Molecular tests revealed influenza D viruses of D/OK lineage widely circulating in farmed animal species in Guangdong Province, southern China. In particular, we found high levels of influenza D virus infection in goats and pigs. We also detected viral RNA in serum specimens and feces of animals with certain severe diseases.

Four types of influenza viruses (A–D) have been confirmed (https://www.cdc.gov/flu/about/viruses/types.htm). The recently discovered influenza D virus is thought to cause respiratory diseases primarily in cattle and to a lesser extent in pigs (1–4). Moreover, serologic evidence for influenza D virus infection in small ruminants and humans has been established (5,6). Since the initial influenza D virus isolation in the United States in 2011 (1), the virus has been reported in China, Mexico, France, Italy, and Japan (7–11). Genetic analysis of the hemagglutinin-esterase-fusion gene demonstrated that these viruses had 2 distinct lineages, represented by D/OK and D/660 (12). Recently, a novel influenza D virus that emerged in Japan has been proposed as the third lineage (11). D/OK lineage–related viruses were previously identified in native Luxi yellow cattle in Shandong Province, northern China (7). Despite good progress in identifying domestic cattle as the primary reservoir of influenza D virus, we know little about prevalence in other animals. We conducted a study to clarify the origin and transmission dynamics of influenza D virus in goats, buffalo, and pigs as well as farmed cattle.

The Study
In 2016, we collected 607 clinical samples from 4 species of animals with different clinical diseases and 250 nasal swab samples from asymptomatic animals (Table) from 16 farms in 4 cities of Guangdong Province: Guangzhou, Qingyuan, Heyuan, and Jiangmen (Figure 1). In addition, we randomly chose 200 archived Holstein dairy cattle serum samples, 40 per year, from 2011–2015 to investigate possible early RNA distribution of influenza D virus in this region. We used the reverse transcription PCR method and subcloning protocol (online Technical Appendix, https://wwwnc.cdc.gov/EID/article/23/8/17-0059-Techapp.pdf). We performed sequence alignment using ClustalW implemented in DNASTar software (DNASTar, Madison, WI, USA), and we conducted phylogenetic analyses based on our obtained sequences and reference truncated sequences (496-bp) of influenza D viruses from GenBank by using MEGA 5.1 software (http://www.megasoftware.net; online Technical Appendix Table).

After testing by reverse transcription PCR with further sequencing confirmation, we found influenza D virus–positive rates in 230 total nasal swab samples of 12.8% (20/156) for dairy cattle, 7.3% (4/55) for native yellow cattle, and 36.8% (7/19) for pigs. Rates in 324 total serum samples were 7.8% (15/193) for dairy cattle, 5.9% (3/51) for buffalo, and 33.8% (27/80) for goats. The influenza D virus–positive rate was also high (28.9%, 13/45) in swine lung samples. In contrast, we found no or low prevalence (<2%) in asymptomatic animals tested (Table). Moreover, all of the archived serum samples were found to be influenza D virus negative. Interestingly, 1 of 8 rectal swabs of goats with severe diarrhea tested positive (Table). Samples from animals with reproductive problems had a positive rate of 4.3% (5/116) (Table).

Sequence alignment analysis showed that the nucleotide sequences of influenza D viruses found in this study shared high similarity (99%–100%) with previously described sequences from China (7) and low similarity (93.8%–98.8%) with sequences originating from the United States, France, Italy, Mexico, and Japan (1,8–12). Similarly, phylogenetic analysis revealed that all influenza D virus sequences in this study clustered together with previous sequences from China and belonged to the D/OK lineage (Figure 2).

Conclusions
When first discovered, influenza D virus was reported in diseased pigs in the United States (1). Later, it was
identified in cattle and swine herds in several other countries, with or without clinical manifestation (7–11). Moreover, antibodies to influenza D virus were detected in goats, sheep, and humans (5–6). Under experimental conditions, influenza D virus replicated and transmitted among ferrets and guinea pigs (13). We confirmed that influenza D virus is widely present in cattle species (dairy cattle, yellow cattle, and buffalo). We also found influenza D virus at a high prevalence (>30%) in pigs and goats (Table), which is in contrast to the low prevalence found in previous investigations (1,5,10). The high prevalence may be caused by poor biosecurity measures and high-density feeding mode practices in China’s animal industry as well as possible cross-species transmission (13). Taken together, our findings expand the host range of influenza D virus and further emphasize the health concern this virus poses to multiple animal species.

Previous studies have shown that influenza D viruses are mainly found in respiratory tract samples (14–17) and that they have played an etiologic role in bovine respiratory diseases (2–4). In this study, we found that influenza D virus RNA was present in cattle and goat serum samples; it was also present in goat rectal swabs, accompanied by peste des petits ruminants virus and caprine kobuvirus (data not shown). The distribution of influenza D virus in our study is not the same as that described under experimental conditions (3).

Influenza viremia, an indicator of disease severity (14), has been detected in 20.9% of severe cases during the acute phase of infection or before host death. Our detection of influenza D virus genome in serum samples from severely diseased animals (Table) implies that the virus could enter transiently into the animal’s circulatory system through capillaries lining the respiratory tract, which

<table>
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<th>Animal species and farm</th>
<th>Farm type†</th>
<th>Farm location</th>
<th>No. animals</th>
<th>Age range of animals</th>
<th>Sample type</th>
<th>No. positive/no. samples</th>
<th>Detection rate, %</th>
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<td>6/70‡</td>
<td>8.57</td>
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<td>5/99‡</td>
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<td>Lung</td>
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<td>5–15 wks</td>
<td>Nasal swab</td>
<td>4/10‡</td>
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<td>Lung</td>
<td>1/8‡</td>
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<td>7/25‡</td>
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<td>1/8†</td>
<td>12.5</td>
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<td>Serum</td>
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<td>4</td>
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<td>0/50§</td>
<td>0</td>
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<tr>
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<td>1–4 y</td>
<td>Nasal swab</td>
<td>0/50§</td>
<td>0</td>
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<td>Qingyuan: Yingde</td>
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<td>1–4 y</td>
<td>Nasal swab</td>
<td>0/50§</td>
<td>0</td>
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</table>
further contributes to the possibility of detecting virus in other organs. Similar to previous studies (2,4), we also found that the reverse transcription PCR positive rate was significantly higher (4%–40%) in diseased animals than the rate (≤2%) observed in asymptomatic animals (p<0.05), which suggests a potential correlation between the disease severity and presence of influenza D virus. For influenza D virus found in rectal swabs, it might be that animals have swallowed the virus. Another possibility is that, similar to influenza A and B viruses, influenza D virus can replicate within the intestinal tract (15).

We detected influenza D virus in cattle with reproductive disorders. However, we could not determine whether influenza D virus is associated with reproductive problems. Future studies can be designed to investigate these scientific issues.

To date, 2 lineages of influenza D virus (D/OK and D/660) co-circulate in North America and Europe (8–10,12). However, only the D/OK lineage has been found in China, and a potential third lineage was found in Japan (7,11). Our study confirms and further extends the previous observation that D/OK lineage circulates in East Asia. The viral, host, and ecologic factors that shape the observed contrasting phylodynamics of influenza D viruses among different geographic regions warrant further investigation.

In addition, we found different minor genetic variants circulating on the same farm (Figure 2), indicating the ongoing evolution of influenza D viruses in their hosts (7,8,11). In comparing our sequences to the reference sequences from different animal species, we found 4 frequent nucleotide mutations (at positions 136, 231, 263, and 486) (online Technical Appendix Figure 1), which caused 2 amino acid mutations at positions 77 and 88 (online Technical Appendix Figure 2). Interestingly, among 4 nucleotide mutations, 1 unique nucleotide (T at position 486) was originally from the D/660 lineage. Moreover, we found several consistent sequences co-circulating in multiple animal species (online Technical Appendix Figure 1). Our speculation is that homologous recombination among different influenza D viruses and potential cross-species transmission under field conditions are possible, but further study is needed.

In summary, our study investigating the infection status of influenza D virus in different farmed animal species in Guangdong Province provides novel insights into the epidemiology and evolution of this virus. In particular, we document the molecular evidence for influenza D virus infection in goats and buffalo.

**Acknowledgments**

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Influenza D Virus Animals, Guangdong Province, China

People’s Republic of China (Grant no. 2015GA780010). The work was also supported in part by SDSU AES Fund 3AH-477 to F.L. and D.W. S.-L.Z. is sponsored by Guangdong Academy of Agricultural Sciences, Guangzhou, China.

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References


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The work of art shown here depicts the interrelationship of human, animal, and environmental health.

Stained-glass windows have been appreciated for their utility and splendor for more than 1,000 years, and this engaging work of art by stained glass artist Jenny Hammond reminds us that influenza A viruses—which can be easily spread between animals and humans, use various host species, and exist in many different environments—remain an enduring and global health concern.

Visit our website to listen: https://www2c.cdc.gov/podcasts/player.asp?f=8644950
Influenza D Virus in Animal Species in Guangdong Province, Southern China

Technical Appendix

RT-PCR method and subcloning protocol used in this study

A classic reverse transcription PCR (RT-PCR) method was developed using a pair of primers (HEF-F: 5′-AAC CRC ATC TTC TTG TTC TTC A-3′ and HEF-R: 5′-TGC TTC TTC WGT GGC ATT ATC T-3′) targeting at the partial hemagglutinin-esterase-fusion (HEF) gene (at positions 582–1077) to test the presence of IDVs and further define IDV genetic lineages in those samples. For RT-PCR, the primers were diluted to 10 μM with ddH₂O.

A 25 μL RT-PCR system was constructed using a one-step RT-PCR kit (Takara Bio Inc.), which contained 0.5 μL of each primer, 0.5 μL of enzyme mixture (including PrimeScript RTase, DNA polymerase, RNase inhibitor), 12.5 μL of 2 × buffer, 8.5 μL of ddH₂O, and 2.5 μL of viral RNA. RT-PCR was performed as follows: reverse transcription (RT) at 50°C for 30 min, 95°C for 5 min, followed by 35 PCR cycles (95°C for 30 sec, 55°C for 30 sec, 72°C for 30 sec), and a final extension at 72°C for 10 min. The positive PCR fragments (~500 bp) were purified and cloned into the pGEM–T vector (TIANGEN, Inc., Beijing). Positive recombinant plasmids were purified according to the manufacturer’s instructions (TIANGEN, Inc., Beijing) and sequenced using the Sanger sequencing method (Sangon Biotech Co., Ltd., Shanghai). RT-PCR detection data were analyzed by chi-square test (Ziyue software), and significance was set at p < 0.05.
**Technical Appendix Table.** Reference influenza D virus isolates with partial hemagglutinin-esterase-fusion sequences used in this study

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Technical Appendix Figure 2. Multiple sequence alignment results of the current IDV amino acid sequences (165 aa) and corresponding published reference sequences.