In-Host Adaptation of Salmonella enterica Serotype Dublin during Prosthetic Hip Joint Infection

Technical Appendix

High-Throughput Genome Sequencing

High-throughput whole-genome sequencing was conducted on strains at the Plateforme de Microbiologie Mutualisée from the Pasteur International Bioresources network (Institut Pasteur, Paris, France). DNA extraction was carried out by using the MagNAPure 96 system (Roche). The libraries were prepared using the Nextera XT kit (Illumina) and the sequencing was done with the NextSeq 500 system (Illumina) generating 100–146-bp paired-end reads, yielding a mean of 61-fold coverage (minimum 32-fold, maximum 101-fold). Reads were trimmed and filtered using AlienTrimmer (1) with a quality Phred score threshold of 28 on a minimum length of 70 nt. De novo assembly was performed with the SPAdes V.3.6.0 assembler (2). For each isolate, the paired-end reads were aligned against 2 Salmonella enterica serotype Dublin CT_020221853 and 3246 reference genomes, to increase the single-nucleotide polymorphism (SNP) detection (GenBank accession nos. CP001144.1 and CM001151.1, respectively) using Bowtie2 with default parameters (3). SAMtools (4) were then used to build a genome index and identify SNPs from the Bowtie alignments. Several criteria were used to filter resulting SNPs: a minimum coverage (number of reads mapped to the reference genome) of 20 and a minimum quality score of each SNP at 25. For each approach, the resulting SNPs were concatenated to generate a filtered multiple alignment. The resulting sequences were further filtered to remove all SNPs present in insertion sequences identified by ISfinder (5). Other repetitive regions were identified by a self-self-BLAST analysis (6) of the reference sequence, using the following parameters: megablast (word size 28), identity percentage >95% and match length >400 bp. Finally, clusters of SNPs introduced by horizontal sequence transfer were detected and removed with Gubbins (7). Alignment was used as input for the construction of a phylogenetic tree using

MEGA6 using a maximum-likelihood approach (8). MEGA6 was ran using the general time reversible model and a Gamma distribution to model site-specific rate variation (i.e., the GTR+ Γ substitution model). Hundred bootstrap replicate analyses were performed to assess maximum-likelihood phylogeny. The final tree was visualized in FigTree version 1.4.2 (http://tree.bio.ed.ac.uk/software/figtree/).

Availability of data

Genomics sequence assemblies used in this project are available online on the NCBI network under accession PRJNA433355 (available at http://www.ncbi.nlm.nih.gov/bioproject/433355).

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Left hip Prosthetic joint infection (PJI) History of febrile diarrhea due to dubious meat consumption						Night fever Right hip infl	ammation
Left hip prosthetic implant	S. Dublin from: -Blood culture -Left hip joint aspiration -Intraoperative periprosthetic tissue (Str. 2011) DAIR ^a	S. Dublin from: -Blood culture -Right hip intraoper periprosthetic tiss (Str. 2014) DAIR ^a	erative ue				S. Dublin from: -Right hip biopsy ^c -Right hip intraoperative periprosthetic tissue (Str. 2015) One-stage right hip prosthesis exchange
Right hip prosthetic implant	6 weeks intravenous antibiotherapy	2 weeks intraveno	ous antibiotherapy	10 weeks ora	l antibiotherapy		6 weeks intravenous antibiotherapy
	Cefotaxime 4g three times a day Ciprofloxacin 400mg three times a day	5 days: Daptomycin 10mg/kg once a day PTZ ^b 4g three times a day	12 days: Amoxicillin 2g three times a day Ofloxacin 200mg thrice a day	4 weeks: Amoxicillin 2g three times a day Ofloxacin 200mg three times a day	4 weeks: Ciprofloxacin 750 mg twice a day	2 weeks: Amoxicillin 1g three times a day	Ceftriaxone 2g once a day Ciprofloxacin 500mg twice a day
1998 2001 2011 2014 2015							
^a Debridment, antibiotics, irrigation and retention; ^b Piperacillin/tazobactam; ^c negative left hip biopsy culture							

Technical Appendix Figure. Clinical case summary for immunocompetent patient who acquired Salmonella Dublin infection after receiving a prosthetic hip joint.