

in the treatment of associated infections (8). Prolonged exposure to microbiocides, including chlorhexidine, has been shown to result in a stable increase in the expression of antibiotic-resistance mechanisms (1,6), and elevated chlorhexidine resistance has been reported in multidrug-resistant strains of *B. cenocepacia* from cystic fibrosis patients (9). Three resistance-nodulation-division (RND) efflux pump genes (RND3, RND4, and RND9) have been shown to be essential for chlorhexidine tolerance in *B. cenocepacia* (9). Examination of the complete genome of *B. lata* isolate A05 revealed the presence of RND3, RND4, and RND9 in each strain ( $\geq 94\%$  sequence identity) (online Technical Appendix Figure 2).

*B. contaminans* is the cause of widespread pharmaceutical product contamination, and infection outbreaks by this species are well-documented (3,10). Our findings suggest that the other member of Bcc group K, *B. lata*, also represents an important opportunistic pathogen of relevance to infection control, particularly given its intrinsic biocide tolerance.

### Acknowledgments

The authors gratefully acknowledge Deborah Williamson and Mark Schultz for their helpful discussions, Trang Nguyen for providing the New South Wales isolates, and microbiological laboratory staff from South Australia Pathology, Pathology North of New South Wales, and Pathology West of New South Wales for their technical input.

### About the Author

Dr. Leong is a research scientist at the South Australian Health and Medical Research Institute. His research interests include pathogen genomic epidemiology and infection outbreaks.

### References

- Rushton L, Sass A, Baldwin A, Dowson CG, Donoghue D, Mahenthalingam E. Key role for efflux in the preservative susceptibility and adaptive resistance of *Burkholderia cepacia* complex bacteria. *Antimicrob Agents Chemother*. 2013;57:2972–80. <http://dx.doi.org/10.1128/AAC.00140-13>
- Pope CE, Short P, Carter PE. Species distribution of *Burkholderia cepacia* complex isolates in cystic fibrosis and non-cystic fibrosis patients in New Zealand. *J Cyst Fibros*. 2010;9:442–6. <http://dx.doi.org/10.1016/j.jcf.2010.08.011>
- Medina-Pascual MJ, Valdezate S, Carrasco G, Villalón P, Garrido N, Saéz-Nieto JA. Increase in isolation of *Burkholderia contaminans* from Spanish patients with cystic fibrosis. *Clin Microbiol Infect*. 2015;21:150–6. <http://dx.doi.org/10.1016/j.cmi.2014.07.014>
- Martina P, Bettiol M, Vescina C, Montanaro P, Mannino MC, Prieto CI, et al. Genetic diversity of *Burkholderia contaminans* isolates from cystic fibrosis patients in Argentina. *J Clin Microbiol*. 2013;51:339–44. <http://dx.doi.org/10.1128/JCM.02500-12>
- Oie S, Kamiya A. Microbial contamination of antiseptics and disinfectants. *Am J Infect Control*. 1996;24:389–95. [http://dx.doi.org/10.1016/S0196-6553\(96\)90027-9](http://dx.doi.org/10.1016/S0196-6553(96)90027-9)
- Knapp L, Rushton L, Stapleton H, Sass A, Stewart S, Amezcuita A, et al. The effect of cationic microbicide exposure against *Burkholderia cepacia* complex (Bcc): the use of *Burkholderia lata* strain 383 as a model bacterium. *J Appl Microbiol*. 2013;115:1117–26. <http://dx.doi.org/10.1111/jam.12320>
- Drevinek P, Holden MT, Ge Z, Jones AM, Ketchell I, Gill RT, et al. Gene expression changes linked to antimicrobial resistance, oxidative stress, iron depletion and retained motility are observed when *Burkholderia cenocepacia* grows in cystic fibrosis sputum. *BMC Infect Dis*. 2008;8:121. <http://dx.doi.org/10.1186/1471-2334-8-121>
- George AM, Jones PM, Middleton PG. Cystic fibrosis infections: treatment strategies and prospects. *FEMS Microbiol Lett*. 2009;300:153–64. <http://dx.doi.org/10.1111/j.1574-6968.2009.01704.x>
- Coenye T, Van Acker H, Peeters E, Sass A, Buroni S, Riccardi G, et al. Molecular mechanisms of chlorhexidine tolerance in *Burkholderia cenocepacia* biofilms. *Antimicrob Agents Chemother*. 2011;55:1912–9. <http://dx.doi.org/10.1128/AAC.01571-10>
- Martin M, Christiansen B, Caspari G, Hogardt M, von Thomsen AJ, Ott E, et al. Hospital-wide outbreak of *Burkholderia contaminans* caused by prefabricated moist washcloths. *J Hosp Infect*. 2011;77:267–70. <http://dx.doi.org/10.1016/j.jhin.2010.10.004>

Address for correspondence: Lex Leong, South Australian Health and Medical Research Institute, North Terrace, Adelaide, South Australia 5000, Australia; email: lex.leong@sahmri.com

## Estimating Latent Tuberculosis Infection Using Interferon- $\gamma$ Release Assay, Japan

Tomoyasu Nishimura, Masaki Ota, Masaaki Mori, Naoki Hasegawa, Hiroshi Kawabe, Seiya Kato

Author affiliations: Keio University, Tokyo, Japan (T. Nishimura, M. Mori, H. Kawabe); Research Institute of Tuberculosis—Japan Anti-Tuberculosis Association, Tokyo (M. Ota, S. Kato); Keio University School of Medicine, Tokyo (N. Hasegawa)

DOI: <https://doi.org/10.3201/eid2411.171948>

We estimated the latent tuberculosis infection (LTBI) rate for foreign-born students at Keio University, Tokyo, Japan, using an interferon- $\gamma$  release assay. The LTBI rate for students from countries with estimated tuberculosis incidence  $>100$  cases/100,000 persons was high (10.0%). Universities should screen for LTBI in students from countries with high tuberculosis incidence.

The proportion of foreign-born tuberculosis (TB) patients among all TB patients in Japan is increasing, particularly for those 20–29 years of age (57.7% in 2016) (1). The Tokyo metropolitan government revealed a foreign-born student-related TB outbreak at a Japanese language school in 2016 (2). TB outbreaks involving foreign-born students create concerns that TB infection from such students, particularly those from countries with a high incidence of TB, might spread to the population of Japan.

In Japan, university students, including foreign-born students, undergo TB screening with chest radiograph; however, a chest radiograph cannot detect LTBI; it detects only pulmonary TB. Because immigrants may develop TB after entry (3), screening with chest radiograph might be ineffective; therefore, screening for LTBI may be necessary to prevent TB outbreaks. However, only a few surveys of TB infection among foreign-born persons have been conducted in Japan (4). We conducted a survey of LTBI among foreign-born students by using an interferon- $\gamma$  release assay (IGRA).

Keio University has 6 campuses in the greater Tokyo area comprising  $\approx$ 33,000 students, of whom  $\approx$ 1,600 are foreign-born from 74 countries. During September 2016–September 2017, we recruited foreign-born students  $\geq$ 20 years of age studying at Keio University who had no history of mycobacterial diseases or HIV infection. After obtaining informed consent, we collected whole blood specimens for the T-SPOT.TB test (Oxford Immunotec Ltd., Abingdon, UK), an IGRA available in Japan. All participants were screened for pulmonary TB with chest radiograph. We interviewed participants using a structured questionnaire on identification and demographic information, the date of first arrival in Japan, and history of TB. We derived country-specific estimated TB incidence rates from the World Health Organization website (5). Statistical results were computed by using R software (The R Foundation, Vienna, Austria). This study was conducted in compliance with the Declaration of Helsinki and approved by the

institutional ethics review committee for human research of the Keio University School of Medicine and Hospital (no. 20160080).

We enrolled 177 participants 20–42 years of age (median 23 years), of whom 98 (55.1%) were female (Table). Participants were from China (55 students), Indonesia (24 students), France (19 students), Germany (9 students), and Thailand (8 students); the remaining participants were from 28 different countries, including 50 from countries with estimated TB incidence rates  $>$ 100 cases/100,000 persons. We excluded data for 1 participant with an indeterminate IGRA result. A total of 117 (66.1%) students participated in this study within 1 month after arriving in Japan.

Overall, 8 (4.5% [95% CI 2.0%–8.7%]) students tested positive on IGRA (2 each from China and Thailand and 1 each from Ghana, Indonesia, South Korea, and the Philippines). The rate of the positive IGRA result for students from countries with an estimated TB incidence rate of  $>$ 100 cases/100,000 persons was 10.0% (95% CI 3.3%–21.9%) and relative risk was 4.2 (95% CI 1.1–17.1), whereas the rate for students from countries with an estimated TB incidence rate of  $<$ 100 cases/100,000 persons was 2.4% (95% CI 0.49%–6.7%). Even IGRA positivity of students 20–29 years of age from countries with estimated TB incidence rates of  $>$ 100 cases/100,000 persons was 9.4% (95% CI 2.0%–25.0%). Chest radiograph found no students with pulmonary TB. We recommended that all IGRA-positive students receive LTBI treatment and close follow-up to detect the development of TB as early as possible.

The overall rate of LTBI among foreign-born students at Keio University was 4.5%. This rate was significantly higher for these students than for Keio University students from Japan assessed during 2009–2013 (0.73% [95% CI 0.39%–1.2%]; T. Nishimura et al., unpub. data). Our findings are consistent with those of previous studies. Ogiwara et al. showed that 7.8% of study participants tested positive for LTBI using the QuantiFERON-TB Gold test on 384 foreign-born students, of whom 363 were from countries with high TB incidence rates (4).

**Table.** Characteristics and IGRA results of participants in a study of latent TB infection, Japan, 2016–2017\*

Characteristic	IGRA positive, no. (%), 95% CI, n = 8	IGRA negative, no., n = 169	p value†
Sex			
F	5 (5.1, 1.7–11.5)	93	0.733
M	3 (3.8, 0.79–10.7)	76	
Age, y			
20–29	6 (3.9, 1.4–8.3)	148	0.278
$\geq$ 30	2 (8.7, 1.1–28.0)	21	
TB incidence rate in country of origin			
$<$ 100 cases/100,000 population	3 (2.4, 0.49–6.7)	124	0.042‡
$>$ 100 cases/100,000 population	5 (10.0, 3.3–21.8)	45	
Time living in Japan, y			
$<$ 1	5 (3.4, 1.1–7.9)	140	0.158
$\geq$ 1	3 (9.4, 2.0–25.0)	29	

\*IGRA, interferon- $\gamma$  release assay; TB, tuberculosis.

†Differences between positive and negative groups were tested using Fisher exact test.

‡ $p <$  0.05.

Our study had a few strengths and limitations. The number of study participants was large enough for us to stratify the participants by estimated TB incidence rates for their countries of origin. One limitation was that the participation rate was small. Just  $\approx 11\%$  of foreign-born students at Keio University participated; therefore, the results obtained might not be representative of LTBI in all foreign-born students.

In conclusion, we found that estimated LTBI rates for foreign-born students in Japan from countries with high TB incidence rates were higher than those for students from countries with low TB incidence rates and for students from Japan. Based on our findings, we recommend that universities screen for LTBI using IGRAs in students from countries with high TB incidence rates (i.e.,  $>100$  cases/100,000 persons).

### Acknowledgments

We are grateful to Satoshi Mitarai for his critical reading of our manuscript.

This research was supported by the Research Program on Emerging and Re-emerging Infectious Diseases from the Japan Agency for Medical Research and Development (JP17fk0108304) and Keio Gijuku Academic Development Funds. This research study was completed as part of our collaborative research with Oxford Immunotec, Ltd. (Abingdon, UK).

### About the Author

Dr. Nishimura is a physician working as an assistant professor at Keio University. His primary research interest is mycobacterial diseases.

### References

1. Kekkaku Yobo kai (Japan Anti-Tuberculosis Association). Kekkaku no toukei 2017 (Statistics of TB 2017) [in Japanese]. Tokyo: Kekkaku Yobo kai (Japan Anti-Tuberculosis Association); 2017.
2. The Japan Times. Tokyo reveals rare outbreak of tuberculosis, plays down ongoing risk [cited 2017 Nov 23]. <https://www.japantimes.co.jp/news/2016/05/18/national/tokyo-reveals-rare-outbreak-of-tuberculosis-plays-down-ongoing-risk/>
3. Public Health England. Non-UK born TB cases. In: Tuberculosis in England. 2015 Report (presenting data to end of 2014) version 1.1 [cited 2017 Nov 20]. [https://www.gov.uk/government/uploads/system/uploads/attachment\\_data/file/564649/TB\\_annual\\_report\\_2015.pdf](https://www.gov.uk/government/uploads/system/uploads/attachment_data/file/564649/TB_annual_report_2015.pdf)
4. Ogiwara T, Kimura T, Tokue Y, Watanabe R, Nara M, Obuchi T, et al. Tuberculosis screening using a T-cell interferon- $\gamma$  release assay in Japanese medical students and non-Japanese international students. *Tohoku J Exp Med*. 2013;230:87–91. <http://dx.doi.org/10.1620/tjem.230.87>
5. World Health Organization. Tuberculosis country profiles [cited 2017 Nov 21]. <http://who.int/tb/country/data/profiles/en/>

Address for correspondence: Tomoyasu Nishimura, Health Center, Keio University, 35 Shinanomachi, Shinjuku-ku, Tokyo 160-8582, Japan; email: [tnishimura@keio.jp](mailto:tnishimura@keio.jp)

## Effect of Inactivated Poliovirus Vaccine Campaigns, Pakistan, 2014–2017

Nicholas C. Grassly, Mufti Zubair Wadood, Rana M. Safdar, Abdirahman Sheikh Mahamud, Roland W. Sutter

Author affiliations: Imperial College London, London, United Kingdom (N.C. Grassly); World Health Organization, Geneva, Switzerland (M.Z. Wadood, R.W. Sutter); Ministry of National Health Services, Islamabad, Pakistan (R.M. Safdar); World Health Organization, Islamabad (A.S. Mahamud)

DOI: <https://doi.org/10.3201/eid2411.180050>

Pakistan began using inactivated poliovirus vaccine alongside oral vaccine in mass campaigns to accelerate eradication of wild-type poliovirus in 2014. Using case-based and environmental surveillance data for January 2014–October 2017, we found that these campaigns reduced wild-type poliovirus detection more than campaigns that used only oral vaccine.

Routine immunization with  $\geq 1$  dose of inactivated poliovirus vaccine (IPV) in all countries using oral poliovirus vaccine (OPV) was recommended by the World Health Organization (WHO) in November 2012, before the global withdrawal of the serotype 2 component from OPV (1). IPV has also been used since 2014 in mass campaigns to help interrupt wild poliovirus transmission and stop serotype 2 vaccine-derived poliovirus (VDPV2) outbreaks. The IPV supply was severely constrained during 2016–2017; only 2 manufacturers supply the United Nations Children's Fund, and their failure to produce the expected bulk product has meant that only about half the awarded quantities were supplied (2). As a result of these unplanned reductions in IPV supply, countries have delayed the introduction of IPV to routine immunization or faced stockouts, and mass campaigns with IPV in response to VDPV2 are no longer recommended by WHO (3). Nonetheless, where possible, IPV continues to be used in mass campaigns for outbreak response; for example, Pakistan, Afghanistan, Nigeria, and Syria all used IPV in mass campaigns in 2017.

Given that IPV supply constraints are likely to continue until at least the end of 2018, it is crucial that available IPV be optimally allocated between routine immunization and mass campaigns. We recently published estimates of the impact of OPV mass campaigns with and without the inclusion of IPV in Nigeria and Pakistan during January 2014–April 2016 (4). These estimates demonstrated