Powassan virus (POWV) lineage II, also known as deer tick virus, is an emerging tickborne flavivirus (1) transmitted by *Ixodes scapularis* ticks, which are also the primary vector for *Borrelia burgdorferi* (Lyme disease pathogen). In POWV-endemic regions, up to 7% of ticks carry the virus, and seroprevalence among small mammalian hosts can exceed 90% (2,3). Because the territory of *I. scapularis* is expanding and the prevalence of POWV in ticks and mammals is increasing, POWV poses an increasing threat (2–5). The seroprevalence of POWV in humans in some regions of North America is known (range 0.5%–3.3%), but because the geographic distribution is quite extensive, the seroprevalence of most at-risk populations is uncertain (6).

POWV is typically detected with an IgM antibody capture ELISA or an IgM immunofluorescence antibody (IFA) assay. Cases are confirmed by ≥90% or ≥50% plaque reduction neutralization test (PRNT$_{90}$ or PRNT$_{50}$), detection of virus-specific nucleic acids, isolation in culture, or a ≥4-fold increase in antibody titers from paired acute and convalescent sera (7–9). Using these assays, investigators have identified ≥100 cases of POWV encephalitis; however, the actual incidence is likely higher (1,6).

Although nonneuroinvasive disease has been described for other arboviral illnesses, our knowledge of POWV has been limited to patients with neuroinvasive disease (1,8,10,11). In this study, we evaluated the seroreactivity for POWV in US Midwest patients, many of whom did not have neuroinvasive disease.

**The Study**

We selected patients with suspected tickborne disease (TBD; n = 95) and patients undergoing routine chemical screening (n = 50) who sought treatment during July–August 2015 at the Marshfield Clinic in northern Wisconsin, a TBD-endemic area. Patients were considered to have suspected TBD if a serologic test for *B. burgdorferi* was ordered. The chemical screening cohort included patients who had a complete metabolic or lipid panel ordered as part of their clinical care. We evaluated POWV seroreactivity of specimens from these patient cohorts and, of the patients with serologic evidence of POWV infection and available clinical data, described the clinical features of their disease. All human subject research protocols were approved by the Marshfield Clinic Research Institute Institutional Review Board.

We performed screening assays on all specimens for tick-borne encephalitis virus complex (TBEV-C) and *B. burgdorferi* and performed POWV serology on TBEV-C–positive specimens (Figure; detailed methods in online Technical Appendix, https://wwwnc.cdc.gov/EID/article/23/8/16-1971-Techapp1.pdf). To evaluate heterologous flavivirus cross-reactivity, we performed the West Nile virus (WNV) enzyme immunoassay (EUROIMMUN, Mountain Lakes, NJ, USA) with TBEV-C–positive samples. We also performed the Flavivirus Mosaic Panel (EUROIMMUN), an IgG IFA assay panel including tests for TBEV, WNV, yellow fever virus, dengue viruses 1–4, and Japanese encephalitis virus, on samples positive for POWV encephalitis by the IFA assay. Patient vaccination status and travel history were also considered.

Clinical data were available for 51 (53.7%) TBD patients and 50 (100%) patients with routine chemistry screening completed. For those with clinical data available, we classified their cases as probable or confirmed by using the Centers for Disease Control and Prevention case...
Powassan Virus Infection and Lyme Disease

TBD and chemistry screening residual specimens

1. TBEV-C IgG and IgM EIA
2. Two-tiered Lyme serology

TBEV-C EIA negative → Negative for POWV
TBEV-C EIA positive

1. POWV IgG and IgM EIA
2. WNV EIA

POWV IgG and IgM negative or WNV EIA positive → Negative for POWV
POWV IgM IFA ≥20 or IgG IFA ≥40

1. POWV PRNT90 positive and POWV RT-PCR negative
2. POWV RT-PCR*
3. Flavivirus IgG IFA mosaic panel†

POWV PRNT90 positive or POWV RT-PCR positive

Laboratory-confirmed POWV infection

Serologic evidence for POWV infection

Powassan Virus Infection and Lyme Disease

Figure. Flow chart showing series of tests performed on specimens obtained from patients with suspected TBD and patients undergoing routine chemical screening to determine POWV seroreactivity, Wisconsin, July–August 2015. *Performed for TBD samples positive for POWV IgG or IgM and chemical screening samples positive for POWV IgM by IFA assay. †Performed for samples positive for POWV IgG by IFA assay. EIA, enzyme immunoassay; IFA, immunofluorescence antibody assay; POWV, Powassan virus; PRNT90, ≥90% plaque reduction neutralization test; RT-PCR, reverse transcription PCR; TBD, tickborne disease; TBEV-C, tick-borne encephalitis virus complex; WNV, West Nile virus.

Serologic evidence of POWV infection was present in 9 (9.5%) TBD patients and 2 (4.0%) patients with routine chemistry screening completed (p = 0.33) (Table 1). POWV infection was confirmed in 3 (3.2%) TBD patients (2 by

Table 1. TBEV-C and Borrelia burgdorferi serologic test results and POWV RT-PCR test results of patients with positive POWV IFA assay results, Wisconsin, July–August 2015

<table>
<thead>
<tr>
<th>Patient no.</th>
<th>Suspected TBD patients</th>
<th>TBEV-C IgM EIA</th>
<th>TBEV-C IgG EIA</th>
<th>POWV IgM IFA assay†</th>
<th>POWV IgG IFA assay‡</th>
<th>POWV PRNT§</th>
<th>POWV RT-PCR¶</th>
<th>B. burgdorferi#</th>
</tr>
</thead>
<tbody>
<tr>
<td>1**††</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>2††</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>IgG and IgM</td>
<td>–</td>
</tr>
<tr>
<td>3††</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>IgG and IgM</td>
<td>–</td>
</tr>
<tr>
<td>4††</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>IgG and IgM</td>
<td>–</td>
</tr>
<tr>
<td>5</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>IgG and IgM</td>
<td>–</td>
</tr>
<tr>
<td>6</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>IgG and IgM</td>
<td>–</td>
</tr>
<tr>
<td>7</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>IgG and IgM</td>
<td>–</td>
</tr>
<tr>
<td>8</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>IgG and IgM</td>
<td>–</td>
</tr>
<tr>
<td>9††</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>IgG and IgM</td>
<td>–</td>
</tr>
<tr>
<td>Patients screened by chemical methods</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1c</td>
<td>+</td>
<td>–</td>
<td>–</td>
<td>+</td>
<td>–</td>
<td>NA</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>2ct††</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>–</td>
<td>NA</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>

†Titers ≥1:20 were considered positive.
‡Titers ≥1:40 were considered positive.
§Positive if sample had a PRNT90 titer.
¶Not performed in specimens with a negative POWV IgM IFA assay result.
#Samples were screened by EIA and followed up by Western blot.
**Cross-reactivity on POWV IgG IFA assay is consistent with a history of West Nile virus infection.
††Clinical data were available.

definitions (7). We performed statistical analysis with SAS 9.3 (SAS Institute, Inc., Cary NC, USA) and compared categorical variables by using Fisher exact tests. Significance was defined as p<0.05.
PRNT<sub>90</sub> [titer range 1:160–1:320] and 1 by reverse transcription PCR) and 0 chemical screening patients (p = 0.55). Of the 3 patients with confirmed POWV infection, evidence of acute infection (IgM positivity) was found in 2 (2.7%). Patients positive only for IgM by IFA assay did not have PRNT<sub>90</sub> titers, which was expected because neutralizing

<table>
<thead>
<tr>
<th>Suspected TBD patients</th>
<th>POWV test results</th>
<th>Borrelia burgdorferi test results</th>
<th>Clinical features</th>
<th>Comorbidities</th>
<th>CDC case classification</th>
<th>Travel history</th>
<th>Location of tick exposure</th>
<th>Vaccine history</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>IgG &gt;1:40</td>
<td>IgG and IgM</td>
<td>56-year-old man with 2-wk history of erythema migrans. Treated with doxycycline for 14 d.</td>
<td>Metabolic syndrome, hypertension, 9 y previous had WNV infection</td>
<td>Probable</td>
<td>Midwest</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>2</td>
<td>IgG &gt;1:40, PRNT 1:160</td>
<td>IgG and IgM</td>
<td>53-year-old man with 3-d history of urticarial rash, malaise, fever, and fatigue. Patient had chills 3 wks prior that resolved. CBC results: leukocytes 7.3 × 10&lt;sup&gt;9&lt;/sup&gt;/L, Hb 13.6 g/dL, Hct 39.9%, Plt count 322 × 10&lt;sup&gt;3&lt;/sup&gt;/µL; CRP 3.9 nmol/L. PCR neg for Anaplasma sp., Babesia sp., and Ehrlichia muris. Treated with doxycycline for 21 d with complete resolution of symptoms. No history of neuroinvasive disease or TBD.</td>
<td>Hyperlipidemia</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>IgM &gt;1:20</td>
<td>IgG and IgM</td>
<td>14-year-old girl with 3-d history of urticarial rash. CBC results: leukocytes 8.8 × 10&lt;sup&gt;9&lt;/sup&gt;/L, Hb 13.0 g/dL, Hct 40.3%, Plt 393 × 10&lt;sup&gt;3&lt;/sup&gt;/µL; CRP 3.6 nmol/L. Treated with doxycycline for 14 d.</td>
<td>None</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>IgM &gt;1:20</td>
<td>IgG and IgM</td>
<td>4-year-old girl with 1-wk history of fever (103&lt;sup&gt;°&lt;/sup&gt;F), listless, headache, fatigue, and maculopapular rash. PCR neg for Anaplasma sp., Babesia sp., and Ehrlichia muris. Treated with amoxicillin for 21 d.</td>
<td>None</td>
<td>Probable</td>
<td>–</td>
<td>–</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>IgM &gt;1:20</td>
<td>IgG and IgM</td>
<td>3-year-old girl with 1-wk history of intermittent fever, fussiness, and erythema migrans. After development of an urticarial rash, treatment with cefuroxime was changed to amoxicillin for 21 d.</td>
<td>None</td>
<td>Probable</td>
<td>Midwest</td>
<td>–</td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Patients screened by chemical methods</th>
<th>POWV test results</th>
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<th>Comorbidities</th>
<th>CDC case classification</th>
<th>Travel history</th>
<th>Location of tick exposure</th>
<th>Vaccine history</th>
</tr>
</thead>
<tbody>
<tr>
<td>1c</td>
<td>IgG &gt;1:40</td>
<td>Neg</td>
<td>68-year-old man with no signs or symptoms of acute infectious disease. No history of neuroinvasive disease or TBD. Died from liver cirrhosis.</td>
<td>Coronary artery disease, liver cirrhosis, end stage renal disease</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td></td>
</tr>
<tr>
<td>2c</td>
<td>IgM &gt;1:20, IgG &gt;1:40</td>
<td>Neg</td>
<td>76-year-old woman with 2-d history of fever, chills, and MRSA infection of the right hand. Mild abdominal pain and diarrhea occurred later in course. CBC results: leukocytes 13.7 × 10&lt;sup&gt;9&lt;/sup&gt;/L, Hb 9.2 g/dL, Hct 29.7%, Plt 180 × 10&lt;sup&gt;9&lt;/sup&gt;/µL; CRP 1.5 nmol/L; Procalcitonin 0.1 µg/L. Received daptomycin for 16 d with full recovery. Currently deceased, unknown cause of death.</td>
<td>Congestive heart failure, rheumatoid arthritis on immunosuppressive medications</td>
<td>Probable</td>
<td>–</td>
<td>–</td>
<td></td>
</tr>
</tbody>
</table>

*CBC, complete blood cell count; CDC, Centers for Disease Control and Prevention; CRP, C-reactive protein; Hb, hemoglobin; Hct, hematocrit; IFA, immunofluorescence antibody; MRSA, multidrug-resistant Staphylococcus aureus; neg, negative; Plt, platelet; POWV, Powassan virus; PRNT, plaque reduction neutralization test; WNV, West Nile virus; TBD, tickborne disease; –, no history.
†Samples were screened by EIA and followed up by Western blot.
‡Patient-reported tick exposure.
§Known history of vaccination against yellow fever virus, Japanese encephalitis virus, or tick-borne encephalitis virus.
¶Cross-reactivity on POWV IgG IFA assay is consistent with a history of West Nile virus infection.
antibodies are often not present during early infection (12). The 2 patients screened by chemical methods who were positive for POWV IgG failed to show neutralization by PRNT; however, rather than PRNT 90, we used POWV PRNT 99, which has greater specificity but lower sensitivity. In addition, our PRNT was based on POWV lineage I; thus, our test was potentially less sensitive at detecting POWV lineage II–specific antibodies and thus less capable of detecting previous POWV lineage II infection.

Similar to other flavivirus serologic assays, considerable cross-reactivity occurred with the Flavivirus Mosaic IgG IFA assay (online Technical Appendix Table) (13). The fluorescence intensity was stronger for TBEV than it was for other flaviviruses in all TBD patients except for 1 patient with prior confirmed WNV infection. Both patients with routine chemistry screening completed who were POWV IgG–positive were TBEV IgM–positive. Neither had a history of yellow fever or dengue virus exposure or vaccination, although the panel showed cross-reactivity with these viruses.

Evidence of current or prior B. burgdorferi infection was present in 63 (66.3%) TBD patients and 4 (8%) patients with routine chemistry screening completed (p<0.0001). Of the 41 (43.2%) TBD patients with evidence of B. burgdorferi infection, 7 (17.1%) had serologic evidence of acute POWV infection and 3 (7.3%) had laboratory-confirmed POWV infection. When controlling for differences in seroprevalence rates of B. burgdorferi, no statistical differences were evident for POWV seroprevalence (p = 1.0) or confirmed infections (p = 1.0) between patients with routine chemistry screening completed and TBD patients, although the study was underpowered in this regard.

B. burgdorferi IgM was detected in 6 (85.7%) of the 7 patients with serologic evidence of acute POWV infection, suggesting concurrent infection, which is consistent with surveillance data indicating that POWV and B. burgdorferi co-infect I. scapularis ticks (2,3). The rate of concurrent antibodies we report is higher than that described for regions of Europe endemic for TBE and Lyme disease (14).

Clinical data were available for 7 of the patients with serologic evidence of POWV infection (Table 2). Infection probably occurred in 3 patients. A laboratory-confirmed nonacute infection was found in a patient (patient no. 2) who did not meet Centers for Disease Control and Prevention criteria. Patient symptoms could not be attributed specifically to POWV because all TBD patients with clinical data available were positive for B. burgdorferi antibodies, and testing for the possibility of infection with additional endemic tick pathogens was performed for only 2 patients.

Consistent with previous studies showing increased susceptibility of children to arboviral diseases, 3 patients who might have had POWV infection were children (Table 2) (15). Fever was present in all patients with evidence of POWV acute infection; other common symptoms were fatigue, malaise, fussiness, listlessness, and headache. Complete blood cell count and C-reactive protein did not indicate severe infection. Consistent with other arboviral diseases, urticarial or maculopapular rash was documented in 3 patients (15). No patients had neuroinvasive disease.

This study had limitations. Similar to other serologic studies, cross-reactivity and prior exposure to POWV cannot be completely excluded in serologically positive cases. Analysis for other flaviviruses, prior yellow fever virus vaccination, and history of travel to dengue-endemic regions, as well as PRNT, were completed to address this concern. The study population was limited to persons in the US upper Midwest, although POWV is likely an increasing problem throughout the territory I. scapularis ticks occupy. Our study results might not be applicable to these other regions.

Conclusions

In a Lyme disease–endemic area, POWV seroreactivity and confirmed POWV infection were present. The spectrum of disease is broader than previously realized, with most patients having minimally symptomatic infection (1,10,11). Further studies are needed to characterize clinical disease of POWV mono-infection, document POWV seroprevalence in humans, and monitor epidemiologic trends.

Acknowledgments

We thank Marshfield Clinic Research Foundation staff for supporting this study and Marshfield Labs staff for collecting specimens. We also thank Diep Johnson for assisting with this study.

Dr. Frost is a pediatrician and physician scientist at Marshfield Clinic in Minocqua, Wisconsin. Her research interests include tickborne pathogens, blastomycosis, and antimicrobial stewardship.

References


3. Knox K, Thomm A, Harrington Y, Baewer D, Carrigan D. Arbovirus co-infections in Wisconsin tick populations. Poster presentation at: IDWeek; October 7–11, 2015; San Diego, CA, USA.


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**etymology**

**influenza**

[in"flo-en'za]

Acute viral infection of the respiratory tract. From Latin *influentia*, "to flow into"; in medieval times, intangible fluid given off by stars was believed to affect humans. The Italian *influenza* referred to any disease outbreak thought to be influenced by stars. In 1743, what Italians called an *influenza di catarro* ("epidemic of catarrh") spread across Europe, and the disease came to be known in English as simply "influenza."


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https://wwwnc.cdc.gov/eid/article/12/1/et-1201_article