Amebaborne Attilina massiliensis Keratitis, France

Alexandre Battaini, Bernard La Scola, Gaëlle Ho Wang Yin, Louis Hoffart, Michel Drancourt

Author affiliations: Aix Marseille Université, Marseille, France (A. Battaini, B. La Scola, M. Drancourt); Aix-Marseille University–APHM, Hôpital de la Timone, Marseille (G.H.W. Yin, L. Hoffart)

DOI: https://doi.org/10.3201/eid2402.170541

We report a case of Acanthamoeba castellani keratitis in a person who wore contact lenses. The amebae hosted an ameba-resistant bacterial symbiont, provisionally named "Attilina massiliensis," a yet undescribed α-Proteobacterium.

Amebal keratitis is an aggressive ocular infection that can lead to blindness (1). It is usually associated with wearing soft contact lenses; Dart et al. documented that in countries with a high prevalence of contact lens wear, 85%–88% of Acanthamoeba keratitis cases occurred in contact lens users (1). These amebae host ameba-resistant bacteria, and increase their pathogenicity to the host (2). Ameba hosting intra-amebal microorganisms have been rarely documented in cases originating in contaminated contact lenses (3) and never in mixed keratitis. We report a case of mixed ameba–amebal-resistant bacterial keratitis.

A 17-year-old woman who wore contact lenses consulted the ophthalmology department of the clinic associated with Hôpital de la Timone, Marseille, France, in July 2016, after experiencing 1 month of keratoconjunctivitis symptoms related to an undocumented clinical diagnosis of herpes virus keratitis of the left eye. The patient had been prescribed a 1-week treatment with valacyclovir (3×/d) and a corneal dressing. Examination of the left eye showed 4/10 visual acuity; the right eye was normal. Slit-lamp examination showed a central radial keratoneuritis, central corneal edema, central diffuse infiltrate, and a punctate superficial keratitis with no predesmecetic precipitates and no satellite lesions (Figure).

The patient was admitted to the hospital and was administered hourly topical treatments of polyhexamethylene biguanide eye drops, hexamidine, and 1% atropine. The patient, whose diagnosis was early-stage Acanthamoeba keratitis infection, was discharged after 5 days of treatment; a corneal swab sample at discharge was negative for herpes virus, varicella zoster virus, adenovirus, enterovirus, cytomegalovirus, and Chlamydia trachomatis. Follow-up 7 days later yielded reduced symptoms. We followed up on the patient biweekly and slowly tapered drugs over 4 months; the previously negative

References


Address for correspondence: Janet Foley, University of California, Department of Medicine and Epidemiology, 1320 Tupper Hall, Davis, CA 95616, USA; email: jefoley@ucdavis.edu

This project was funded by the One Health Institute and the Wildlife Health Center of University of California, Davis.

About the Author

Miss Vogel is a veterinary student at University of California Davis School of Veterinary Medicine and works as a laboratory intern in J.F.’s laboratory. Her research interest is the ecology of emerging vectorborne diseases.
pathogen tests remained negative. However, culture yielded *Corynebacterium ureicelerivorans*, identified on the basis of a 98.7% partial rpoB gene sequence similarity with the reference sequence (GenBank accession no. FJ392018.1), *Acanthamoeba castellani*, identified on the basis of a 99% 18S rRNA gene sequence similarity with reference genotype T4 (GenBank accession no. U07416.1). Further culture of the amebal isolate in sterile peptone-yeast-extract-glucose broth (Culture-Top, Courtaboeuf, France) by using both optic microscopy and electron microscopy (Figure) yielded an intra-amebal *Holosporaceae* bacterium observed in the cytoplasm of the ameba. This symbiont, a yet undescribed α-Proteobacterium of the family of *Holosporaceae* that had been provisionally named “Attilina massiliensis,” was identified on the basis of a 100% 16S rRNA gene similarity with the reference sample (GenBank HM138368). After subculture, this “*A. massiliensis*” isolate was shown to be flagellated and highly mobile. Moreover, it was shown to lyse the *A. castellani* Neff strain (ATCC 30010), an observation suggesting motility-linked pathogenicity. Culturing the lens storage case yielded *Serratia liquefaciens* and *Stenotrophomonas maltophilia*, identified by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (4).

In this patient, mixed *C. ureicelerivorans*, *A. castellani*, and “*A. massiliensis*” keratitis was firmly documented by the isolation and culture of the pathogens identified by using appropriate controls and validated protocols. The fact that we had never documented such an infection in our laboratory excludes mere false-positive results caused by contamination. *C. ureicelerivorans* has mainly been reported to cause septicemia, not ophthalmologic infections (5,6). Its sources and potential relationships with waterborne amebae are unknown. *Acanthamoeba* spp. amebae cause severe keratitis, which may cause visual loss (1). *Acanthamoeba* spp. amebae are ubiquitous in tap water (7). Tap water could be a source of contamination of contact lenses through the wearer’s handwashing habit before lens manipulation (8). Wide varieties of amebae have been documented in contaminated contact lenses, eventually leading to amebal keratitis outbreaks (9).

Culturing an *A. castellani* ameba isolated from a diseased cornea yielded “*A. massiliensis*,” which we isolated once 8 years ago from an *Acanthamoeba polyphaga* ameba collected from a contact lens storage case that belonged to a patient unrelated to the case-patient we report here. The potential for this emerging ameba-resistant bacterium to cause keratitis remains to be analyzed, but we observed that this mobile symbiont lysed the reference amebal strain, demonstrating its cytopathogenicity. Also, ameba-resistant organisms do comprise acknowledged opportunistic pathogens (2), and corneal toxicity was previously reported as significantly higher for *Acanthamoeba*-hosting endosymbionts (10). This investigation illustrates that amebae present in cases of keratitis may shelter organisms that should be provisionally regarded as potential opportunistic pathogens under these circumstances.

This study was supported by IHU Méditerranée Infection, Marseille, France.
About the Author
Dr. Battaini is a laboratory resident at Aix Marseille University, Marseille, France, with particular interest in the prevention of corneal infection in contact lens wearers.

References

Address for correspondence: Michel Drancourt, URMITE, UMR CNRS 7278, IRD 198, INSERM U1095 IHU Méditerranée Infection, 19-21 Bd Jean Moulin 13005 Marseille, France; email: michel.drancourt@univ-amu.fr

Influenza D Virus in Cattle, Ireland

Orla Flynn, Clare Gallagher, Jean Mooney, Claire Irvine, Mariette Ducatez, Ben Hause, Guy McGrath, Eoin Ryan

Author affiliations: Department of Agriculture, Food, and Marine Laboratory Services, Celbridge, Ireland (O. Flynn, C. Gallagher, J. Mooney, C. Irvine, E. Ryan); École Nationale Vétérinaire de Toulouse, Toulouse, France (M. Ducatez); Kansas State University College of Veterinary Medicine, Manhattan, Kansas, USA (B. Hause); University College Dublin, Dublin, Ireland (G. McGrath)

DOI: https://doi.org/10.3201/eid2402.170759

We detected influenza D virus in 18 nasal swab samples from cattle in Ireland that were clinically diagnosed with respiratory disease. Specimens were obtained from archived samples received for routine diagnosis during 2014–2016. Sequencing showed that virus from Ireland clustered with virus sequences obtained in Europe within the D/swine/OK/1334/2011 clade.

Influenza D virus is a recently characterized member of the family Orthomyxoviridae. This virus was originally detected in pigs in the United States (1); however, cattle are now believed to be the main reservoir species (2). Evidence suggests that this virus plays a role in bovine respiratory disease, although experimentally, it caused only mild disease by itself (3). Influenza D virus has been found to be associated with respiratory disease in feedlot cattle (4).

The zoonotic potential of influenza D virus remains unclear; this virus can replicate in ferrets (a model for human influenza infection), and a seroprevalence of 91% was found in persons working closely with cattle (5). However, a study of 3,300 human respiratory samples from Scotland did not detect any influenza D virus–positive samples (6). This virus has been detected in bovine samples in several other countries, including France (7), Italy (8), Japan, and China (9).

Cattle are a major part of the economy in Ireland, where there are ≈7 million (10). To determine whether influenza D virus was present in cattle in Ireland and to investigate epidemiologic factors that might be related to this virus, we conducted a cross-sectional study by using 320 nasal swab specimens from cattle with respiratory disease that were submitted to the Central Veterinary Research Laboratory (Celbridge, Ireland) for routine bovine viral pathogen testing during 2014–2016.

We tested swab specimens by using real-time PCR for influenza D virus as described (11). We selected samples...