th>
<th>Mean infectious titer, LD50/g (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>vCJD-1</td>
<td>5/5</td>
<td>458 ± 37</td>
<td>10−1.0 (10−2.5 – 10−1.0)</td>
</tr>
<tr>
<td>vCJD-2</td>
<td>6/6</td>
<td>373 ± 35</td>
<td>10−3.7 (10−3.0 – 10−3.4)</td>
</tr>
<tr>
<td>vCJD-3</td>
<td>4/6</td>
<td>504 ± 10</td>
<td>10−3.3 (10−2.1 – 10−3.7)</td>
</tr>
<tr>
<td>vCJD-4</td>
<td>6/6</td>
<td>447 ± 91</td>
<td>10−3.9 (10−4.4 – 10−4.5)</td>
</tr>
<tr>
<td>PBS control</td>
<td>0/6</td>
<td>&gt;600</td>
<td>NA</td>
</tr>
</tbody>
</table>

*A 10% wt/vol bone marrow homogenate prepared from 4 symptomatic vCJD patients (Table 3) was inoculated intracerebrally into 6 tgBov mice (20 µL/mouse). One mouse (inoculated with homogenate from patient vCJD-1) died within the first few days after intracerebral inoculation. Mice were euthanized when they showed clinical signs of prion infection or after 600-d postinoculation. Mice were considered prion infected when abnormal PrP deposition was detected in brain. Infectious prion titers were estimated by using the method of Arnold et al. (17). The method uses the probability of survival (attack rate at each dilution) and the individual mouse incubation periods at each dilution to estimate infectious load. Infectious titers are given as estimated values. LD50, 50% lethal dose; NA, not applicable; PBS, phosphate-buffered saline; PrP, prion protein; vCJD, variant Creutzfeldt-Jakob disease.
indicates the potential for iatrogenic transmission of this fatal neurologic condition by surgical procedures. Furthermore, this finding shows that, for certain peripheral tissues, a level of infectivity equivalent to an end stage titer (and attendant risk) is reached at a preclinical stage.

Several hundred cases of iatrogenic CJD have been reported worldwide. These cases appear to result from transmission of sporadic CJD, and most cases have occurred in recipients of human dura mater grafts or after administration of human growth hormone extracted from cadaveric pituitaries (39). Although in sporadic CJD the distribution of the agent is largely restricted to the nervous system (central and peripheral), the wide distribution of the vCJD agent in the asymptomatic infected patient we report might serve to increase the range of medical procedures, including dentistry, organ transplant, and surgery involving nondisposable equipment, that might result in iatrogenic transmission of vCJD (40–43).

Nevertheless, >20 years after identification of the first vCJD patients, only 5 cases that are a probable consequence of iatrogenic vCJD transmission are known, all in the United Kingdom and associated with blood and blood products. These cases were caused by transfusion of non–leukocyte-depleted erythrocyte concentrates or by treatment involving large amounts of pooled plasma from the United Kingdom that were known to include donations from persons who later showed development of vCJD (12,44–46).

None of the 220 other vCJD cases identified worldwide have been linked to any other medical or dental procedure. Whereas this fact is reassuring, it would be unwise to disregard the threat that vCJD still poses for public health. Despite the relatively low number (n = 178) of vCJD clinical cases observed in the United Kingdom, the most recent epidemiologic studies indicate that ≥1 of 2,000 persons in the United Kingdom could be infected with the vCJD agent (as indicated by the presence of abnormal prion protein detected by immunohistochemical analysis of lymphoid follicles in the appendix). Each asymptomatic vCJD-infected person represents a potential source of secondary infection. The data in our report offer an opportunity for refining measures that were implemented in many countries to limit the risk for vCJD iatrogenic transmission. The apparent concordance between PMCA biochemical and infectivity bioassay data, and the higher analytical sensitivity of PMCA, suggest that future research need not rely exclusively on time-consuming and costly animal bioassay.

Our results indicate the need for vCJD screening assays. After more than a decade of effort, several vCJD blood detection tests have reached a stage in their development that could enable their evaluation as screening or confirmatory assays (11,47,48). In particular, there is now a strong case for use of PMCA in a highly sensitive and specific blood test for vCJD, as indicated by our previous studies (11,16) and studies by Bougard et al. (35) and Concha-Marambio et al. (36). The relationship shown here between PrPres amplification by PMCA and detection of infectivity by bioassay indicates that PMCA seeding activity is a good surrogate marker of infectivity and could provide a sound basis for a vCJD blood test for use with blood or tissue donors.

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References

RESEARCH


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etymologia

Creutzfeldt-Jakob [ˈkroʊtʃəfɪlt-ˈdʒækəb] Disease

Ronnie Henry, Frederick A. Murphy

In 1920, German neuropathologist Alfons Maria Jakob described a series of 6 patients with spasticity and progressive dementia associated with neural degeneration. Shortly thereafter, in 1921, another German neuropathologist, Hans Gerhardt Creutzfeldt, independently published a similar case. Jakob gave credit to Creutzfeldt for describing the syndrome first, without realizing he had also uncovered the new syndrome. Walther Spilmeier first used the term “Creutzfeldt-Jakob disease” (CJD) in 1922. CJD occurs worldwide as a rare, sporadic disease, with genetic and iatrogenic forms. A zoonotic form, variant CJD (vCJD), is caused by infection with a prion derived from bovines and occurs predominantly in the United Kingdom.

This issue of Emerging Infectious Diseases’ long-running Etymologia series is dedicated to the memory of Richard T. Johnson, MD (1931–2015), the leading prion disease authority in the United States for many years and great friend of CDC’s infectious disease programs, so many of which involve central nervous system disorders.

Sources


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